

BATTLING INFECTIONS OF THE CENTRAL NERVOUS SYSTEM

Organisms, Risk Factors, and Laboratory Diagnostics

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Outline

- Overview of meningitis
- Address P.A.C.E. Goals
 - Identify organisms commonly associated with CNS infection
 - Discuss the factors which put patients at risk for these infections
 - Explain the strengths and weaknesses of current diagnostic methods
- Clinical and financial impact of rapid results
- Conclusion

CNS Infections

Meningitis vs. encephalitis

- Meningitis
 - Infection/ inflammation of meninges (3 layers)
 - Brain, spinal cord, or intracranial spaces (CSF)
 - Bacterial, viral, fungal
- Encephalitis
 - Infection/inflammation of brain parenchyma
 - Infections & non-infectious causes (Injury, cancer, drugs)
 - Diffuse → more typically viral
 - Cerebritis is more focal presentation

Meningitis (Coverings of the Brain)

Encephalitis

CNS Infections

Routes of infection

- Direct invasion
 - "Natural" → Access through sinus, conjunctiva
 - URT flora, amoeba, HSV/GBS (neonates)
 - "Traumatic" → Open cranial or spinal wound
 - Environmental GNR, Skin flora, Mycobacteria, fungi
 - "iatrogenic" → Medical device related, e.g. shunt, drain
 - Skin flora
- Haematogenous
 - Following infection
 - Pneumonia, BSI
- Endogenous
 - Reactivation of latent infection → HSV, CMV, EBV, JC

Organisms

Common causes

- Viruses (65-75%)
 - Commonly self-resolving "aseptic meningitis"
 - May be life-threatening in immunocompromised host
- Bacteria (15-20%)
 - Severe, acute meningitis
 - High mortality if untreated
- Fungi (5-8%)
 - Most commonly yeast, dimorphic fungi
 - More common in compromised
- Amoebic (<1%)
 - Associated with environmental exposure
 - Almost uniformly fatal

CSF characteristics

- General rule
 - Subject to variation by species, severity of infection, etc.
 - Viral may initially have neutrophilic predominance

Test	Appearance	Protein	WBC μ L	Protein mg/dL	Glucose mg/dL	Chloride
Normal CSF	Clear	90 – 180 mm	0-8 lymph.	15-45	50-80	115-130 mEq/L
Acute bacterial meningitis	Turbid	Increased	1000 -10000	100 – 500	< 40	Decreased
Viral meningitis	Clear	Normal to moderate increase	5-300, rarely >1000	Normal to mild increased	Normal	Normal
Fungal meningitis	Clear	Increased	40-400 mixed	50-300	Decreased	Decreased

Risk factors

- **Age**
 - Neonates
 - Congenital infection → CMV, HSV
 - Vertical transmission during birthing → HSV, GBS
 - Young children
 - High rate of URT colonization → *S. pneumoniae*, *H. influenzae*, *N. meningitidis*,
 - Questionable hygiene → Enterovirus
- **Immune state**
 - Compromised → HIV/AIDS, HSCT, SOT
 - Typically more severe in compromised
 - Often re-activation of latent infection (VZV, HSV, JCV); fungal infections
- **Medical hardware**
 - CNS shunts/drains/catheters
 - Skin flora, GNRs → biofilm

BACTERIAL MENINGITIS

Clinical impact of bacterial meningitis

- **Acute bacterial meningitis is life-threatening condition (i.e. critical value!)**
 - Critical role for Laboratory
 - Differentiate from viral meningitis (more common/less severe)
 - Benefit from prompt abx
- **General risk factors**
 - Age, colonization status, indwelling devices

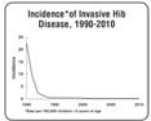
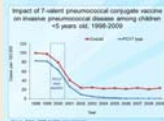
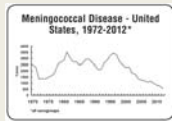
Demographic	Common Bacterial Etiology
Neonate	<i>S. agalactiae</i> , <i>E. coli</i> , <i>L. monocytogenes</i>
Infants, young children	<i>H. influenzae</i> , <i>S. pneumoniae</i> , <i>N. meningitidis</i>
Young adult	<i>N. meningitidis</i>
Adult	<i>S. pneumoniae</i> , <i>N. meningitidis</i>
Elderly	<i>S. pneumoniae</i> , <i>L. monocytogenes</i> , <i>Enterobacteriaceae</i> , NLFs
CNS shunt/drain	CoNS, <i>S. aureus</i> , <i>Corynebacterium</i> spp., <i>Enterobacteriaceae</i> , NLFs

Epidemiology

- **Initial surveys in early 1980's**
 - Attack rate of 3.0-6.0 cases/100,000
 - 10-20x higher for children <1 yoa
 - Common agents...source of infection?
 - 75-85% → *H. influenzae*, *S. pneumoniae*, *N. meningitidis*
 - 2-5% → *S. agalactiae* (neonates, now elderly as well) *L. monocytogenes*
 - 2-5% → *Enterobacteriaceae*, *Staph* spp., *Strep* spp., *P. aeruginosa*
- **Vaccine impact?**

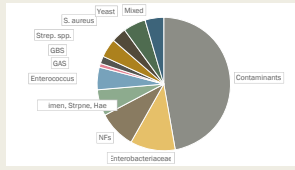
Epidemiology

- **Vaccine impact?**
 - Dramatic reduction, changing epidemiology
 - Hib conjugate (1990) → >99% reduction from 54 to 0.3 cs/100k
 - Pneumococcus (2000)
 - Pediatric conjugate 7/13; 97% effective, 30-60% decrease in pneumococcal meningitis
 - Adult polysaccharide 23; High risk adults → includes 75-90% of CSF isolates
 - *N. meningitidis* ACYW (2005) B (2015) → 65% reduction from 0.92 to 0.33 cs/100k
 - Not recommended for general population in USA (low risk)
 - Recommended for laboratory workers (60-100x higher incidence than general public), college freshmen

Epidemiology

- **Current causes of bacterial meningitis**
 - WDL (2 years)
 - CoNs leading - CSF shunts
 - real vs. contaminant!
 - Broth or plate only? Single CFU? 1st quadrant?

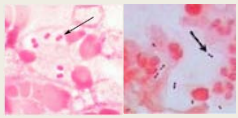


ID	# cultures	% cultures
<i>S. epidermidis</i>	39	35.5%
CoNS	5	4.5%
<i>Corynebacterium</i>	1	0.9%
<i>P. acnes</i>	4	3.6%
<i>Bacillus</i> spp.	2	1.8%
<i>Micrococcus</i>	1	0.9%
<i>S. pneumoniae</i>	4	3.6%
<i>N. meningitidis</i> *	3	2.7%
<i>E. coli</i>	4	3.6%
<i>S. marcescens</i>	3	2.7%
<i>Enterococcus</i>	6	5.5%
<i>P. aeruginosa</i>	7	6.4%
<i>Enterobacter</i> spp.	4	3.6%
<i>S. agalactiae</i>	2	1.8%
<i>S. pyogenes</i>	1	0.9%
<i>Acinetobacter</i>	3	2.7%
<i>Vitidans</i> gr/ <i>Strep.</i>	5	4.5%
<i>S. aureus</i>	4	3.6%
<i>P. mirabilis</i>	1	0.9%
Mixed pathogens	5	4.5%
<i>Candida</i> spp.	2	1.8%
<i>C. neoformans</i>	4	3.6%
Total	110	

Detection methods

- Direct exam
- Antigen
- Culture
- NAAT/PCR

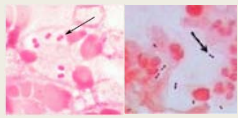
Detection methods



Direct exam

- Critical value!
 - Critical value → establish acceptable TAT for reporting (<2 h)
 - Cellularity (RBCs, PMN vs. Monos)
 - Bacteria (presence, relative abundance, morph, location)
- Sensitivity?

Detection methods




Direct exam

- Critical value!
 - Critical value → establish acceptable TAT for reporting (<2 h)
 - Cellularity (RBCs, PMN vs. Monos)
 - Bacteria (presence, relative abundance, morph, location)
- Sensitivity? **Concentrate!**
 - Stains
 - Gram stain → Morphology, GP/GN
 - Variable sensitivity (LoD ~ 10⁶ cfu/mL), small GNR
 - Specificity for "rare GPC"
 - Acridine Orange → Morphology only
 - Fluorescent nucleic acid stain
 - Increased sensitivity (LoD ~ 10⁴ cfu/mL)
 - Sensitivity: "Rare GNR"; Specificity: "Rare GPC"


Pathogen	CSF Gram stain	Sensitivity (%)
<i>Haemophilus influenzae</i>		25-65
<i>Streptococcus pneumoniae</i>		69-93
<i>Neisseria meningitidis</i>		30-89
<i>Listeria monocytogenes</i>		10-35
<i>Streptococcus agalactiae</i>		80-90
<i>Streptococcus pyogenes</i>		66-73
<i>Streptococcus suis</i>		30
<i>Staphylococcus aureus</i>		20-44

CLINICAL MICROBIOLOGY REVIEWS, July 2010, p. 467-492




Journal of Medical Microbiology (2005), 54, 943-950

Detection methods



Antigen

- Types
 - Latex agglutination, Enzyme assay
 - *N. meningitidis*, HIB, *S. pneumoniae*, GBS
 - Simple, faster than culture (10-20 min.)
- Sensitivity?
 - vs. Gram stain? Non-viable organisms? Abx?



Detection methods

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 - Simple, faster than culture (10-20 min.)
- Sensitivity?
 - vs. Gram stain? Non-viable organisms? Abx?
 - Sensitivity similar or inferior to Gram stain
 - No added sensitivity for patients on Abx
 - Potential for "false sense of security" w/ neg result
 - "No substantial benefit beyond concentrated Gram stain"

Organism	Total no. of specimens	No. (%) of specimens not detected by:	
		BAT	Gram stain
Detected by culture ^a			
<i>Streptococcus pneumoniae</i>	22	6 (27)	3 (14)
Group B <i>Streptococcus</i> species	7	3 (43)	0 (0)
<i>Neisseria meningitidis</i>	7	3 (43)	1 (14)
<i>Haemophilus influenzae</i> type b	2	0 (0)	1 (50)
Not detected by culture ^a	4	1 (25)	0 (0)
Total	42	13 (31)	5 (12)

JOURNAL OF CLINICAL MICROBIOLOGY, Apr. 2010, p. 1504-1505

Detection methods

Antigen

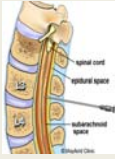

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JOURNAL OF CLINICAL MICROBIOLOGY, Apr. 2010, p. 1504-1505

- MIC.22550 - Back-up cultures required on both AG-positive and negative CSF specimens

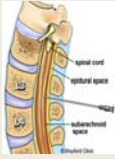

Detection methods

Culture

- “Gold standard”
 - Tube #2 → Centrifuge or plate entire volume (<1 mL)
 - Aerobic culture (BAP, CHOC) + Thio broth
- Sensitivity?

Detection methods

Culture

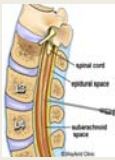

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 - Tube #2 → Centrifuge or plate entire volume (<1 mL)
 - Aerobic culture (BAP, CHOC) + Thio broth
- Sensitivity?
 - 80-95%
 - Factors impacting culture sensitivity
 - Organism: 95% *H. flu.*, 90% *S. pneumo.*, 80% *N. mening.*
 - CSF volume: some infections $\leq 10^3$ CFU/mL
 - Abx usage: 60-80% decrease in sensitivity
 - Gold standard: Cytology (>1000 WBC/uL, >80% PMN)
- Blood culture added benefit?

None	< 4h	> 4h	>12 h	>24 h
84% (146/159)	72% (18/25)	55% (26/47)	58% (19/33)	59% (17/29)

PEDIATRICS Volume 122, Number 4, October 2008

CLINICAL MICROBIOLOGY REVIEWS, July 2010, p. 467-492

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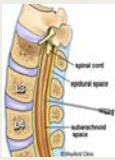

PEDIATRICS Volume 122, Number 4, October 2008

Positive CSF Gram-stain results ^a	95/150 (63)
Positive blood culture results ^b	123/187 (66)
Positive CSF culture results ^c	136/154 (88)

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JOURNAL OF CLINICAL MICROBIOLOGY, Aug. 2003, p. 3851-3853

CLINICAL MICROBIOLOGY REVIEWS, July 2010, p. 467-492

Detection methods

NAAT/PCR

- Potential advantages
 - Speed, sensitivity, less impact of abx treatment
 - Highly desirable for *N. meningitidis*
 - Lowest GS and culture sensitivity among bacterial pathogens
 - Rapidly progressing and fatal
- Performance
 - Sensitivity: 90-97%; Specificity: >99%
 - Result available in 2-3 h
 - ~20% decrease in sensitivity if abx

Test	No. of samples		Sensitivity (%)	Specificity (%)
	Positive	Negative		
Gram staining	25	13	66	100
Culture	21	17	55	100
Green staining culture	33	5	87	100
PCR	37	1	97	99.6

JOURNAL OF CLINICAL MICROBIOLOGY, Aug. 2003, p. 3851-3853

Collection of CSF samples	Test	Tests done	Positive (%)		p-value
			CSF	Blood	
Before onset	PCR	31	70	100	
	of antibiotic treatment	EA	35	70	
After onset	PCR	36	13	82	0.02
	of antibiotic treatment	EA	9	7	0.02
Culture		16	1	17	0.01

Clin Microbiol Infect 2006; 12: 137-141

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 - Result available in 2-3 h
 - ~20% decrease in sensitivity if abx
- PCR considered “gold standard” for *N. meningitidis* in UK
 - Observed a 56% increase in lab-confirmed meningococcal disease

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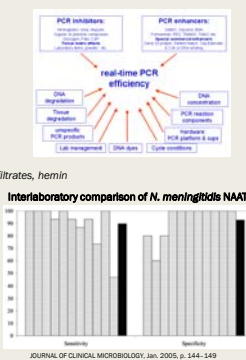
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CLINICAL MICROBIOLOGY REVIEWS, July 2010, p. 467-492

Detection methods

NAAT/PCR

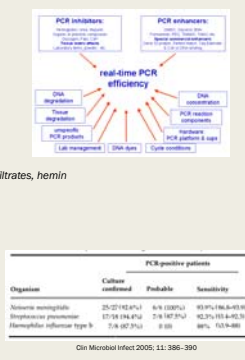
- Obstacles to NAAT
 - Few FDA-cleared options → LDT?
 - Complex to design
 - Inhibitors** - Elevated proteins, globulin, cellular infiltrates, hemin
 - Targets** - Binding affinity/strain diversity
 - Labs/lab variability
 - Lack of clinical samples to validate
 - Singleplex lacks broad applicability
 - Indistinguishable clinical presentation among bacterial (and sometimes viral) meningitis cases



Detection methods

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 - Lack of clinical samples to validate
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 - Indistinguishable clinical presentation among bacterial (and sometimes viral) meningitis cases
 - Multiplex?
 - More complex
 - Annealing temps., bacterial v. bacterial + viral?



VIRAL MENINGITIS

Clinical impact of viral meningitis

- "Aseptic meningitis"**
 - Mild/self-resolving to acute and life-threatening
 - ~35,000 hospitalizations/yr → 14/100,000
 - Critical role for Laboratory
 - Differentiate bacterial meningitis (less common/more severe) and severe viral etiologies
 - Management: Antiviral Rx? Supportive therapy?
- General risk factors**
 - Age
 - Immunocompromise (HIV or suppressive therapies)
 - Exposure
 - Outdoor activities, geographic location, season
 - Endemic areas for virus/vector
 - Outdoor activities
 - Community - pools, daycare

Epidemiology

- Prevalence**
 - Viral etiologies are the most common causes of meningitis (70-80%)
 - 50-70% of "aseptic meningitis" go without specific diagnosis/viral ID
 - Demographic most affected depends on specific virus
- Common agents...source of infection?**
 - Enteroviruses
 - > 10 million cases/yr in US → direct person-person spread (feces, saliva, fomites, water)
 - Arboviruses
 - ~100-200k infections/yr in US, ~1% severe symptoms → Arthropod-borne (mosquito, tick)
 - Herpesviruses
 - Recurrent meningitis in young adults, severe infection in compromised host → reactivation
 - Polyomaviruses
 - Exclusively compromised host, 1-8% of HIV patients pre-HAART → reactivation
- Vaccine Impact?**

Detection methods

- Culture
- Serology
- NAAT/PCR

Best method depends on...

specific virus, time from onset of symptoms, available tests, specimen

Detection methods

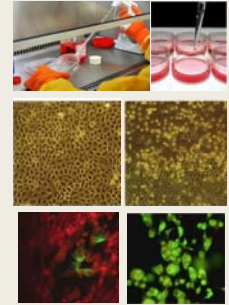
Virus	Effectiveness* of diagnosis by:							
	PCR		Serology [†]		Culture of specimens			
	Serum	CSF	Serum	CSF	Throat	Rectal	Blood	CSF
Arboviruses	+ ^d	+ ^d	++	++	-	-	++	+/-
WNV	+ ^d	+ ^d	++	++	-	-	++	+/-
Enteroviruses	+	++	+	-	+	++	+/-	++
Nonpoliovirus	+	++	+ ^d	-	+	++	-	-
Poliovirus	+	++	-	-	-	-	-	-
Herpesviruses								
CMV	+	++	+	++	+	-	+	Rare
EiV	+	+	++	++	+	-	+/-	+/-
HHV-6	+/-	+/-	+/-	-	-	-	+ ^d	+ ^d
HSV-1, and HSV-2	-	-	+/-	-	-	-	-	-
VZV	++	+	+ ^d	++	-	-	-	+
JCV	-	++	-	-	-	-	-	-

CLINICAL MICROBIOLOGY REVIEWS, Oct. 2004, p. 903-925

Detection methods

Culture

- Traditional "Gold standard"
 - Prepare monolayer of permissive cells (CAP MIC.61180)
 - Green monkey kidney, MRC-5, A549, MDCK
 - Seed to microwell plate or culture tube
 - Inoculate w/ specimen
 - Antibiotics - non-sterile site, lab contamination
 - Incubate depending on virus
 - Observe for CPE
 - On regular schedule, typically every 48-72
 - Cell lysis, vacuolization, syncytia, inclusion bodies
 - Semi-specific
 - Stain w/ panel of virus specific Abs
 - Final ID



Detection methods

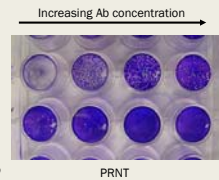
Culture - Disadvantages

- Poorly sensitive
 - CSF culture yield especially low, not typically recommended for diagnosis
 - HSV ~ 20% sensitive, EVs 30-35% sensitive, JC not cultivable using standard cell lines
 - Potentially due to presence of neutralizing Ab, low VL in CSF
 - Preventing uptake of viruses by host cells
- Extended TAT
 - Growth rate of some viruses e.g. VZV, can take up to 14-28 days (CAP MIC.61210)
 - Limits clinical utility for diagnosis
- Technical aspects of culture
 - Maintaining multiple cell lines
 - Contamination
 - Maintain proficiency of personnel

Detection methods

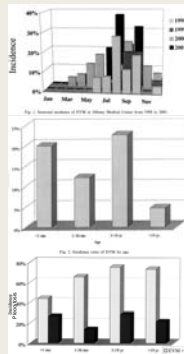
Culture - Advantages

- High specificity
 - Growth indicates viable virus, infectious etiology
- Broad inclusivity
 - Not limited by design of PCR target, availability of specific Ab
 - Discovery of novel viruses
- Increased specificity
 - Plaque-reduction neutralization assay (PRNT)
 - Add virus + dilutions of specific AB to each row
 - Determine 50% reduction from no ab control
 - Differentiate by w closely related viruses (e.g. flaviviridae)
- Epidemiology
- Antiviral resistance testing



Enteroviruses

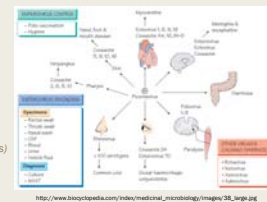
- Virus
 - Picornaviridae (enterovirus, echovirus, coxsackievirus)
 - Non-enveloped >60 serotypes
- Epidemiology
 - Summer-fall, primarily in children <5 yo
 - Transmitted in feces, saliva, environmental sources (water)
 - 80-90% of aseptic meningitis when etiology is found
- Symptoms
 - Largely asymptomatic or sub-clinical
 - Non-specific rash, fever, headache, URT symptoms, etc.
 - <5% Progress to more severe symptoms
 - Severe meningitis/encephalitis, Guillain-Barre syndrome
 - Cause severe sepsis syndrome in newborns
 - Pleocytosis not significantly different from controls in <1mo



Enteroviruses

Diagnostics

- Culture
 - CSF → Insensitive (20-60%), slow (5-8 days)
 - Resp/GI → non-specific (shed in stool, resp for 4-16 weeks)
- Serology
 - Non-specific, high rate of seropositivity
 - Many serotypes complicate diagnosis
- NAAT
 - Fast → <24 h
 - Comprehensive → 5'UTR target encompasses all serotypes
 - Sensitive → 10²-10³ copies/mL
 - Surrogate less invasive specimens...CSF vs. Blood?
 - Viremia in only 40-60% of CSF (+) patients → not rule out EV meningitis
 - EVs shed in resp. GI...NAATs may cross-react with rhinovirus



Xpert EV (2009)

- Xpert EV (Cepheid)**
 - Qualitative detection of >60 EV serotypes → 140 ul CSF, 2.5 h TAT
 - coxsackievirus, echovirus, and enterovirus



Xpert EV (2009)

- Performance**
 - Multicenter**
 - 199 prospective, 235 retrospective
 - Compared to LDTs and culture
 - Sensitivity: 95%, specificity: 100%**
 - Culture positive in only 35% of specimens

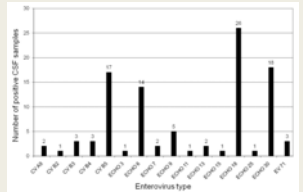
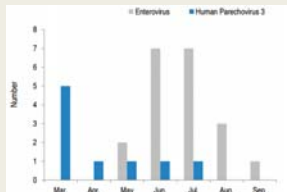


TABLE 1. Xpert EV assay for diagnosis of enteroviral meningitis (n = 434)

Site	No. of samples*				Sensitivity	Specificity	SP†
	Total	TP	FP	FN			
1	15	2	0	13	0	100	100
2	34	5	0	24	2	80	100
3	44	3	0	41	0	100	100
4	64	27	0	37	0	100	100
5	22	6	0	16	0	100	100
6	235	61	0	170	4	93.85	100
Total	434	107	0	321	6	94.69	100

Xpert EV (2009)

- Performance**
 - Multicenter**
 - 199 prospective, 235 retrospective
 - Compared to LDTs and culture
 - Sensitivity: 95%, specificity: 100%**
 - Culture positive in only 35% of specimens
- Drawbacks**
 - May suffer cross-reactivity w/ Rhinovirus
 - ...but this should not be in CSF
 - Does not include hPeV
 - 20-30% of "enteroviral aseptic meningitis"
 - Indistinguishable symptoms
 - Similar seasonality (summer-fall)



	Walters 2011	Sharpe 2013	Han 2013	Seo 2015
Location	USA	USA	Korea	Korea
EV	8.3%	14.0%	21.3%	7.5%
hPeV 3	2.4%	17.0%	6.5%	3.4%

Majority of patients < 5 years of age

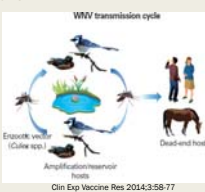
Xpert EV (2009)

- Clinical Impact**
 - EV leading cause of meningitis in children < 5 yoa
 - Rapid result, suggests non-bacterial meningitis
 - 50 children presenting with meningitis symptoms, EV positive
 - If EV positive result reported in < 24 h
 - Abx usage reduced by ~20 h
 - Hospital charges reduced by ~\$2,800

Variable	Time from Specimen Collection to Positive PCR Report			P
	<24 h (n = 17)	>24 h (n = 33)	Difference	
Antibiotic duration				
Mean (h)	22.6	42.2	19.6	0.006
Range	0-48	0-106		
Hospital charges				
Mean	\$3015	\$5831	\$2796	0.001
Range	\$801-\$5937	\$1406-\$16761		

Arboviruses

- Virus**
 - Diverse group of viruses transmitted by arthropods
 - Togaviridae - Eastern Equine, Western Equine, Venezuelan Equine
 - Bunyaviridae - La Crosse, Jamestown Canyon, California encephalitis
 - Flaviviridae - West Nile, St. Louis, Powassan, Tickborne encephalitis
- Epidemiology**
 - Largely driven by season/climate/vector range
 - Reservoir (amplifying host)
 - Likely underreported, >99% asymptomatic
- Symptoms**
 - Fever, rash → meningitis, encephalitis, flaccid paralysis
 - Attack rate, severity of symptoms highly variable
 - Hemorrhagic fever groups
 - Dengue, YFV, Rift Valley, Crimean-congo



Arboviruses - USA, 2014

2014. MMWR Morb Mortal Wkly Rep 2015;64:929-34

Characteristic	Virus type					
	West Nile (N = 2,295)	La Crosse (N = 80)	Jamestown Canyon (N = 11)	St. Louis encephalitis (N = 10)	Powassan (N = 8)	Eastern equine encephalitis (N = 8)
Age group (yrs)†						
<18	65 (3)	12 (9)	4 (6)	0 (0)	0 (0)	0 (0)
18-59	1,165 (51)	4 (3)	1 (6)	6 (60)	3 (38)	4 (50)
≥60	974 (44)	4 (3)	6 (53)	4 (40)	5 (62)	4 (50)
Sex						
Male	1,403 (64)	38 (48)	5 (45)	4 (40)	6 (75)	4 (50)
Female	892 (36)	42 (53)	6 (55)	6 (60)	2 (25)	4 (50)
Period of illness onset						
January-March	3 (1)	1 (1)	0 (0)	1 (10)	0 (0)	0 (0)
April-June	18 (3)	1 (1)	3 (27)	1 (10)	3 (38)	0 (0)
July-September	1,085 (48)	73 (91)	8 (73)	6 (60)	5 (62)	8 (100)
October-December	159 (7)	3 (4)	0 (0)	2 (20)	0 (0)	0 (0)
Clinical syndrome						
Non-neuroinvasive	858 (38)	4 (5)	5 (45)	4 (40)	1 (13)	0 (0)
Neuroinvasive	1,347 (61)	76 (95)	6 (55)	6 (60)	7 (88)	8 (100)
Encephalitis	620 (28)	63 (79)	3 (27)	4 (40)	5 (62)	6 (75)
Meningitis	945 (43)	12 (15)	2 (18)	1 (10)	2 (25)	1 (13)
Acute flaccid paralysis‡	132 (6)	0 (0)	1 (9)	1 (10)	0 (0)	1 (13)
Other neurologic	30 (1)	1 (1)	0 (0)	0 (0)	0 (0)	0 (0)
Outcome						
Hospitalization	1,589 (72)	79 (99)	7 (64)	10 (100)	8 (100)	8 (100)
Death	97 (4)	3 (4)	0 (0)	0 (0)	0 (0)	2 (25)

Arboviruses

Diagnostics

- **NAAT**
 - Blood → Viral replication/viremia precedes CNS involvement by 5-8 d
 - Non-specific fever/rash
 - Not detectable by time of CNS symptom onset
 - CSF → May be detected early in CNS symptoms
 - Still poor sensitivity – 60-70%

NAAT not recommended as primary test for arboviral meningitis
...but...
Specificity of NAAT is useful in epidemiologic studies

Arboviruses

Diagnostics

- **Serology**
 - Blood → not specific for CNS involvement
 - IgG persists for many years, if not life
 - IgM persists for 3-12 months
 - CSF → Preferred method
 - Intrathecal IgM indicates recent viral infection – likely cause
- **Method - IFA**
 - Infected host cells spotted to slide
 - Serum added, observe for fluorescence
 - Often multiple arbos on panel → same symptoms
 - Cross-reactive!
 - PRNT to definitively ID the specific arbovirus present

Herpesviruses

- **Virus**
 - HSV, VZV, CMV, HHV-6, EBV
- **Epidemiology**
 - Meningitis resulting from re-activation of latent infection
 - Competent – HSV, VZV
 - Compromised – VZV, CMV, HHV-6, EBV
- **Symptoms**
 - Clinical presentation consistent with meningitis
 - Fever, headache, photophobia
 - Severe/life-threatening
 - Immunosuppressed
 - Neonates

Herpesviruses

Site of latency

HSV-1
Herpesvirus (Type 1) Infection

HSV-2
Spinal Nerves

HSV

- **Clinical**
 - HSV-2 → Recurrent aseptic meningitis (Mollaret's)
 - Immune-competent, young adults
 - Self-resolving, optimal treatment not established
 - HSV-1 → Sporadic encephalitis
 - More common in compromised/HIV
 - Life-threatening, requires immediate treatment
 - 20% mortality, >95% of cases suffer long term neurologic defects

Simplexa HSV (2014)

- **Simplexa (Focus)**
 - Qualitative detection of HSV-1 and HSV-2 → 50 uL CSF, 1 h TAT

Simplexa HSV (2014)

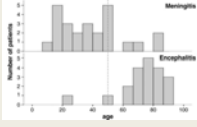
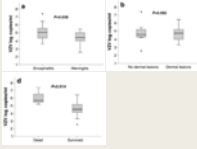
- **Performance**
 - Single center
 - 100 retrospective characterized CSF
 - Compared to LDts
 - Sensitivity: HSV-1: 100%, HSV-2: 100%
 - Specificity: HSV-1: 100%, HSV-2: 98.3%
 - 3 samples resulted as IND by Roche were negative by 3rd molecular comparator (Artus HSV)
- **Clinical Impact**
 - Competent adults
 - Recurrent HSV-2 meningitis common, self-limited no specific therapy
 - Compromised adults
 - HSV-1 severe encephalitis, require immediate treatment
 - Children
 - Important cause of neonatal meningitis → Assay off-label for blood, superficial (SEM screen)

Simplexa HSV-1/2 Direct	Roche ASR HSV-1/2 HSV Type		
	Positive	Negative	Indeterminate ^a
HSV-1			
Positive	11	0	1
Negative	0	85	3 ^d
HSV-2			
Positive	37	1 ^d	1
Negative	0	58	3 ^d

Herpesviruses

VZV

- Clinical**
 - Immunosuppressed → reactivation
 - cerebellar ataxia, meningitis or encephalitis
 - Rash is present in only 42% of patients with confirmed VZV CNS disease
- Serology**
 - CSF IgM
 - may be positive during asymptomatic reactivation or viremia episodes
 - Compare serum to CSF titer (?)
- NAAT**
 - Fast → <24 h
 - Sensitive → 10²-10³ copies/mL
 - Quant vs. Qual
 - Qualitative typically associated with causality
 - Quant prognostic?

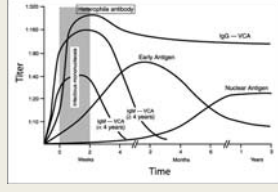



Diagnostic Microbiology and Infectious Disease 79 (2014) 174–177

Herpesviruses

EBV

- Virus**
 - ~90% seropositivity
 - Establishes latency in B-cells
 - Intermittent asymptomatic shedding in saliva
- Clinical**
 - Immunocompetent
 - Acute IM → adenopathy, malaise
 - CNS symptoms follow primary infection (pediatric/young adult)
 - Aseptic meningitis, encephalitis
 - Immunosuppressed → reactivation
 - 20-100% Burkitt, 40% Hodgkin, 10% DBC lymphomas
 - >95% of primary central nervous system lymphomas (PCNSLs)



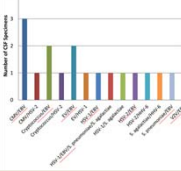
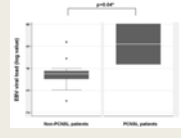
Serologic diagnosis: Heterophile Ab followed by VCA and EBNA

Herpesviruses

EBV

- Qualitative NAAT**
 - 75-100% sensitive
 - HIV-pos patients, histologically confirmed CNS lymphoma
 - 66-88% specific
 - Positive result NOT correlated with increased risk of PCNSLs
 - 25-70% of EBV-positive specimens also pos for another likely pathogen
 - 30-50% PPV
- Quantitative NAAT**
 - Can a threshold increase specificity?
 - Threshold of 10⁴ copies/mL → 96% specific
 - Not-standardized...lab-lab variability
 - Comparison of serum vs. CSF VL?

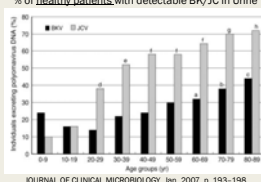
NAAT not extremely helpful, should not be used as sole means of diagnosis for CNS infections

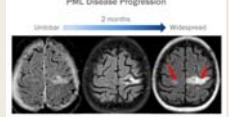
J. Clin. Microbiol. March 2016 vol. 54 no. 3 785-787

Polyomaviruses

- Virus**
 - JC, BK, circa 1970; nine others circa 2000-2005
- Epidemiology**
 - Seroprevalence 60-95%
 - JC/BK commonly shed in urine - asymptomatic
 - Compromised - severe focal organ disease
- Illness (JC)**
 - Progressive multifocal leukoencephalopathy (PML)
 - Destructive viral replication
 - demyelination of white matter in the brain
 - confusion, ataxia, paresis, and death if untreated
 - Immunosuppressed
 - AIDS, HSCT, therapy for MS (natalizumab)



JOURNAL OF CLINICAL MICROBIOLOGY, Jan. 2007, p. 193-198



PML Disease Progression
0 months 2 months 24 months

Polyomaviruses

JC

- Culture**
 - Non-cultivable in routinely used cell lines
- Serology**
 - Not useful for diagnosis, 60-90% seropositivity
 - Potential to screen/stratify risk for those considering certain MS therapies
- NAAT**
 - Fast → <24 h
 - Sensitivity is key!
 - 10¹-10² copies/mL (95% sensitive)
 - 10²-10³ copies/mL (75% sensitive)
 - Surrogate less invasive specimens...CSF vs. Blood, Urine?
 - Urine → detected in 40-70%, not correlated with PML
 - Blood → detected in 0.3-1%, none developed PML
 - No FDA-cleared assays, reference labs LDTs

Diagnostic criteria for PML

Definite (histopathologic) diagnosis

CSF confirmed PML

- 1. Evidence of PML findings consistent with PML, and
- 2. Evidence of JC's DNA in CSF

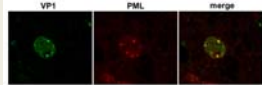
Tissue-confirmed PML

- 1. Evidence of PML neuropathology in brain tissue (biopsy or autopsy) with JC's DNA or genome detected by in situ hybridization

Presumptive (clinical) diagnosis

- 1. Evidence of typical clinical and MRI findings and
- 2. Blood biopsy and/or urine genome either not performed or JC's DNA not detected in CSF

Lancet Infect Dis. 2009 October ; 9(10): 625-636



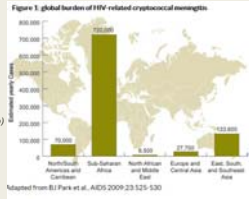
The American Journal of Pathology, Vol. 180, No. 3, March 2012

FUNGAL MENINGITIS

Clinical impact of viral meningitis

Epidemiology

- Historically rare compared to viral/bacterial
 - Increasing prevalence with increasing immunosuppressed population
 - HIV/AIDS, hematologic malignancies, direct spinal surgeries/therapies
 - Pathogens
 - Cryptococcus - >90% of fungal CNS infections
 - Dimorphic fungi - Coccidioides, Blastomyces
 - Filamentous fungi - "dematiaceous molds"



General risk factors

- Age
- Immunocompromise (HIV -100-fold higher incidence of Crypto)
- Exposure
 - Geographic location
 - Medical procedures

Detection methods

- Culture
- Antigen
- Serology
- NAAT/PCR

Best method depends on...
specific fungus, available tests, specimen

Detection methods

Culture

- Direct exam of specimen
 - Low yield, not routinely performed
- Plating
 - > 2mL → centrifuge, plate entire pellet
 - Supernatant can be used for serologic tests
 - < 2mL → plate entire volume to fungal culture media
- Sterile source
 - Critical to differentiate contamination from true infection
 - Do NOT streak inoculum → consider growth only at inoculation site

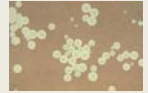


CSF culture for Fungi is typically low yield, augment with second approach when available

Detection methods

Cryptococcus

- Direct exam of specimen
 - India ink smear → Poor sensitivity
- Culture
 - Highly dependent on specimen volume/abx exposure
- Cryptococcal Ag
 - Latex agglutination - capsular polysaccharide
 - Most sensitive method for diagnosis of Cryptococcal meningitis
 - CSF
 - Serum
 - Persists after resolution of symptoms
 - Positive in culture-negative/NAAT negative samples
 - Not a test of cure!
 - Can cross-react with other capsulated yeast → Trichosporon, Rhodotulura



Time of findings	Leukocyte (10 ⁶ /liter)	No. of positive fungal cultures (%)	Cryptococcal titers
Before therapy	112.4 ± 228.5	20 (0.00)	1.049.1 ± 4.1
After therapy	12.9 ± 11.2	0 (0.00)	238.4 ± 4.4

JOURNAL OF CLINICAL MICROBIOLOGY, June 2005, p. 2989-2990

Detection methods

Cryptococcus

- Direct exam of specimen
 - India ink smear → Poor sensitivity
- Culture
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JOURNAL OF CLINICAL MICROBIOLOGY, June 2005, p. 2989-2990

No. of patients	Host condition	No. positive by CSF Ag (%)	No. positive by CSF culture (%)	No. positive by India ink (%)	No. positive by serum Ag (%)
401	AIDS	307/333 (92.2)	380/401 (94.7)	302/375 (80.5)	181/183 (98.9)
331	HIV negative, immunocompromised	214/219 (97.7)	251/282 (89)	139/231 (60.2)	129/148 (87.2)
116	Immunocompetent	72/78 (92.3)	91/102 (89.2)	89/116 (76.7)	181/183 (98.9)
848	Overall total	593/630 (94.1)	722/785 (92)	430/722 (59.6)	310/311 (93.6)

JOURNAL OF CLINICAL MICROBIOLOGY, Nov. 2005, p. 5828-5829

Detection methods

Dimorphs

- Coccidioides, Blastomyces, Histoplasma
 - CNS infection secondary to resp. infection
 - More common in compromised host
 - Histo - 5-10% of disseminated infections
 - Blasto - <5% of disseminated infections
 - Coccidioides - 30-50% of disseminated infections
- Culture - Poor sensitivity from CSF, long TAT. **HANDLE WITH CARE!**
- Serology - Poor sensitivity in compromised host, high seroprevalence
- Antigen tests
 - Blasto/Histo
 - Urine antigen test >90% sensitive for disseminated disease
 - Cross-reactive (Blasto, Histo, Paracoccidioides)



Detection methods

Dematiaceous

- "Dark walled" fungi, contain melanin
 - Saprophytic → soil/decaying plant material
 - Filamentous and yeast-like organisms
 - *Cladophialophora bantiana*
 - Inhalation → neuroinvasive/parenchymal growth
 - *Exophiala* → yeastlike
 - Traumatic introduction → injury, lines, ports
 - *Scedosporium*
 - Inhalation → neurotropic
- Opportunistic skin/soft tissue and respiratory pathogens
 - **Triplasm for CNS**
 - **Affect young/healthy individuals**



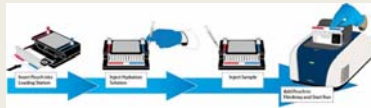
MOLECULAR DIAGNOSTICS

Syndromic panels

"Syndromic panel"

FilmArray ME (BioFire, 2015)

- Simultaneous detection of 14 targets → 200 uL CSF, 1 h TAT
 - Bacteria → *E. coli K1*, *H. influenzae*, *L. monocytogenes*, *N. meningitidis*, *S. agalactiae*, *S. pneumoniae*
 - Viruses → CMV, VZV, HSV-1, HSV-2, HHV-6, Enterovirus, Human Parainfluenza
 - Fungi → *Cryptococcus neoformans/gattii*



FilmArray ME (2015)

■ Performance

- Single center
 - 174 retrospective characterized CSF
 - Compared to LDTs (viral), culture (bacterial)
 - Discordant results tested with third LDT NAAT

TABLE 1 Distribution of organisms identified by conventional methods and the FilmArray meningitis/encephalitis (FA ME) panel

Organism identification ^a	Conventional detection, no.	FA ME panel detection, no.	Baseline agreement, no.	Resolution result, no. ^b			Sensitivity, % (95% CI) ^c	Specificity, % (95% CI) ^c
				FA+/R+	FA+/R-	FA-/R+		
Bacteria								
<i>H. influenzae</i>	4	3	4	1	0	0	100 (47.8-100)	100 (97.4-100)
<i>S. pneumoniae</i>	3	6	3	2	1	0	100 (47.8-100)	99.3 (96.1-100)
<i>S. agalactiae</i>	1	3	1	2	2	1	66.7 (16.4-99.2)	98.6 (93.0-99.8)
<i>Escherichia coli</i>	1	1	1	NA ^d	NA	NA	100 (2.5-100)	100 (97.5-100)
<i>Listeria monocytogenes</i>	0	0	1	NA	NA	NA	NA	100 (97.5-100)
<i>Neisseria meningitidis</i>	1	1	1	NA	NA	NA	100 (2.5-100)	100 (97.5-100)
Bacteria not in the FA ME panel ^e	7	0	7	NA	NA	NA	NA	NA

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				FA+/R+	FA+/R-	FA-/R+		
Viruses								
EV	37	37	36	1	0	1	97.4 (86.2-99.9)	100 (98.2-100)
HSV-1	12	13	13	0	2	1	92.9 (66.1-99.8)	98.0 (89.1-99.9)
HSV-2	29	29	29	NA	NA	NA	100 (98.1-100)	100 (92.4-100)
HHV-6	13	18	12	6	0	1	94.7 (74.0-99.9)	100 (92.6-100)
VZV	32	32	32	NA	NA	NA	100 (99.1-100)	100 (79.4-100)
CMV	7	4	4	0	0	3	37.3 (16.4-60.1)	100 (91.6-100)
EBV	13	25	11	5	9	1	94.1 (71.3-99.9)	84.2 (72.1-92.5)
PV	0	1	0	1	0	0	100 (2.5-100)	100 (92.5-100)
Yeast								
<i>C. neoformans/gattii</i>	14	9	8	1	0	0 ^f	64.3 (35.1-87.2)	NA
Total	174	186	161	19	14	0	92.8 (88.2-96.0)	92.8 (88.2-96.0)

FilmArray ME (2015)

■ Potential pitfalls

- *Cryptococcus*
 - 64% sensitive compared with CrAG
 - All discordants were negative by alternative NAAT and culture
 - CrAG more sensitive than NAAT? AG persist?



FilmArray ME (2015)

- Potential pitfalls**
 - Cryptococcus**
 - 64% sensitive compared with CrAG
 - All discordants were negative by alternative NAAT and culture
 - CrAG more sensitive than NAAT? AG persist at low level!!!!

JOURNAL OF CLINICAL MICROBIOLOGY, June 2005, p. 2989-2990

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 - Cryptococcus**
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 - All discordants were negative by alternative NAAT and culture
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 - EBV**
 - 84% specific, EBV in 14/20 (70%) of "mixed infections"
 - Lymphocytic infiltrates → latent virus?
 - CMV? HHV-6? → clinical significance?

JOURNAL OF CLINICAL MICROBIOLOGY, June 2005, p. 2989-2990

FilmArray ME (2015)

- Potential pitfalls**
 - Cryptococcus**
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 - EBV**
 - 84% specific, EBV in 14/20 (70%) of "mixed infections"
 - Lymphocytic infiltrates → latent virus?
 - CMV? HHV-6? → clinical significance?
 - S. pneumoniae**
 - 99% (1544/1556) specific but....
 - Only 5/12 FP confirmed by alternative NAAT
 - 9 TP, 3 FP → PPV 75%
 - Data from package insert

With low prevalence population, specificity is key!!!!!!

J. Clin. Microbiol., March 2016 vol. 54 no. 3 785-787

FilmArray ME (2015)

- Utilization**
 - Pediatrics**
 - Rapid, effective method to determine cause of symptoms
 - Enterovirus/HPeV vs. HSV vs. bacterial
 - Adult outpatient, acute onset**
 - Rapid method for HSV, but more \$\$\$ than Simplex HSV
 - Consider clinical picture (severe symptoms, elderly), do results fit?
 - S. pneumoniae? Other herpesviruses?
 - Compromised patient**
 - Rapid, but is it comprehensive?
 - 14/110 (13%) positive bacterial cultures were on-panel targets
 - Analytical vs. clinical specificity for herpesviruses
 - Inpatient with hardware**
 - No! Common bugs not on panel (CoNS, P. aeruginosa, Enterococcus, Acinetobacter)

ID	# culture	% culture
S. epidermidis	59	35.5%
CoNS	5	4.5%
Corynebacterium	1	0.9%
P. aeruginosa	4	3.6%
Staphylococcus	2	1.8%
Micrococcus	1	0.9%
S. pneumoniae	4	3.6%
N. meningitidis	3	2.7%
E. coli	4	3.6%
S. marcescens	3	2.7%
Enterococcus	6	5.5%
P. aeruginosa	7	6.4%
Enterobacter spp.	4	3.6%
S. agalactiae	2	1.8%
S. pyogenes	1	0.9%
Acinetobacter	3	2.7%
W. meningitidis	5	4.5%
S. aureus	4	3.6%
P. mirabilis	1	0.9%
Helicobacter pylori	5	4.5%
Candida spp.	2	1.8%
E. pneumoniae	4	3.6%
Total	110	

Conclusion

- Meningitis remains a common, potentially serious condition**
 - Critical to get result to clinician as fast as possible**
 - Major impact on care and management (antibiotics, antivirals? supportive care?)
 - No single approach is sufficient to detect all causes
 - In choosing orderable test consider**
 - Symptoms
 - Patient population
 - Current and previous infections/anatomic sites
 - Geographic locale
 - Molecular tests are typically the most sensitive method for diagnosis however...**
 - Few FDA-cleared options
 - "Only find what you are looking for" – potential for false sense of security
 - Must always be accompanied by culture

THE END

Are we still awake?