


2014 Update: STEC Diagnosis and Surveillance in Wisconsin

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**WCLN Teleconference
March 19, 2014**

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Objectives

- Describe the history of Shiga toxin-producing *E. coli* (STEC) in Wisconsin
- Discuss available diagnostic tests for the detection of Shiga toxins in clinical specimens
- Gain an appreciation for the significance of STEC disease in WI and nationally
- Understand testing that is performed at WSLH and CDC on STEC specimens submitted by clinical health systems


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Objectives

- Become aware of ongoing developments in STEC diagnostic testing (Culture-independent diagnostic testing- CIDT) and their potential effects on both clinical and public health laboratories and officials
- Understand the importance of the partnership between clinical, public and environmental health for effective disease surveillance and prevention

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Shiga toxin-producing *Escherchia coli* (STEC)



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What is STEC?

- Shiga toxin-producing *E. coli* (STEC)—STEC may also be referred to as Verocytotoxin-producing *E. coli* (VTEC) or enterohemorrhagic *E. coli* (EHEC). This pathotype is the one most commonly heard about in the news in association with foodborne outbreaks.

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Pathotypes of *E. coli*

- enterotoxigenic *E. coli* (ETEC)
- enteropathogenic *E. coli* (EPEC)
- enteroinvasive *E. coli* (EIEC)
- enteroaggregative *E. coli* (EAEC)
- diffusely adherent *E. coli* (DAEC)
- enterhemorrhagic *E. coli* (EHEC)

7

Serogroups

Table 1. Laboratory confirmed STEC infections, by serogroup, STEC, with the most commonly reported serogroups listed individually, United States, 2011

| Rank | Serogroup | Number Reported | Percent |
|-------------------------|-----------|-----------------|-------------|
| 1 | O157 | 2366 | 41.3 |
| 2 | O26 | 613 | 10.6 |
| 3 | O103 | 548 | 9.5 |
| 4 | O111 | 322 | 5.6 |
| 5 | O121 | 176 | 3.1 |
| 6 | O45 | 135 | 2.3 |
| 7 | O145 | 74 | 1.3 |
| 8 | O118 | 47 | 0.8 |
| 9 | O89 | 22 | 0.4 |
| 10 | O76 | 14 | 0.2 |
| 11 | O81 | 14 | 0.2 |
| 12 | O8 | 9 | 0.2 |
| 13 | O153 | 6 | 0.1 |
| 14 | O104 | 7 | 0.1 |
| 15 | O113 | 6 | 0.1 |
| 16 | O165 | 6 | 0.1 |
| 17 | O174 | 6 | 0.1 |
| 18 | O71 | 6 | 0.1 |
| 19 | O146 | 6 | 0.1 |
| 20 | O85 | 5 | 0.1 |
| Sub Total | | 4387 | 76.3 |
| All other non-O157 STEC | | 90 | 1.5 |
| Unknown ^a | | 100 | 1.8 |
| Brough | | 79 | 1.4 |
| Undetermined | | 14 | 0.2 |
| Sub Total | | 1376 | 23.8 |
| | | 5763 | 100 |

8

Virulence Factors

- Two distinct Shigatoxins (Stx1 and Stx2)
- Shigella dysenteriae 1
- Range in virulence
- Additional virulence factors.
 - *eae* gene for intimin
 - *ehxA* gene for plasmid encoded hemolysin

9

Virulence Factors

Table 3. Presence of Virulence Genes in Shiga Toxin-Producing *Escherichia coli* Isolates From Patients With Postdiarrheal Hemolytic Uremic Syndrome by Serogroup, 2000-2010

| Serogroup | Shiga Toxin | | | <i>eae</i> ^a | <i>ehxA</i> ^b |
|------------------------------|--------------------|----------------------|-----------------------------|-------------------------|--------------------------|
| | <i>stx1</i> Alone | <i>stx2</i> Alone | <i>stx1</i> and <i>stx2</i> | | |
| O157 (n = 100) ^c | 0/100 | 78/100 (75.7) | 25/100 (24.3) | | |
| O111 (n = 4) ^c | 0/4 | 0/4 | 4/4 (100) | 4/4 (100) | 4/4 (100) |
| O121 (n = 2) ^c | 0/2 | 2/2 (100) | 0/2 | 2/2 (100) | 2/2 (100) |
| O145 (n = 5) | 0/5 | 5/5 (100) | 0/5 | 5/5 (100) | 5/5 (100) |
| O26 (n = 2) ^c | 1/2 (50.0) | 0/2 | 1/2 (50.0) | 2/2 (100) | 1/2 (50.0) |
| O153 (n = 1) | 1/1 (100) | 0/1 | 0/1 | 1/1 (100) | 1/1 (100) |
| O130 (n = 1) | 0/1 | 1/1 (100) | 0/1 | 0/1 | 1/1 (100) |
| Unknown (n = 1) ^d | 0/1 | 0/1 | 1/1 (100) | | |
| Total | 2/119 (1.7) | 86/119 (72.3) | 31/119 (26.1) | 14/15 (93.3) | 14/15 (93.3) |

^a *eae* encodes intimin, and *ehxA* encodes enterohemolysin; *eae* and *ehxA* data were not available for STEC O157 (test not generally performed because genes almost invariably present) or for isolates of unknown serogroup.
^b Shiga toxin data collected for 103 of 135 STEC O157 isolates (76.3%) from 2007 through 2010.
^c Virulence factor information is not available for 1 isolate from each of the following serogroups: O111, O121, and unknown.
^d Excluded 1 Stx1-only STEC O26 isolate from a patient whose stool sample also yielded a stx2-only STEC O157 isolate.

From: Infections in Pediatric Postdiarrheal Hemolytic Uremic Syndrome: Factors Associated With Identifying Shiga Toxin-Producing *Escherichia coli*
Arch Pediatr Adolesc Med. 2012;166(10):902-909. doi:10.1001/archpediatrics.2012.471

10


STEC- Background Information

- CDC estimates 19% of O157 STEC and 9% of non-O157 STEC are outbreak related (majority of cases sporadic)
- STEC are low-infectious dose organisms (10-100 cells)
- STEC virulence dependent upon which virulence factors are present in a given strain (Stx1/Stx2, *eae*, *Ehly*); evidence suggests Stx and *eae* are most significant predictors of serious illness

11

Common Symptoms of STEC Disease

- Diarrhea (sometimes bloody)
- Severe stomach cramps
- Vomiting
- Fever less than 101°F



12

STEC Complications

- Hemolytic Uremic Syndrome (HUS)
 - Occur in about 5–10% of cases
 - Usually STEC O157
 - Renal failure
 - Hemolytic anemia
 - Thrombocytopenia
 - Most recover within a few weeks, but some suffer permanent damage or die

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Sequence of events in STEC infection

| STEC O157 ingested | Non-O157 STEC ingested |
|--|--|
| ↓ 3 - 4 days | ↓ 3 - 4 days |
| non-bloody diarrhea, abdominal cramps <small>(short-lived fever)</small> | non-bloody diarrhea, abdominal cramps <small>(short-lived fever)</small> |
| 80% ↓ 1 - 2 days | 45% ↓ 1 - 2 days |
| bloody diarrhea | bloody diarrhea |
| 94% ↙ 5 - 6 days <small>(up to 2-3 weeks)</small> ↘ | >98% ↙ 5 - 6 days <small>(up to 2-3 weeks)</small> ↘ |
| resolution HUS | resolution HUS |
| 6% | <2% |

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Where does STEC come from?

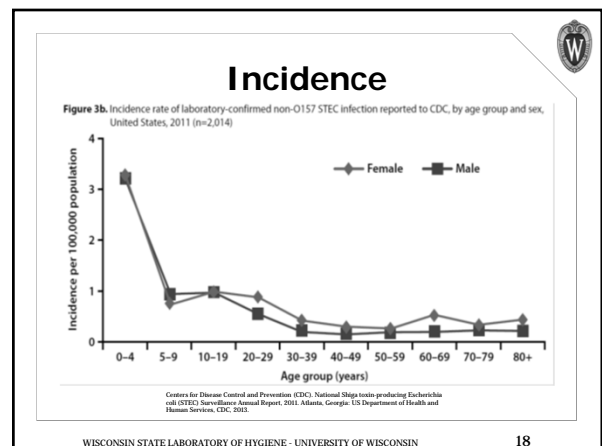
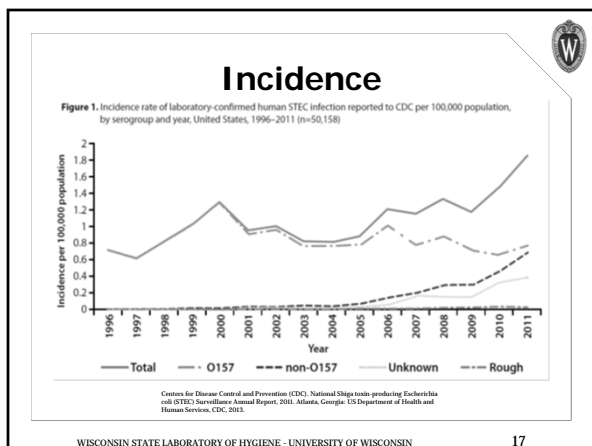
It wasn't me,
the dog did it!

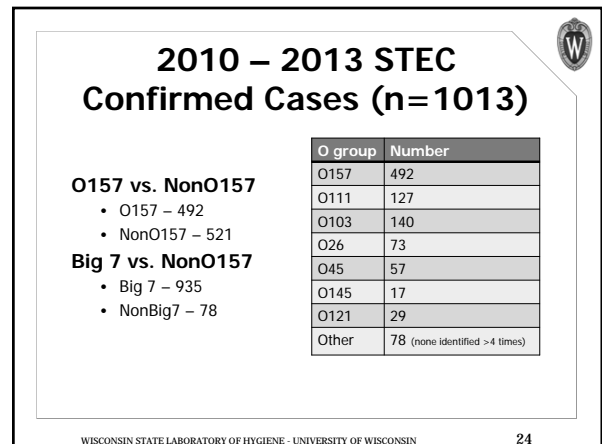
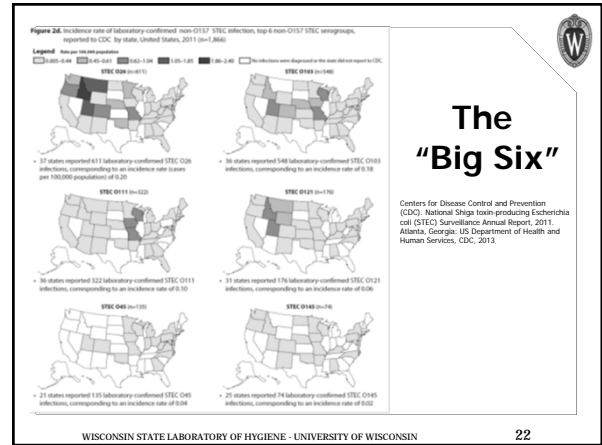
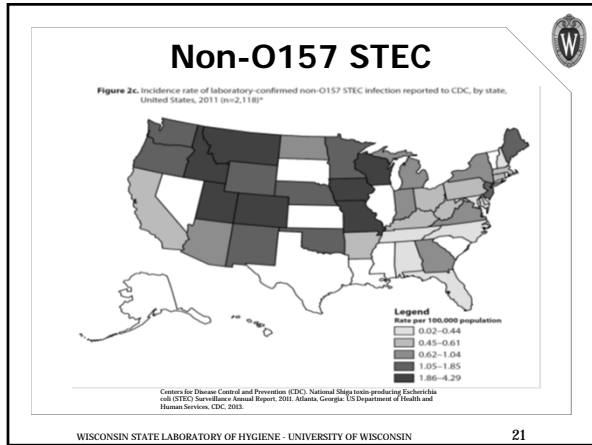
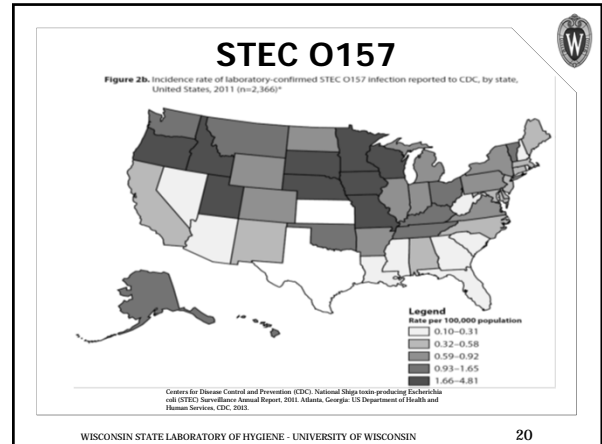
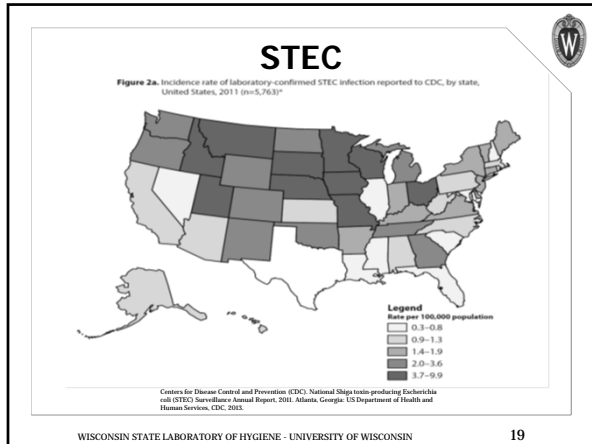
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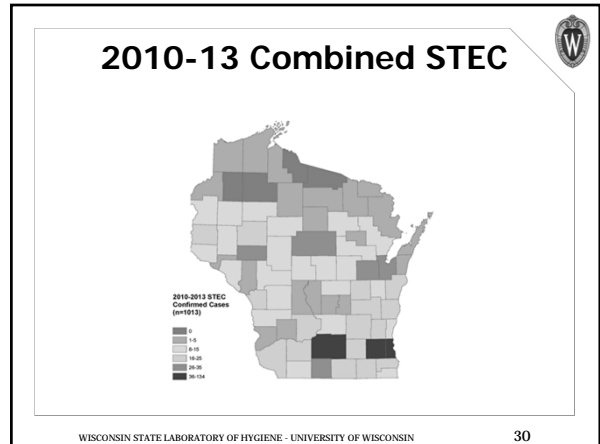
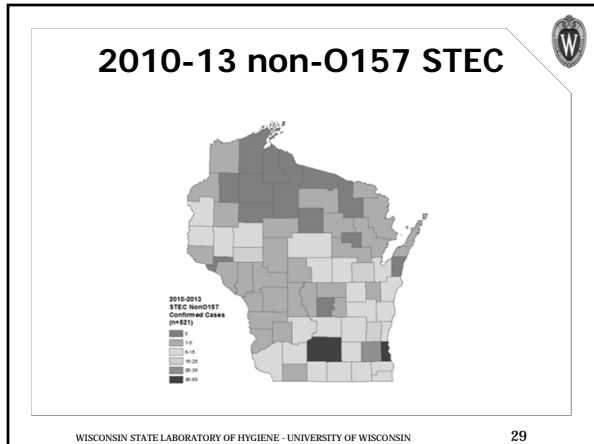
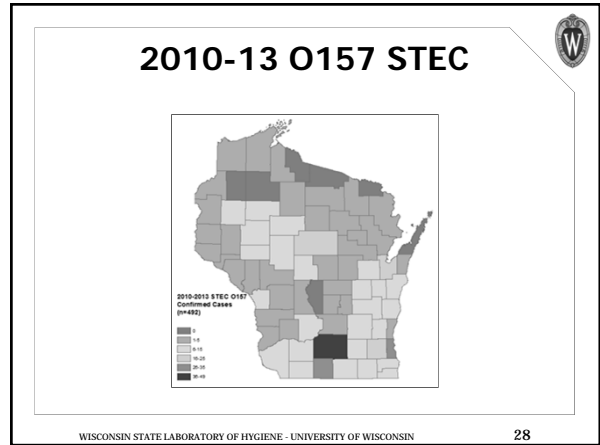
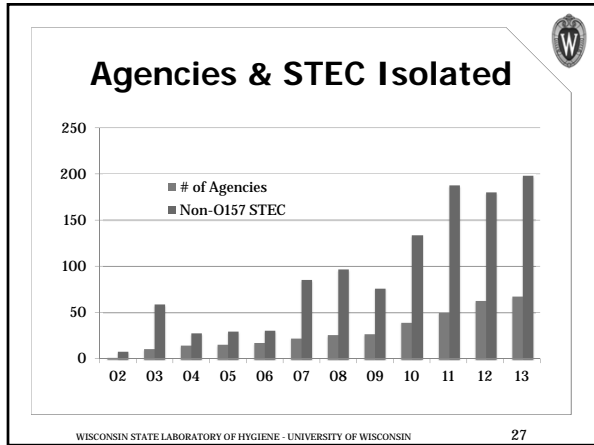
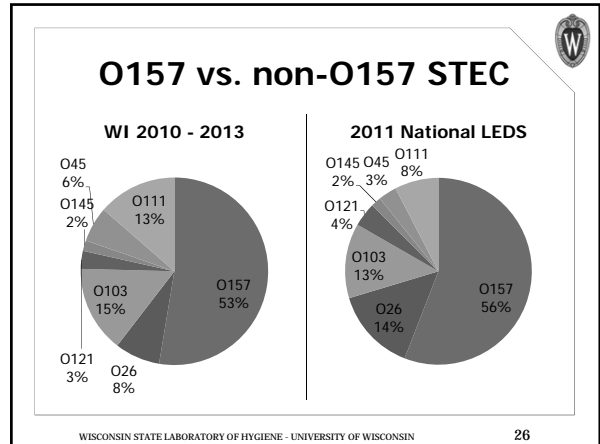
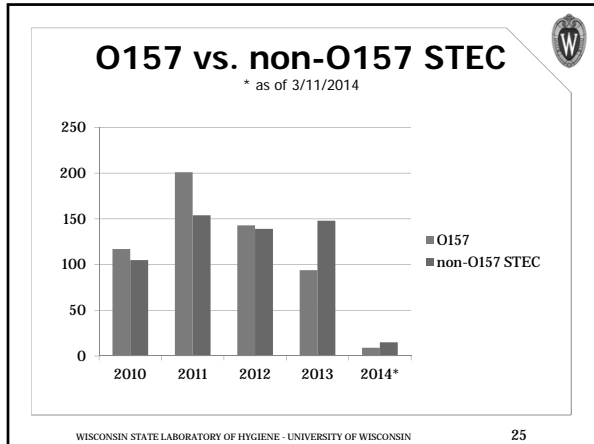
Incidence

- 265,000 illnesses annually in U.S.
- 3,600 plus hospitalizations
- 30 deaths
- 96,534 STEC O157
- 168,698 non-O157

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Wisconsin Case Definition:

- **Confirmed:** A case that meets the laboratory criteria for confirmation. When available, O and H antigen serotype characterization should be reported
- **Probable:** A case with isolation of *E. coli* O157 from a clinical specimen, without confirmation of H antigen or Shiga toxin production, OR a clinically compatible case that is epidemiologically linked to a confirmed or probable case, OR identification of an elevated antibody titer to a known Shiga toxin-producing *E. coli* serotype from a clinically compatible case
- **Suspected:** Identification of Shiga toxin in a specimen from a clinically compatible case without the isolation of the Shiga toxin-producing *E. coli*

Laboratory Criteria for Confirmation:

- Isolation of Shiga toxin-producing *Escherichia coli* from a clinical specimen.
- *E. coli* O157:H7 isolates may be assumed to be Shiga toxin-producing.
- For all other *E. coli* isolates, Shiga toxin production or the presence of Shiga toxin genes must be confirmed to be considered STEC. Examples of assays used to detect Shiga toxin production include Shiga toxin EIA or PCR.

<http://www.cdc.gov/mmwr/PDF/rr/rr5812.pdf>



Recommendations for Diagnosis of Shiga Toxin-Producing *Escherichia coli* Infections by Clinical Laboratories

Guidelines for the Clinical Laboratory:

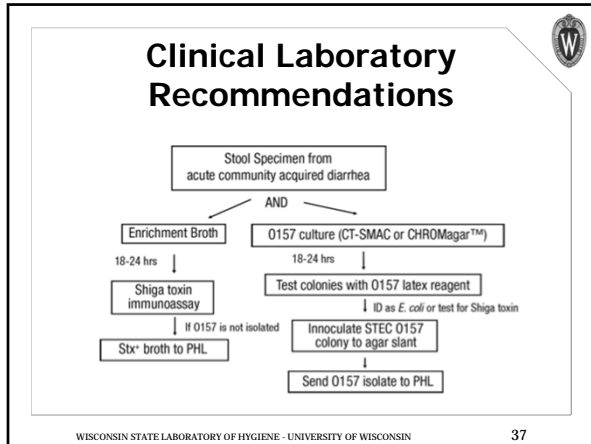
- All stools submitted for testing from patients with acute, community-acquired diarrhea should be cultured for O157 STEC on selective and differential media
- Stools from patients with acute, community-acquired diarrhea should be simultaneously tested for non-O157 STEC with a test that detects either Shiga toxins or the genes that encode for these toxins

Guidelines for the Clinical Laboratory:

- All O157 STEC isolates should be forwarded as soon as possible to a state or local public health laboratory (PHL) for confirmation and molecular characterization (PFGE, MLVA, virulence genes)
- Detection of STEC or Shiga toxin should be promptly reported to the physician, PHL and proper public health authorities

Guidelines for the Clinical Laboratory:

- Any specimens or enrichment broths in which Shiga toxin or STEC has been detected but from which no O157 STEC isolates are recovered, should be forwarded as soon as possible to a state or local public health laboratory for isolation of the STEC



Justification for STEC Testing

- STEC may be as or more prevalent than other enteric bacterial pathogens routinely tested for:
 - STEC may be found in 0-4% of stools
 - *Salmonella* may be found in 1.9-4.8%
 - *Shigella* may be found in 0.2-3.1%
 - *Campylobacter* may be found in 0.9-9.3%

FoodNet Data, CDC

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Early HUS Identification and Treatment is Critical

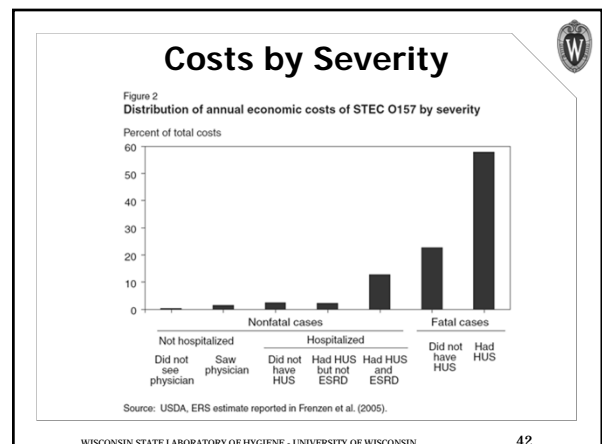
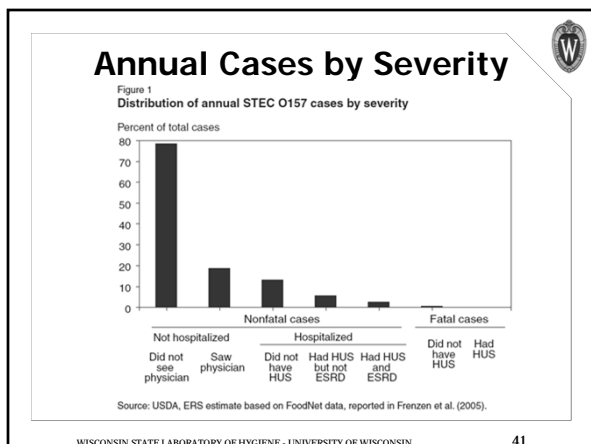
- 15% of culture + *E. coli* O157:H7 patients < 10 yrs of age will develop HUS
- Strong evidence suggesting that STEC isolates that produce Stx2, and in particular Stx2 only, have a higher likelihood to lead to HUS development
- Evidence supporting improved outcomes in individuals that receive prompt intervention (parenteral fluid admin. may prevent renal damage)

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Prevention of Illness is Hard to Measure; Hard to Sell

- Even though non-O157 STEC are low prevalence, single case of HUS may cost the healthcare system over \$6.1 million

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Economic Costs


Table 1
Annual economic cost of illness due to Shiga toxin-producing *E. coli* (STEC) O157

| Severity category | Estimated annual STEC O157 cases ¹ | Estimated annual economic cost ² | Estimated average cost per case ² |
|--|---|---|--|
| | Number | Million dollars | Dollars |
| Nonfatal cases | | | |
| 1. Not hospitalized, did not see a physician | 57,656 | 1.5 | 26 |
| 2. Not hospitalized, saw a physician | 13,656 | 6.0 | 441 |
| 3. Hospitalized, did not have HUS | 1,797 | 10.1 | 5,599 |
| 4. Hospitalized, had HUS but not ESRD | 900 | 9.3 | 30,998 |
| 5. Hospitalized, had HUS and ESRD | 10 | 51.7 | 5,173,594 |
| Fatal cases | | | |
| 6. Did not have HUS | 23 | 92.0 | 3,998,265 |
| 7. Had HUS | 38 | 234.7 | 6,175,500 |
| Total | 73,480 | 405.2 | 5,515 |

Notes: HUS = hemolytic uremic syndrome. ESRD = end-stage renal disease.
¹Derived from the CDC estimate of the annual number of STEC O157 cases, hospitalizations, and deaths in 1997. See Frenzen et al. (2005).
²2003 dollars. The average cost per case was calculated from unrounded totals.
 Source: USDA, ERS using data from Frenzen et al. (2005).

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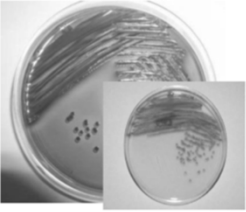
Testing at WSLH



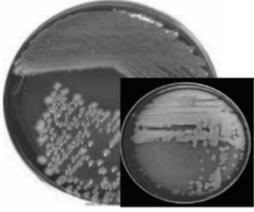
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WSLH Culture Results

CT-SMAC Plate
(sorbitol fermentation positive; sorbitol negative example inset)

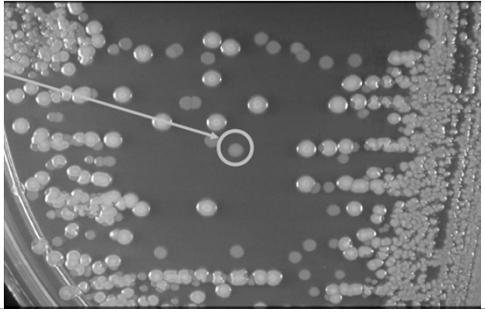


Washed SBA Plate
(non-hemolytic; example of an expected STEC Ehly + result inset)



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E. Coli O157:H7 on SMAC Plate

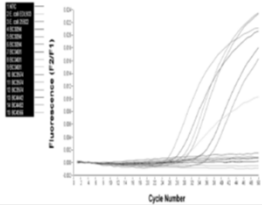


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WSLH PCR Results

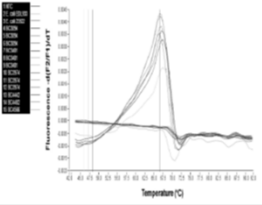
***stx1* and *stx2* detection- fluorescent probes**

LightCycler Analysis Settings Report



***stx1* and *stx2* differentiation by melting temperature**

LightCycler Manual Tm Estimation Report



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WSLH Serogrouping

- O157 and big six non-O157 by slide agglutination



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WSLH PFGE Subtyping

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WSLH PFGE Subtyping

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PFGE

The Pulsed-field Gel Electrophoresis Process

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MLVA

Multiple Locus Variable-Number Tandem Repeat Analysis

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PulseNet

- PulseNet USA is made up of 87 federal, regional, state, and local laboratories. There is at least one PulseNet laboratory in every state.
- In 2012, participating laboratories reported over 60,000 isolates to PulseNet. Over 4,000 of these isolates were from food, animal, or environmental sources.
- Each year in the United States, 1,200 clusters of disease are identified by state and local health agencies and 10–15 multistate or national outbreaks are identified.

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PulseNet

- PulseNet tracks subtypes of:
 - *E. coli* O157 and other Shiga toxin-producing- *E. coli*
 - *Campylobacter jejuni*
 - *Clostridium botulinum**
 - *Listeria monocytogenes*
 - *Salmonella*
 - *Shigella*
 - *Vibrio cholerae*
 - *Vibrio parahaemolyticus*

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Available Testing Options

- ELISA's
- RCA's
- CIDT's
- Future?



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ELISA



ProSpecT

- Common procedure and requires for all tests
- Flexible specimen transport options
- Results within 100 minutes
- Visual or spectrophotometric reading


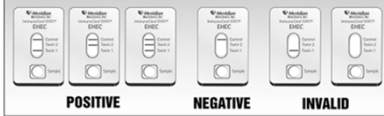


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Rapid Cartridge Assays


DEFINITIVE ANSWERS, CONFIDENT RESULTS

- Fully comply with CDC guidelines
- Detects ALL Shiga toxin-producing E. coli, not just E. coli O157:H7
- Rapid identification to avoid contraindicated antibiotic therapy





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Rapid Cartridge Assays



Detect STEC toxins direct from fecal samples with the new SHIGA TOXIN QUIK CHEK test.



Direct Fecal Testing Results

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Culture-Independent Diagnostic Tests (CIDT)

- They have arrived!
- Two multiplex GI panel tests are now available for use (Luminex xTAG GPP and Prodesse ProGastro SSCS)
- Many others at various stages of development
- Will change the landscape of GI pathogen diagnostics and surveillance

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Luminex xTAG GPP-ASR/RUO Kits

NEW xTAG® Gastrointestinal Pathogen Panel (GPP) for In Vitro diagnostic use

Multiplexed molecular testing for a reliable, actionable result in under 5 hours

- xTAG GPP is the only test that simultaneously detects and identifies the bacterial, viral, and parasitic pathogens responsible for over 90% of cases of infectious diarrhea*
- xTAG GPP provides:
 - More answers per sample than any other method
 - A reliable answer, in time to affect patient care
 - Better use of time and resource

| Method | Tests for | Turn-around time | Diagnostic yield |
|------------------------------|---|--|---|
| Stool culture | Single bacterial pathogen per test | 24-36 hrs | 50 to 75% |
| One and parasite (OAP) assay | Parasitic pathogens | Several days - several weeks for collected stool | 10 to 30%* |
| Rapid tests | Single pathogen per test | 30-30 min | Varies |
| Rapid stool PCR | 1-3 pathogens per test | Under 5 hours | Varies, depends on the pathogen target, individual performance and method of sample |
| ELISA | Single antigen/antibody per test | 0-24 hours | Varies |
| xTAG GPP | 10 to 12 bacterial, viral, and parasitic pathogens in a single test | Under 5 hours* | 90%* |

*xTAG GPP - the only available multiplexed molecular test that simultaneously detects and identifies the bacterial, viral and parasitic pathogens in a single test

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Prodesse ProGastro SSCS GI Panel

HOLOGIC



Instructions for Use

For the detection and differentiation of Salmonella, Shigella, Campylobacter (C. jejuni and C. coli only, undifferentiated), nontyphoid Shigella, and Shiga Toxin Producing E. coli (STEC) Shiga Toxin 1 (stx1) and Shiga Toxin 2 (stx2) genes.

ISO 9001 CE REF


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bioMérieux (BioFire) GI Panel

FilmArray Gastrointestinal Panel

1 Test. 23 Targets. All in about an hour.

| Bacteria | Protozoa | Viruses |
|---|---|--|
| <ul style="list-style-type: none"> Aeromonas Campylobacter Clostridium difficile (Toxin A/B) Plasmodium shigeloides Salmonella Yersinia enterocolitica Vibrio Vibrio cholerae | <ul style="list-style-type: none"> Cryptosporidium Cyclospora cayentensis Entamoeba histolytica Giardia lamblia | <ul style="list-style-type: none"> Adenovirus F 40/41 Astrovirus Norovirus GI/GII Rotavirus A Sapovirus |



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The Future of "DNA Fingerprinting"

- Whole genomic sequencing
 - Requires pure cultures of bacteria
 - Use whole genome sequencing which by itself is a very powerful DNA fingerprinting method to help identifying and investigating outbreaks
 - Identify genetic targets for surveillance
 - Build reference database for metagenomics

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The Future of "DNA Fingerprinting"

| Key Factors | Next Generation Sequencing (NGS) | Current "Gold Standard" (PFGE) |
|--|--|---|
| Time to get data | 1-2 days | Days |
| Time to analyze data | Hours to days | Hour or less |
| Cost per bacteria isolate (for reagents only) | ~\$150-300 (depends on platform) | ~\$10-15 |
| Ability to differentiate and cluster isolates | To be determined but likely very good | Well-established for bacteria tracked by PulseNet, few exceptions |
| Potential for automation | Yes | No |
| Intended objective | Genetic serotyping, virulence, antimicrobial resistance profiling, and subtyping possibly based on SNPs or other variant regions | Only subtyping |

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The Future of "DNA Fingerprinting"

- Metagenomics
 - No pure culture of bacteria required
 - Sequence select genetic targets or all bacteria in a patient sample (stool, blood, etc.)
 - Identify bacteria making a patient sick and subtype at the same time

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Studies

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2011-2012 FoodCORE Study

- **FoodCORE: Foodborne diseases Centers for Outbreak Response Enhancement**
- Public Health systems (Lab, Epi and Environ) granted funding to improve and model enhanced foodborne disease diagnostics, surveillance and prevention of further disease
- WI was one of 7 systems awarded funding in 2009; began laboratory activities in 2010
- Addressing STEC disease is one of the major foci for WI FoodCORE activities

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70

2011-2012 FoodCORE Study

- Three clinical sites not performing routine Shiga toxin (Stx) screening of stool specimens were enlisted to participate
 - **Site 1** inoculated GN enrichment broth and sent to WSLH for microplate EIA Stx screening
 - **Site 2** performed microplate EIA screening for Stx in-house
 - **Site 3** performed Stx screening using a lateral flow rapid cartridge assay

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71

2011-2012 FoodCORE Study

- Total of 8180 stools were screened for Stx from the three clinical system sites (Mar 1, 2011-Dec 31, 2012)
- There were 83 primary STEC isolates detected, of which 46 were O157 and 37 non-O157 STEC
- STEC recovered from the three clinical system sites represented 15% of the O157 STEC and 12% of the non-O157 STEC reported in WI over the study period (14% of total STEC WSLH rec'd)

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72

STEC isolated during study:

- E. coli* O5:NM
- E. coli* O121:H undetermined
- E. coli* O145:NM
- E. coli* O26:NM and H undetermined
- E. coli* O146:H21
- E. coli* O103:H undetermined
- E. coli* O157:H7 and O157:NM
- E. coli* O45:H2 and H undetermined
- E. coli* O111:NM
- E. coli* O91:H14
- E. coli* O undetermined: H undetermined*

*CDC unable to serotype

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73

2011-2012 FoodCORE Study

- All of the non-O157 STEC and 6% of the O157 STEC (negative by culture at the clinical system site) would have been unreported had the three clinical system sites not been screening for Stx
- Of the 75 cases (out of 83) for which epidemiological data is available, there were 23 hospitalizations (18 O157, 5 non-O157), no deaths or HUS cases and the median age was 19 (age range from 0-89 years)

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74


2011-2012 FoodCORE Study

- Positivity rates for Stx among all three clinical system site stool specimens averaged near 1% over the study period
- There were a total of 6 O157 STEC isolates (7.2%) during the study period that were included in national PulseNet clusters; no non-O157 STEC were included in national clusters
- Both the microplate EIA and RCA Stx diagnostic assays performed well; of all positive GN broths, WSLH isolated STEC from 92%

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75

It Takes Teamwork...



Effective STEC disease diagnosis, surveillance and prevention takes a concerted effort by clinical health systems, public health laboratories, public health epidemiologists and env. health officials

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How do We Partner to Prevent STEC and Other Diseases?




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Role of Clinical Health Systems

- Disease diagnosis and patient management is the primary focus; Clinical laboratories will generally be the first to see infectious disease cases
- Responsibility to notify local health officials, within the patient jurisdiction, of reportable disease cases as they detect them
- Prevention of spread of further communicable disease from index case(s)
- Responsible use of antibiotics; reduce development of antimicrobial resistance

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Role of WSLH

- Laboratory service for the Wisconsin Division of Public Health (WDPH)
- Laboratory response to clusters of disease or pathogens of public health significance; emergency response
- Surveillance for public health pathogens
- Antimicrobial resistance monitoring
- Public health education and safety
- Research and specialized testing

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Role of Local Health Officials

- Receive reportable disease information from clinical health systems or the public within their jurisdiction
- Investigate disease cases or clusters
- Disseminate disease information to WDPH



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Role of WDPH

- Receive disease reports from local health, clinical health and environmental health officials
- SOS (Surveillance and Outbreak Support) Team will interview and enter epi data from cases in real time into a database; Epi's often have data in hand when they receive lab data from WSLH

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Role of WDPH

- Receive lab data (PFGE subtyping, AST, Stx) from WSLH
- Work with other state and CDC epi's as needed to respond to regional or national clusters of illness
- Lead and coordinate cluster investigations
- Public Health education

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82

Role of Environmental Health Sanitarians

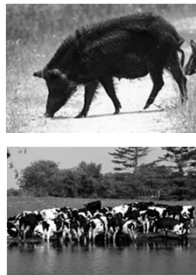
- Work closely with Dept of Agriculture, Trade and Consumer Protection officials
- Conduct environmental health inspections and related activities for locations such as:
 - Restaurants
 - Hotels
 - Swimming pools and beaches
- Enforce public health laws and sanitary codes
- Investigate outbreaks, injuries, exposures, etc



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83

2006 Multi-State Spinach *E. coli* O157: H7 Outbreak



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84

O157:H7 Spinach Outbreak- 2006

- Cluster of *E. coli* O157:H7 detected by PFGE subtyping in WI
- Reports to WDPH of multiple cases of HUS in the Milwaukee area
- Total of 204 cases of *E. coli* O157:H7 attributed to the spinach exposure; 49 of which were WI residents
- There were 104 patients hospitalized (51%), 31 cases of HUS (16%) and 3 deaths (1 WI)

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85

O157:H7 Spinach Outbreak- 2006

- Epi investigation traced source back to spinach field in CA
- Environmental sampling in the area led to the *E. coli* O157:H7 outbreak strain being detected from cattle on an adjacent farm, wild pig feces in the field and water in an irrigation channel in the field (confirmed by PFGE and MLVA subtyping)

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86

O157:H7 Spinach Outbreak- 2006

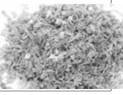
- Interestingly, one WI case was also determined to have a dual STEC infection; the outbreak O157:H7 and a non-O157 STEC
- NM state laboratory isolated the same non-O157 STEC from a bag of the implicated spinach
- How many other non-O157 STEC infections were there from the implicated spinach exposure?

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87

O104:H4 STEC Outbreak


- May 2011 cluster of HUS cases detected in Germany
- No equivalent of PulseNet in Germany; Surveillance based on HUS case reporting
- Almost 4000 cases, 1000 HUS cases and 50 deaths attributed to the outbreak in the end
- Eventually linked to contaminated sprouts germinated from fenugreeek seeds that originated in Egypt



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O104:H4 STEC Outbreak

- O104:H4 novel strain; only seen in 2006 Japan and 2009 Rep. of Georgia human illness cases
- High interest in enhanced virulence ; genomic sequencing revealed the origin of this strain was likely an O157:H7 strain exchanging genetic material with an EAEC O104:H4 in the gut of a human



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O104:H4 STEC Outbreak



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O45 STEC Associated with a WI Meat Processor- 2010-12

- Dec 2010, WSLH identifies matching/ closely matching PFGE patterns for four O45 STEC isolates (3 WI residents and 1 MI resident)
- Epi investigation leads to 3 more probable cases (1WI and 2 MI) and an interesting foodborne exposure scenario...

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O45 STEC Associated with a WI Meat Processor- 2010-12

- Case patient food exposure histories lead to consumption of bear meat sticks as illness source
- Meat sticks were prepared with beef and bear meat
- Venison contamination found in beef sausage
- True source (beef, bear, venison)?



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O45 STEC Associated with a WI Meat Processor- 2010-12

- WDPH Epi's thought this investigation was in the books but...
- Nov 2011, a new O45 STEC isolate with the same PFGE pattern is identified at WSLH
- Epi investigation finds that patient had venison processed at the same meat processor implicated in the 2010 outbreak

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O45 STEC Associated with a WI Meat Processor- 2010-12

- Fast forward to May 2012: Dane county resident O45 STEC isolate determined to match the outbreak strain by PFGE
- Epi investigation by WDPH and local HD reveals (you guessed it) this patient also had venison processed at the same meat processor
- Health officials determined to find source of these infections

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94

O45 STEC Associated with a WI Meat Processor- 2010-12

- In addition to environmental sampling, all employees were screened for Stx
- One employee Stx positive in June 2012
- Interestingly though, the culture yields an O146:H21 STEC, not O45 STEC as was suspected
- To date, no further O45 STEC cases linked to the meat processor...but hunting season begins again next Fall!

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95

WI Tiger Meat Outbreaks- 2012 and 2013



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96

WI Tiger Meat Outbreaks- 2012 and 2013

- It's a Holiday Tradition!
- Jan 2013, two cases of *E. coli* O157:H7 with matching PFGE patterns reported to WDPH; two more would be linked later
- Epi investigation links the cases to raw ground beef purchased at the same meat market
- 17 cases would be identified with raw beef exposure (14 consumed/ 3 cross contam.)

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97

WI Tiger Meat Outbreaks- 2012 and 2013

- Dec 2013...It's still a Holiday tradition!
- 3 *E. coli* O157:H7 STEC isolates identified that have matching PFGE patterns
- Epi investigation determines 2/3 had consumed raw ground beef as Tiger Meat/ cannibal sandwiches
- Definitive source of beef not identified after a thorough investigation by PH officials

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98

Culture Independent Diagnostic Tests- Impact on Patient Mgmt

- In many cases, will detect pathogens more quickly than traditional detection methods
- Multiplex assays will pick up multiple pathogens in a single specimen...clinical significance of one, both, all?
- Loss of isolate for antimicrobial susceptibility testing (if needed)

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99

Culture Independent Diagnostic Tests- Impact on Public Health

- Potential loss of isolates if clinical health system discontinues enteric bacterial culture (current basis for surveillance)
- Loss of isolate for serotyping and subtyping
- Loss of isolate for antimicrobial resistance testing
- Greater resources necessary to perform enteric culture on specimens received from clinical health systems

100

Specimen Submissions

- As with other enteric specimens, STEC specimens may be submitted to WSLH via overnight courier as part of the **Wisconsin Enteric Pathogen Surveillance (WEPS) Program**
- Contact Dunham Express courier service: 800-236-7127 (account 7271)
- For technical questions, call WSLH: 800-862-1013

101

CDD Requisition Form A

| Bacteriology | Bacteriology | Parasitology |
|---|---|---|
| MP0020 Bacterial ID, Non-Enteric Public Health Panel (PHS) Suspect Organism | MP0019 ETEC Shiga Toxinogen Exotoxins | MP0060 Antigen Identification |
| MP0043 Bacterial Typing (PFGE) Organism | WSLH assigned | MP0062 Cryptosporidium, Giardia, Tox |
| MP0047 Bacteroid Culture | WSLH assigned | MP0063 Ova & Parasites, Intestinal |
| MP0051 Bacteroid PCR | • Burkholderia mallei/paradeimatis • Bacillus anthracis • Francisella • Yersinia pestis | MP0068 Parasites Blood Stain Suspect Parasite |
| Bordetella Testing (C&I 800-662-2018) | Investigative Bacteriology Surveillance (IBS) St | MP0069 Parasites Tissue Stain Suspect Parasite |
| IC00111 Chlamydia GC NAAT | MP0066 Group A Beta Streptococcus | MP0064 Worm Identification |
| IC00113 Chlamydia trachomatis NAAT | MP0068 Group B Beta Streptococcus | Unassigned |
| IC00112 Chlamydia pneumoniae NAAT | MP0067 Group D Beta Streptococcus | Y001701 Enterovirus PCR (CF) |
| Y001541 Chlamydia trachomatis Culture | MP0068 Group E Beta Streptococcus | Y001706 Herpes Simplex Virus PCR (Genital, Derm) |
| MP0046 CRE (KPC) NDH-1 PCR | MP0064 Streptococcus pneumoniae | Y001705 Herpes Simplex Virus PCR (CSF) |
| Organism Antibiogram Susceptibility Results | MP0065 Streptococcus pneumoniae | Y001711 Measles Virus PCR |
| MP0060 Diphtheria Culture | Mycobacteriology | Y001714 Mumps Virus PCR |
| MP0069 Bacterial Pathogen Matrix Identification (PHS) Suspect Organism | MD0026 Mycobacteria (AFB) Stain and Culture | Y001715 Norovirus PCR |
| MP0047 Haemophilus ducreyi Culture | MD0025 Mycobacteria Dot-Blot Identification | Y001723 Rubella Virus PCR |
| MP0045 Legionella PCR | MD0022 Mycobacterium avium Complex (MAC) Susceptibility | Y001727 Varicella Zoster Virus PCR |
| MP0049 Shiga Toxin-EDA | MD0024 Mycobacterium tuberculosis Susceptibility to Line Drugs | Y001725 Rubella Virus PCR |
| MP0036 Shiga Toxin-PCR Screen | MD0027 Mycobacteria Rapid Growth Susceptibility | Y001722 Varicella Zoster Virus PCR |
| MP0064 S. aureus in VISA, VISA Confirmation | MD0026 Mycobacterium tuberculosis PCR (Disseminated) Var. Sp. | Other Tests Specimen |
| MP0049 Stool Culture, Routine Suspect Organism | Stain Result | |

WHITE-RETURN TO WSLH
YELLOW-KEEP FOR YOUR RECORDS

102

Summary

- STEC disease is a significant cause of morbidity and mortality in WI and nationally; there are significant costs, both human and monetary
- here are multiple diagnostic testing options available for Stx detection with many more in development
- WI clinical health systems have been testing for Stx for many years; currently there is effective statewide Stx diagnostic testing which is critical for STEC diagnosis, surveillance and prevention

103

Summary

- CIDT's have proven beneficial to STEC and other disease diagnoses; however they have the potential for adverse impact on current isolate-based public health surveillance systems
- Effective STEC disease detection, surveillance and prevention requires a strong partnership among clinical and public health agencies and officials; we believe such a partnership exists today in WI

104

Acknowledgements

- Rachel Klos, Traci DeSalvo and Justin Kohl-WDPH state foodborne disease epidemiologists
- Three FoodCORE STEC enhancement initiative sites:
 - Marshfield Clinic
 - St. Mary's Hospital- Madison
 - Fort Healthcare
- Colleagues in the WSLH Bacteriology and PFGE laboratories
- Clinical health systems and laboratorians in WI

105

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Questions?



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