

Diagnostic Mycology for Laboratory Professionals

Part Three--Opportunistic Molds

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The presenter states no conflict of interest and has no financial relationship to disclose relevant to the content of this presentation. 1

OUTLINE

- I. Introductory statements
 - A. Review of classification
 - B. Important general criteria
- II. Identification of clinically-significant molds
 - A. Macroscopic morphology
 - B. Microscopic morphology
 - C. Other hints
- III. Antifungal susceptibility testing

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"D#*%it, Jim,
I'm not a physician."

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The Basics

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SCOPE OF FUNGI

- At least 100,000 named fungal species
- ~1 million to 10 million unnamed species; 1000 to 1500 new species per year
- Fewer than 500 named species associated with animal or human disease
- Less than 50 are pathogenic in healthy human hosts

Biol. Rev. 73: 203-266; 1998

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PATHOGENICITY OF FUNGI

-- Generally more chronic than acute	
-- Generally involves predisposition	
Chemotherapy-induced neutropenia	HIV
Organ transplantation	Diabetes
Corticosteroids	Alcoholism
Broad-spectrum antimicrobials	Intravenous drug abuse
Parenteral nutrition	Intensive care population (burns, NICU)
Dialysis	Malignancy
Invasive medical procedures	Other immune deficiency
-- Certain infections can be "signal diseases"	

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CLASSIFYING OPPORTUNISTS

- Taxonomy

Holomorph	
Teleomorph	Anamorph
Sexual reproduction	Asexual reproduction
Fusion of two nuclei into zygote	Mitosis
Perfect Fungi	"Fungi Imperfecti"
<i>Pseudallescheria boydii</i>	<i>Scedosporium apiospermum</i>

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SEXUAL REPRODUCTION

Subphylum Mucoromycotina	Zygothores meet and fuse (zygosporangium) 
Phylum Basidiomycota	  Clamp connections facilitate basidium
Phylum Ascomycota	  Nuclear division inside ascus (bag)
Phylum Deuteromycota	NO SEXUAL REPRODUCTION OBSERVED

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CLASSIFYING OPPORTUNISTS

- Taxonomy
- Cell morphology (conidiogenesis)

Blastic	blastoconidia	annelloconidia
Enlarge, then divide	phialoconidia	poroconidia
Thallic	arthroconidia	aleurioconidia
"Divide", then enlarge		chlamydoconidia
- Mode of entry (implantation; inhalation)

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UNIFYING CONCEPTS

- Macroscopic observation of colonial growth
- Microscopic observation of colonial growth
- Growth on selective medium
- Rate of growth
- Pigmentation



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Wild Card




DERMATOPHYTES

- Infrequent mortality
- Tinea (ringworm)
- Immunocompromised host not required
- Some have niche in terms of parasitism



Geophilic	<i>M. gypseum</i>
Zoophilic	<i>M. canis</i> <i>T. mentagrophytes</i>
Anthropophilic	Most

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DERMATOPHYTES

- Some have regions of endemicity

<i>M. audouinii</i>	Africa, Haiti	
<i>T. violaceum</i>	Middle East, North Africa	
<i>T. concentricum</i>	Polynesia	
	Pockets of C. and S. America	

Dermatophyte	Nails	Skin	Hair
<i>Microsporum</i> spp.	NO	Yes	Yes
<i>Epidermophyton floccosum</i>	Yes	Yes	NO
<i>Trichophyton</i> spp.	Yes	Yes	Yes

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DERMATOPHYTES

Group	Agents	Cultures	%	Total	%
Anthropophilic	<i>T. rubrum</i>	319	(48.7)	501	(76.4)
	<i>T. tonsurans</i>	91	(13.8)		
	<i>T. mentagrophytes</i>	64	(9.7)		
	<i>F. floccosum</i>	37	(4.3)		
Zoophilic	<i>M. canis</i>	137	(20.4)	137	(20.9)
Geophilic	<i>M. gypseum</i>	17	(2.5)	17	(2.5)

Rev. Inst. Med. trop. S. Paulo
45: 259-263; 2003

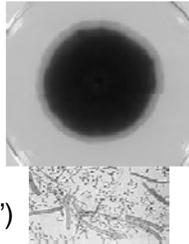
Organisms	Number	Percent
<i>Trichophyton tonsurans</i>	106	56
<i>Epidermophyton floccosum</i>	76	11.8
<i>Trichophyton</i> spp.	27	8.9
<i>T. rubrum</i>	53	6.3
<i>T. violaceum</i>	25	3.9
<i>T. violaceum</i>	21	3.3
<i>M. canis</i>	18	2.5
<i>Microsporum furfur</i>	21	6.3
<i>Phytonospora oval</i>	13	2
Total	641	100

Ann. Trop. Med. Pub. Health
3: 53-57; 2010

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Trichophyton rubrum

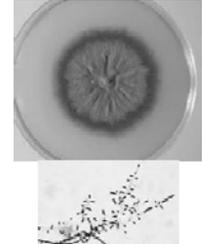
- ~14 days; resistant to cycloheximide
- Diffusible red pigment
- Smooth-walled "pencil" macroconidia (3-8 cells) variable in amount
- Abundant microconidia; tear-shaped ("birds on a wire")
- Urease-negative after 7 days



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Trichophyton tonsurans

- ~12 days; resistant to cycloheximide; scalp
- Suede surface with folds
- Rare, irregular, thick-walled macroconidia
- Abundant microconidia (tears, balloons, clubs); some elongated
- Urease-positive after 4 days



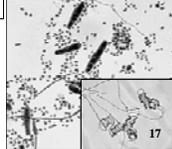
16

Trichophyton mentagrophytes

- ~7-10 days; resistant to cycloheximide; foot

Fluffy, white	Variable-pigment, granular	
Rare macroconidia	Cigar-shaped, smooth, thin-walled (1-6 cells); narrow attachment to hyphae	
Small microconidia; tear-shaped (resembling <i>T. rubrum</i>)	Very round microconidia; clustered on branched conidiophores	

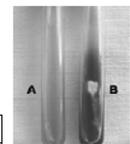
- Spiral hyphae
- Urease-positive after 4 days



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Trichophyton AGARS

- Homogenous suspension of mycelial growth
- Room temperature; 2 weeks



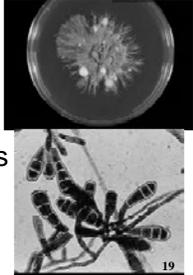
Selected <i>Trichophyton</i> spp.	Growth in Presence of:			
	Casein		Ammonium nitrate	
	Base	+ thiamine	Base	+ histidine
<i>T. rubrum</i>	4+	4+	3+	4+
<i>T. tonsurans</i>	1+	4+	1+	1+
<i>T. mentagrophytes</i>	4+	4+	4+	2+



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Epidermophyton floccosum

- ~10 days; resistant to cycloheximide; jock
- Starts velvety and khaki; becomes fluffy white
- Smooth, thin- or thick-walled macroconidia; rounded ends; single or characteristic clusters
- No microconidia
- Urease-positive after 7 days



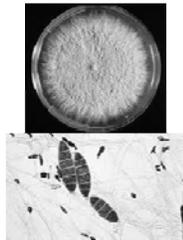
Microsporium canis

- ~6-10 days; resistant to cycloheximide; kids
- Cottony, wooly; lemon periphery closely-spaced grooves
- Rough, thick-walled, spindle-shaped macroconidia; tapers to knob-like ends (6-15 cells)
- Rare, single microconidia
- Urease-positive after 7 days



Microsporium gypseum

- ~6 days; resistant to cycloheximide; kids
- Cinnamon brown to buff; granular (sporulates heavily)
- Very abundant macroconidia; thin-walled with rounded tips (4-6 cells)
- Rare, single microconidia
- Urease-positive after 7 days

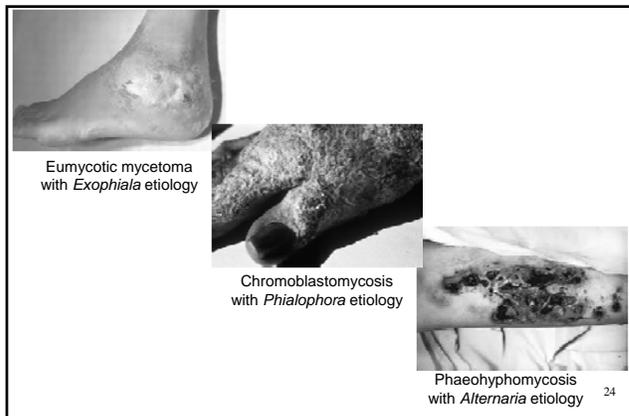


Pictures

DEMATIACEOUS OPPORTUNISTS

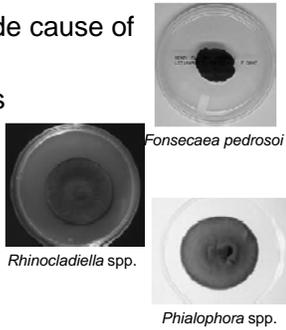
- Soil, plant, moist organics (some air)
- Some tropical; some temperate
- Immunocompromised host not required
- Spectrum of disease

Eumycotic mycetoma
 Chromoblastomycosis
 Phaeohyphomycosis
 Chronic sinusitis (portal for CNS disease)
 Rare systemic disease



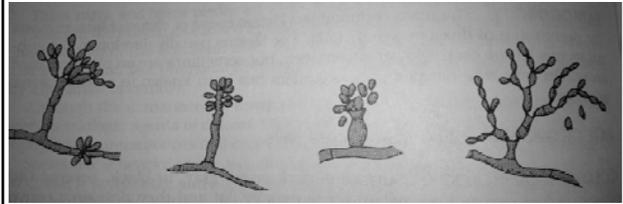
Fonsecaea spp. AND OTHERS

- Most common worldwide cause of chromoblastomycosis
- Maturity in ~14-28 days
- Colony surface dark green, black, or gray; reverse is black
- Conidia (phores), hila, vase-shaped phialides, collarettes, denticles



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Fonsecaea spp. AND OTHERS

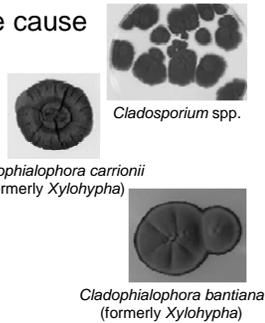


Fonsecaea-type conidiation Rhinocladiella-type conidiation Phialophora-type conidiation Cladosporium-type conidiation

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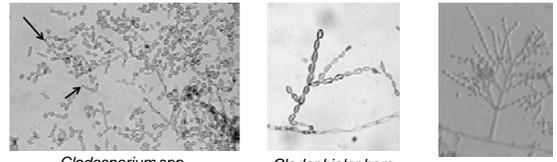
Cladosporium spp., *Cladophialophora*

- Most common worldwide cause of chromoblastomycosis
- Maturity in ~14-28 days
- Colony surface dark green, black, or gray; reverse is black
- Conidia (phores), hila, vase-shaped phialides, collarettes, denticles



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Cladosporium spp., *Cladophialophora*



Cladosporium spp. *Cladophialophora bantiana* *Cladophialophora carrionii*

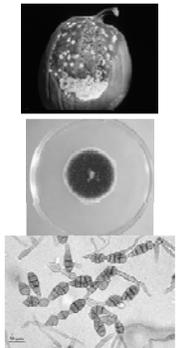
Dematiaceous Mold	Distinct conidiophores	Hila on conidia	Conidial chain length	Conidial chain branching	Gelatin hydrolysis	Growth in 15% NaCl	Max growth ° C
<i>Cladosporium</i> spp.	Yes	Yes	Short	Frequent	Positive	Positive	<37
<i>C. carrionii</i>	Variable	Yes	Medium	Moderate	Negative	Negative	35-37
<i>C. bantiana</i>	No	No	Long	Sparse	Negative	Negative	42-43

D. H. Larone, Medically Important Fungi, fourth ed.

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Alternaria spp.

- Typically contaminant; role in phaeohyphomycosis, allergy
- Maturity in ~5 days
- Colony surface becomes greenish black or brown with light border; reverse is black
- Drumstick macroconidia with longitudinal, transverse septations; poroconidiation (chains)



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Inhalation

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ASPERGILLOSIS

- Nasoorbital
- Endocardial
- Cutaneous
- Disseminated
- Central nervous system disease
- Pulmonary

Allergic bronchopulmonary aspergillosis
Aspergilloma (fungus ball)
Invasive pulmonary aspergillosis

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24 MEDICAL CENTERS; n = 1477

What does your (lab) positive culture result mean???

No. (%) of positive culture results, according to clinical condition

Aspergillus species	Invasive disease (n = 256)	Chronic necrotizing aspergillosis (n = 41)	Aspergilloma (n = 83)	ABPA (n = 87)	Colonization (n = 735)	Contamination (n = 275)
<i>A. flavus</i>	41 (16)	1 (2)	2 (2)	5 (6)	66 (9)	28 (10)
<i>A. fumigatus</i>	171 (67)	33 (80)	57 (69)	80 (92)	485 (63)	102 (37)
<i>A. nidulans</i>	2 (1)	0 (0)	0 (0)	0 (0)	5 (1)	2 (1)
<i>A. niger</i>	14 (5)	4 (10)	11 (13)	0 (0)	101 (14)	66 (24)
<i>A. terreus</i>	8 (3)	0 (0)	0 (0)	0 (0)	8 (1)	1 (1)
Other	2 (1)	3 (8)	4 (5)	0 (0)	28 (4)	17 (6)
Not identified	19 (7)	0 (0)	9 (11)	2 (2)	62 (8)	58 (21)

NOTE: ABPA, allergic bronchopulmonary aspergillosis.

Clin. Infect. Dis. **33**: 1824-1833; 2001

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UNDERLYING RISK AND OUTCOME

Percentage of patients dead at 3 months, according to cause of death

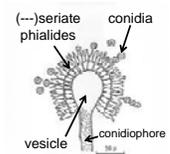
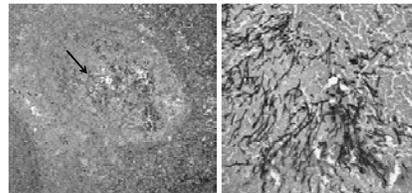
Group, characteristic (no. of patients)	Aspergilloma	Underlying disease	Other/unknown
Disease classification			
Aspergillus colonization (508)	<1	3	9
IA (148)	40	10	12
Risk			
Allogeneic BMT (39)	39	10	8
Autologous BMT (14)	29	7	0
Neutropenia (61)	34	16	13
Hematologic cancer (106)	27	12	11
Solid-organ cancer (124)	5	3	18
Corticosteroid use (381)	11	7	15
Treatment for IA			
Amphotericin B (95)	38	11	11
Itraconazole (43)	21	7	2

NOTE: BMT, bone marrow transplant; IA, invasive aspergillosis.

Clin. Infect. Dis. **33**: 1824-1833; 2001

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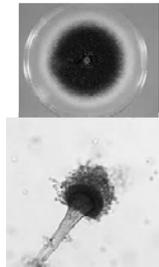
ASPERGILLOSIS



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Aspergillus fumigatus

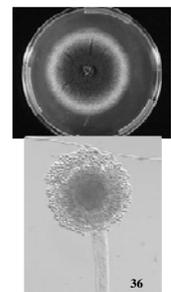
- Maturity in ~3 days
- Conidiophores short & smooth
- Colony surface becomes dark greenish to gray; reverse white to tan
- Uniseriate phialides on upper 2/3 of vesicle; parallel to axis of conidiophore



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Aspergillus flavus

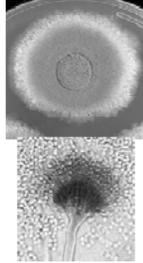
- Commonly associated with aflatoxins
- Conidiophores rough & spiny
- Colony surface velvety, yellow to green or brown; reverse white to tan
- Uniseriate and biseriate phialides covering entire vesicle (all directions)



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Aspergillus terreus

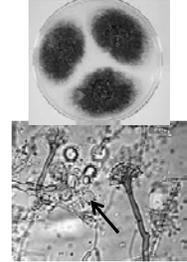
- Commonly considered contaminant
- Conidiophores short & smooth
- Colony surface velvety, cinnamon brown; reverse white to brown
- Biseriate phialides very compact; can be quite lengthy



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Emericella (Aspergillus) nidulans

- Commonly considered contaminant
- Conidiophores short, smooth, brown
- Colony surface typically green (yellow in spots); reverse purplish red
- Biseriate, short, columnar phialides; cleistothecia, Hülle cells



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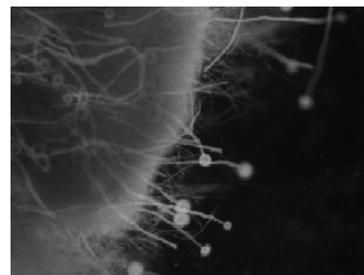
CASE PRESENTATION

- 19-year-old male with three-day history of congestion and ear pain
- PMH of psoriasis in multiple cutaneous sites; daily ibuprofen for tonsillar hypertrophy
- Previous regimens of amoxicillin-clavulanate, amoxicillin, otic neomycin-polymyxin, otic ciprofloxacin-hydrocortisone
- Pain worsened; ENT consult

Courtesy T. K. Block

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CASE PRESENTATION



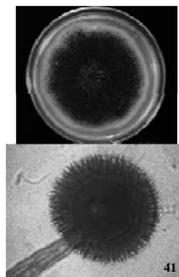
calcofluor white stain;
400x total magnification

Courtesy T. K. Block

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Aspergillus niger

- Can cause disease in debilitated patients
- Conidiophores long & smooth
- Colony surface starts white to yellow, turns black; reverse white to yellow
- Biseriate phialides; forms a "radiate head"



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Aspergillus niger OTOMYCOSIS

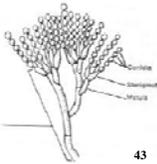
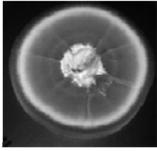
- *A. niger* at least two times more common than *A. flavus* in context of otomycosis
Eur. J. Clin. Microbiol. Infect. Dis. **8**: 413-437; 1989
Am. J. Trop. Med. Hyg. **29**: 620-623; 1980
- Superficial infection; immunocompetent hosts
Eur. J. Clin. Microbiol. Infect. Dis. **8**: 413-437; 1989
- Self-manipulation; manipulation by barbers

Am. J. Trop. Med. Hyg. **29**: 620-623; 1980

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Penicillium spp.

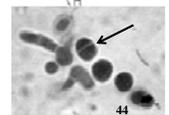
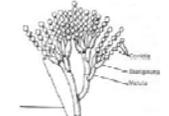
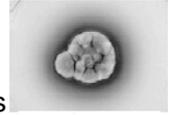
- Typically contaminant; ear, respiratory, cornea, endocarditis
- Maturity in ~4 days
- Colony surface becomes powdery and bluish green with white border; reverse variable
- Branched or non-branched conidiophores; secondary branches known as metulae



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Penicillium marneffeii

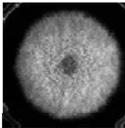
- Endemic to Southeast Asia; compromised and competent
- Mold maturity (25° C) in ~3 days
- Colony surface can become reddish yellow with light edge; reddish pigment diffusion
- Yeastlike cells observed at 35-37° C; central cross wall as result of fission (not budding)



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Fusarium spp.

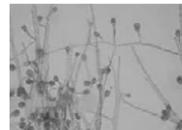
- Common contaminant; mycotic keratitis, disseminated disease
- Maturity in ~4 days
- Cottony surface, develops violet or pink center with light periphery; reverse light
- Canoe-shaped macroconidia ± oval 1- to 2-celled conidia in clusters resembling *Acremonium*



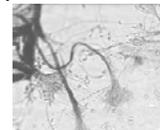
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Pseudallescheria HOLOMORPH

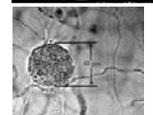
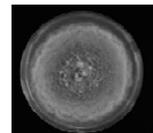
- Mycetoma; respiratory/sinus, disseminates (bone, brain, eyes, meninges)
- Maturity in ~7 days; mouse-like appearance



Scedosporium apiospermum
asexual
no inhibition by cycloheximide



Graphium
asexual
no inhibition by cycloheximide

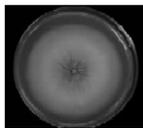


Pseudallescheria boydii
sexual
inhibited by cycloheximide

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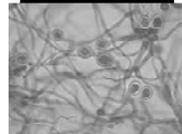
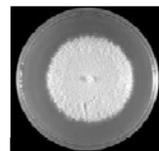
Scedosporium prolificans

- Invasive infection (osteomyelitis, arthritis); competent & compromised
- Maturity in ~5 days; growth inhibited by cycloheximide
- Cottony or moist surface, becomes dark gray/black with white tufts; reverse gray/black
- Olive to brown conidia, ovoid; annellides have swollen base and elongated neck

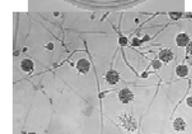


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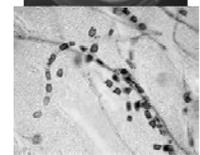
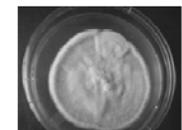
MORE MIMICRY



Chrysosporium spp.



Sepedonium spp.



Malbranchea spp.

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MUCORMYCOSIS

- Rapid growers
- Diabetic susceptibility
- Different cellular nomenclature
- Ribbon-like hyphae; most are aseptate
- www.youtube.com/watch?v=IK0MtXNKgKI



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MUCORMYCOSIS

Organism isolated	No. (%) of all patients	No. of patients who died/total no. with the organism (%)
<i>Rhizopus</i> species	218 (17)	106/218 (49)
Not specified	125 (27)	61/125 (49)
<i>R. oryzae</i>	66 (13)	24/66 (37)
<i>R. rhizodiformis</i>	20 (4)	9/20 (45)
<i>R. microsporus</i>	11 (2)	7/11 (64)
<i>R. nigricans</i>	7 (2)	1/7 (17)
<i>R. stolonifer</i>	1 (1)	1/1 (100)
<i>Mucor</i> species	66 (18)	44/66 (67)
<i>Cunninghamella bertholletiae</i>	34 (7)	29/34 (85)
<i>Apophysomyces</i> species	27 (6)	4/27 (15)
<i>Albizziella</i> species	25 (6)	6/25 (24)
<i>Saksenaella</i> species	21 (5)	8/21 (38)
<i>Rhizomucor pusillus</i>	10 (4)	10/10 (100)
<i>Rhizomucor</i> species	13 (3)	3/13 (23)
<i>Conidiobolus</i> species	10 (2.2)	5/10 (50)
<i>Basidiobolus</i> species	9 (2)	3/9 (33)
<i>Cokeromyces</i> species	3 (0.8)	1/3 (33)
<i>Syncephalastrum</i> species	1 (0.2)	0/1 (0)

Site of infection, or site	Proportion (%) of all patients	No. of patients with the infection who died/total no. with the infection (%)
Brain		
Cerebral	299/329 (91)	180/299 (60)
Intra-orbital	100/100 (100)	100/100 (100)
Sino-orbital	7/69 (10)	18/74 (24)
Skull base	7/69 (10)	12/74 (16)
Extracranial	15/69 (22)	13/74 (18)
Pulmonary		
General	22/492 (4)	1/202 (0.5)
Localized	17/374 (5)	7/371 (2)
Deep subcutaneous	15/254 (6)	13/15 (87)
Subcutaneous	8/371 (2)	8/8 (100)
Soft tissue		
Localized	175/200 (9)	54/176 (31)
Localized	86/176 (5)	10/88 (11)
Deep subcutaneous	43/176 (2)	11/43 (26)
Disseminated	35/176 (20)	33/35 (94)
Other		
General	8/320 (3)	8/87 (9)
Localized	4/287 (1)	7/81 (9)
Local dissemination	4/287 (1)	4/42 (10)
Generalized disseminated	66/329 (20)	66/88 (75)
Kidney	2/329 (0.6)	2/25 (8)
Other solid organ	15/329 (5)	9/15 (60)

Clin. Infect. Dis. 41: 634-653; 2005

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Rhizopus spp.

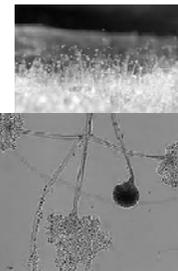
- Common contaminants
- Maturity in ~4 days; growth inhibited by cycloheximide
- Cotton candy--white at first, then gray or yellowish brown; reverse white
- Rhizoids opposite of sporangiophores



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Mucor spp.

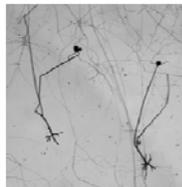
- Common contaminant
- Maturity in ~4 days; growth inhibited by cycloheximide
- Cottony or moist surface, becomes gray; reverse white
- Rhizoids absent



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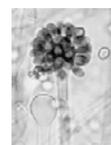
Lichtheimia spp.

- Common contaminant
- Maturity in ~4 days; growth inhibited by cycloheximide
- Coarse, wooly-gray surface--eventually covers surface with "fluff"; reverse white
- Sporangiofores form conical apophysis just below columella; rhizoids alternate



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OTHER MUCORMYCETES



Cunninghamella spp.



Rhizomucor spp.



Apophysomyces spp.



Syncephalastrum spp.

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Antifungal Susceptibility Testing

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CLSI DOCUMENTS OF INTEREST

- M38-A2 Reference Method for Broth Dilution Antifungal Susceptibility Testing of Filamentous Fungi, 2nd ed.; Approved Standard
- M51-A Method for Antifungal Disk Diffusion Susceptibility Testing of Nondermatophyte Filamentous Fungi; Approved Guideline

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BROTH MICRODILUTION

- “Intended for testing common filamentous... moulds, including the dermatophytes, which cause invasive and cutaneous infections, respectively...”

Aspergillus spp. *Fusarium* spp.
Rhizopus spp. *Pseudallescheria boydii*
Scedosporium prolificans *Sporothrix schenckii* (mould)
 Opportunistic monilaceous fungi
 Opportunistic dematiaceous fungi

- “Method has not been used in studies of the yeast or mould form of dimorphic fungi.”

CLSI M38-A2

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BROTH MICRODILUTION

- RPMI 1640 broth (MOPS buffer, 0.2% dextrose)
- 7-day filamentous fungus growth; potato dextrose agar slants
- Flood with saline
Withdraw mixture, particles settle 3-5 min
Upper suspension contains mycotic elements
- Inoculum (OD₅₃₀) dependent upon fungus [0.09-0.30]; range of 0.6 to 3.0 x 10⁶ CFU/mL

CLSI M38-A2

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BROTH MICRODILUTION

Non-dermatophyte filamentous fungi

0.03-16 µg/mL amphotericin B ravuconazole
 ketoconazole itraconazole
 posaconazole voriconazole

0.125-64 µg/mL flucytosine fluconazole

0.015-8 µg/mL flucytosine fluconazole

CLSI M38-A2

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BROTH MICRODILUTION

Dermatophytes

0.06-32 µg/mL ciclopirox

0.125-64 µg/mL griseofulvin
 fluconazole

0.001-0.5 µg/mL itraconazole
 voriconazole
 terbinafine

0.004-8 µg/mL posaconazole

CLSI M38-A2

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BROTH MICRODILUTION

- 35° C ambient air
 - 21-26 hours for mucormycetes
 - 70-74 hours for *Scedosporium* spp.
 - 46-50 hours for most others
 - 21-26 hours for echinocandin testing
 - 46-72 hours for *Scedosporium* spp./echinocandins
 - Amphotericin B: observe 100% inhibition
 - Other agents: observe 50% inhibition
 - Dermatophytes: observe 80% inhibition
 - Echinocandins: lowest concentration resulting in small, compact, rounded hyphae
 - Minimum Effective Concentration (MEC)
- CLSI M38-A2 61

Etest

- Not FDA-approved for filamentous fungi
- Etest MIC and broth microdilution data more comparable for trizoles (>90% agreement) than for amphotericin B (>80% agreement)
- Etest MIC values higher for *S. apiospermum*, *A. flavus*, *S. prolificans* higher than reference values

J. Clin. Microbiol. **39**: 1360-1367; 2001 62

CLINICAL UTILITY

- "Factors related to.....appear to have more value than the MIC as predictors of clinical outcome."
Clin. Infect. Dis. **24**: 235-247; 1997
- "Very few correlations of in vitro results with in vivo response have been reported for mold infections."
Curr. Fungal Infect. Rep. **3**: 133-141; 2009
- "...tests are currently most useful for detecting resistance or outliers based on either assigned in vitro breakpoints or epidemiological cutoffs."
Pfaller *et al.*, Manual of Clinical Microbiology, tenth ed. 63

THE END

- Mostly an observational science (occasional biochemical may help with dermatophytes); note growth distribution and rate of growth
- Antifungal susceptibility testing for moulds continues to be a work in progress
- See you at the Dells



CREDITS

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