

# Updates to CLSI M100



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The presenter states no conflict of interest and has no financial relationship to disclose relevant to the content of this presentation.

# OUTLINE

I. General acclimation to M100 document

II. Objectives of webinar

Describe significant changes relevant to pre-existing antimicrobial susceptibility breakpoints...

Describe significant changes relevant to antimicrobial susceptibility testing methodology...

Identify (new) organism/antimicrobial combinations for which susceptibility breakpoints now exist...

*as outlined in the CLSI M100-S29 document.*



# The Small Roman Numerals



# WHAT'S IN THERE?

- Overview of changes
- Processes for establishing breakpoints, QC ranges
- CLSI reference vs. commercial methods  
CLSI reference vs. FDA breakpoints
- Breakpoint additions/revisions since 2010  
ECV additions/revisions since 2015

# BREAKPOINT ADDITIONS FOR 2019



Investigational cefiderocol BMD for:

*Enterobacteriaceae*

*Pseudomonas aeruginosa*

*Acinetobacter* spp.

*Stenotrophomonas maltophilia*

Requires iron-depleted CAMHB

- Meropenem-vaborbactam  
*Enterobacteriaceae* (DD, BMD)
- Azithromycin  
*Neisseria gonorrhoeae* (MIC, S only)





# Instructions for Use of Tables



# INSTRUCTIONS FOR USE I

- Selecting antimicrobial agents for testing, reporting
- Breakpoint and interpretive category definitions
- Reporting results

Organisms excluded from Table 2 (CLSI M45)

Reporting concentrations (report to hundredths)

$\leq 0.125 \mu\text{g/mL}$  reported as  $\leq 0.12 \mu\text{g/mL}$

- Therapy-related comments

# INSTRUCTIONS FOR USE II

- Testing of repeat isolates (as little as 3-4 days)

3° cepheids      *Klebsiella aerogenes*  
*Citrobacter* spp.  
*Serratia* spp.

All agents      *Pseudomonas aeruginosa*

Fluoroquinolones      Staphylococci

- Supplemental (D-test), screening (vancomycin agar screen), **surrogate agent** (cefoxitin), equivalent agent (amp predict amox for *Haemophilus*) tests

## Surrogate Agent Tests

Surrogate Agent	Organisms	Test Description	Results	Table Location
Cefazolin	<ul style="list-style-type: none"> <li>• <i>E. coli</i></li> <li>• <i>Klebsiella pneumoniae</i></li> <li>• <i>P. mirabilis</i></li> </ul>	Broth microdilution or disk diffusion	<p>When used for therapy of uncomplicated UTIs, predicts results for the following oral antimicrobial agents: cefaclor, cefdinir, cefpodoxime, cefprozil, cefuroxime, cephalexin, and loracarbef</p> <p>Cefazolin as a surrogate may overcall resistance to cefdinir, cefpodoxime, and cefuroxime. If cefazolin tests resistant, test these drugs individually if needed for therapy.</p>	1A, 2A
Cefoxitin	<ul style="list-style-type: none"> <li>• <i>S. aureus</i></li> <li>• <i>S. lugdunensis</i></li> <li>• <i>S. epidermidis</i></li> <li>• Other <i>Staphylococcus</i> spp. (excluding <i>S. pseudintermedius</i> and <i>S. schleiferi</i>)</li> </ul>	Broth microdilution ( <i>S. aureus</i> and <i>S. lugdunensis</i> only) or disk diffusion	<p>Predicts results for <i>mecA</i>-mediated oxacillin resistance</p> <p><b>NOTE: For <i>Staphylococcus</i> spp. other than <i>S. aureus</i>, <i>S. lugdunensis</i>, <i>S. epidermidis</i>, <i>S. pseudintermedius</i>, and <i>S. schleiferi</i>, oxacillin MIC breakpoints may overcall resistance. Isolates for which the oxacillin MICs are 0.5–2 µg/mL have been shown to be <i>mecA</i> positive and <i>mecA</i> negative. Isolates from serious infections with MICs in this range may be tested for <i>mecA</i> or for PBP2a.</b></p>	1A, 2C
Oxacillin	<ul style="list-style-type: none"> <li>• <i>S. pneumoniae</i></li> </ul>	Disk diffusion	Predicts penicillin susceptibility if oxacillin zone is ≥ 20 mm. If oxacillin zone is ≤ 19 mm, penicillin MIC must be done.	1B, 2G
Pefloxacin	<ul style="list-style-type: none"> <li>• <i>Salmonella</i> spp.</li> </ul>	Disk diffusion	Predicts reduced susceptibility to ciprofloxacin	2A
Colistin	<ul style="list-style-type: none"> <li>• <i>Enterobacteriaceae</i></li> <li>• <i>P. aeruginosa</i>*</li> <li>• <i>A. baumannii</i> complex</li> </ul>	Broth microdilution	MICs obtained from testing colistin predict MICs for polymyxin B.	2B-1, 2B-2, Appendix G



Table 1



# TABLE 1A

Table 1A  
Suggested Nonfastidious Groupings  
M02 and M07

**Table 1A. Suggested Groupings of Antimicrobial Agents Approved by the US Food and Drug Administration for Clinical Use That Should Be Considered for Testing and Reporting on Nonfastidious Organisms by Microbiology Laboratories in the United States**

GROUP A PRIMARY TEST AND REPORT	<i>Enterobacteriaceae</i>	<i>Pseudomonas aeruginosa</i>	<i>Staphylococcus</i> spp.	<i>Enterococcus</i> spp. <sup>m</sup>
	Ampicillin <sup>c</sup>	Cefazidime	Azithromycin <sup>b</sup> or clarithromycin <sup>b</sup> or erythromycin <sup>b</sup>	Ampicillin <sup>n</sup> Penicillin <sup>o</sup>
	Cefazolin <sup>d</sup>	Gentamicin Tobramycin	Clindamycin <sup>b</sup>	
	Gentamicin <sup>c</sup> Tobramycin <sup>c</sup>	Piperacillin-tazobactam	Oxacillin <sup>k,t,t,t</sup> & Cefoxitin <sup>k,t</sup> (surrogate test for oxacillin)	
		Penicillin <sup>l</sup>		
		Trimethoprim- sulfamethoxazole		

# TABLE 1A

Table 1A  
Suggested Nonfastidious Groupings  
M02 and M07

Table 1A. (Continued)

**“Warning”:** The following antimicrobial agents that are included in this document should not be routinely reported for bacteria isolated from CSF. These antimicrobial agents are not the drugs of choice and may not be effective for treating CSF infections caused by these organisms (ie, the bacteria included in Tables 2A through 2J):

- Agents administered by oral route only
- 1st- and 2nd-generation cephalosporins and cephamycins
- Clindamycin
- Macrolides
- Tetracyclines
- Fluoroquinolones

## Footnotes

### General

- a. Organisms that are susceptible to tetracycline are also considered susceptible to doxycycline and minocycline. However, some organisms that are intermediate or resistant to tetracycline may be susceptible to doxycycline, minocycline, or both.
- b. Not routinely reported on organisms isolated from the urinary tract.

# NON-FASTIDIOUS GROUPINGS

- Group A Primary test and report
- Group B Optional primary test, report selectively
- Group C Supplemental report selectively
- Group U Supplemental for urine only

# NON-FASTIDIOUS GROUPINGS

These exist for:

*Enterobacteriaceae*

*Pseudomonas aeruginosa* (no group C or group U)

*Acinetobacter* spp. (no group C)

*Burkholderia cepacia* complex (no group U)

*Stenotrophomonas maltophilia* (no group U)

Other non-*Enterobacteriaceae*

*Staphylococcus* spp.

*Enterococcus* spp.

# CHANGES TO TABLE 1A

- Added meropenem-vaborbactam as group B testing agent for *Enterobacteriaceae*
- Clarified staphylococci that can be subject to oxacillin broth microdilution (footnote to Table)

*S. aureus*

*S. lugdunensis*

Other staphylococci

EXCEPT

*epidermidis*

*pseudintermedius*

*schleiferi*

# TABLE 1B

Table 1B  
Suggested Fastidious Groupings  
M02 and M07

**Table 1B. Suggested Groupings of Antimicrobial Agents Approved by the US Food and Drug Administration for Clinical Use That Should Be Considered for Testing and Reporting on Fastidious Organisms by Microbiology Laboratories in the United States**

GROUP A PRIMARY TEST AND REPORT	<i>Haemophilus influenzae</i> <sup>d</sup> and <i>Haemophilus parainfluenzae</i>	<i>Neisseria gonorrhoeae</i> <sup>l</sup>	<i>Streptococcus pneumoniae</i> <sup>l</sup>	<i>Streptococcus</i> spp. β-Hemolytic Group <sup>p</sup>	<i>Streptococcus</i> spp. Viridans Group <sup>p</sup>
	Ampicillin <sup>d,t</sup>	Azithromycin <sup>nt</sup>	Erythromycin <sup>ac</sup>	Clindamycin <sup>c,p</sup>	Ampicillin <sup>m,t</sup> Penicillin <sup>m,t</sup>
	Ceftriaxone <sup>t</sup> Cefixime <sup>t</sup>				
	Ciprofloxacin <sup>t</sup>	Penicillin <sup>k</sup> (oxacillin disk)	Erythromycin <sup>a,co</sup>		
	Tetracycline <sup>b,t</sup>	Trimethoprim- sulfamethoxazole	Penicillin <sup>nt</sup> or ampicillin <sup>nl</sup>		

# TABLE 1B

Table 1B  
Suggested Fastidious Groupings  
M02 and M07

**Table 1B. Suggested Groupings of Antimicrobial Agents Approved by the US Food and Drug Administration for Clinical Use That Should Be Considered for Testing and Reporting on Fastidious Organisms by Microbiology Laboratories in the United States**

GROUP A PRIMARY TEST AND REPORT	<i>Haemophilus influenzae</i> <sup>d</sup> and <i>Haemophilus parainfluenzae</i>	<i>Neisseria gonorrhoeae</i> <sup>l</sup>	<i>Streptococcus pneumoniae</i> <sup>l</sup>	<i>Streptococcus</i> spp. β-Hemolytic Group <sup>p</sup>	<i>Streptococcus</i> spp. Viridans Group <sup>p</sup>
	Ampicillin <sup>d,t</sup>	Azithromycin <sup>st</sup>	Erythromycin <sup>ac</sup>	Clindamycin <sup>c,p</sup>	Ampicillin <sup>m,t</sup> Penicillin <sup>m,t</sup>
	Ceftriaxone <sup>t</sup> Cefixime <sup>t</sup>				
	Ciprofloxacin <sup>t</sup>	Penicillin <sup>k</sup> (oxacillin disk)	Erythromycin <sup>a,co</sup>		
	Tetracycline <sup>b,t</sup>	Trimethoprim- sulfamethoxazole	Penicillin <sup>nt</sup> or ampicillin <sup>nl</sup>		

- o. **Rx:** Recommendations for intrapartum prophylaxis for group B streptococci are penicillin or ampicillin. Although cefazolin is recommended for penicillin-allergic women at low risk for anaphylaxis, those at high risk for anaphylaxis may receive clindamycin. Group B streptococci are susceptible to ampicillin, penicillin, and cefazolin, but may be resistant to erythromycin and dindamycin. When group B *Streptococcus* is isolated from a pregnant woman with severe penicillin allergy (high risk for anaphylaxis), erythromycin and dindamycin (including inducible clindamycin resistance) should be tested, and only clindamycin should be reported. See Table 3G.

# FASTIDIOUS GROUPINGS

- Group A Primary test and report
- Group B Optional primary test, report selectively
- Group C Supplemental report selectively

# FASTIDIOUS GROUPINGS

These exist for:

*Haemophilus influenzae*

*Haemophilus parainfluenzae*

*Neisseria gonorrhoeae* (no group B or no group C)

*Streptococcus pneumoniae*

$\beta$ -hemolytic *Streptococcus*

viridans group *Streptococcus*

# CHANGES TO TABLE 1B

- Added azithromycin as group A testing agent for *Neisseria gonorrhoeae*
- Susceptibility and resistance to dirithromycin (as well as azithromycin and clarithromycin) can be predicted by testing erythromycin

*Streptococcus pneumoniae*

$\beta$ -hemolytic *Streptococcus*

viridans group *Streptococcus*

# THERE'S AN ANAEROBE TABLE

Table 1C  
Suggested Anaerobe Groupings  
M11

Table 1C. Suggested Groupings of Antimicrobial Agents Approved by the US Food and Drug Administration for Clinical Use That Should Be Considered for Testing and Reporting on Anaerobic Organisms by Microbiology Laboratories in the United States

	Gram-Negative Anaerobes	Gram-Positive Anaerobes <sup>a</sup>
GROUP A PRIMARY TEST AND REPORT	Amoxicillin-clavulanate Ampicillin-sulbactam Piperacillin-tazobactam	Ampicillin <sup>b</sup> Penicillin <sup>b</sup> Amoxicillin-clavulanate Ampicillin-sulbactam Piperacillin-tazobactam
	Clindamycin	Clindamycin
	Doripenem Ertapenem Imipenem Meropenem	Doripenem Ertapenem Imipenem Meropenem
	Metronidazole	Metronidazole
GROUP C SUPPLEMENTAL REPORT SELECTIVELY	Penicillin <sup>b</sup> Ampicillin <sup>b</sup>	
	Cefotetan Cefoxitin	Cefotetan Cefoxitin
	Ceftizoxime Ceftriaxone	Ceftizoxime Ceftriaxone
	Chloramphenicol	
	Moxifloxacin	Moxifloxacin Tetracycline

“Testing may not be necessary for isolates associated with polymicrobial anaerobic infections.”  
Test only isolate most likely to be resistant; *Bacteroides* spp., *Parabacteroides* spp.



## Table 2



# TABLE 2

Table 2B-1  
*Pseudomonas aeruginosa*  
M02 and M07

Table 2B-1. Zone Diameter and MIC Breakpoints for *Pseudomonas aeruginosa*

<b>Testing Conditions</b>  <b>Medium:</b> Disk diffusion: MHA Broth dilution: CAMHB; <b>For cefiderocol, special media is required for testing. See comment (13).</b> Agar dilution: MHA <b>Inoculum:</b> Broth culture method or colony suspension, equivalent to a 0.5 McFarland standard <b>Incubation:</b> 35°C ± 2°C; ambient air Disk diffusion: 16–18 hours Dilution methods: 16–20 hours	<b>Routine QC Recommendations</b> (see Tables 4A-1 and 5A-1 for acceptable QC ranges)  <i>Pseudomonas aeruginosa</i> ATCC <sup>®</sup> 27853  Refer to Tables 4A-2 and 5A-2 to select strains for routine QC of $\beta$ -lactam combination agents.  When a commercial test system is used for susceptibility testing, refer to the manufacturer's instructions for QC test recommendations and QC ranges.
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\* ATCC<sup>®</sup> is a registered trademark of the American Type Culture Collection.

## General Comments

- (1) For disk diffusion, test a maximum of 12 disks on a 150-mm plate and no more than 6 disks on a 100-mm plate; disks should be placed no less than 24 mm apart, center to center (see M02,<sup>1</sup> Subchapter 3.6). Each zone diameter should be clearly measurable; overlapping zones prevent accurate measurement. Measure the diameter of the zones of complete inhibition (as judged by the unaided eye), including the diameter of the disk. Hold the Petri plate a few inches above a black background illuminated with reflected light. The zone margin should be considered the area showing no obvious, visible growth that can be detected with the unaided eye. Ignore faint growth of tiny colonies that can be detected only with a magnifying lens at the edge of the zone of inhibited growth.
- (2) The susceptibility of *P. aeruginosa* isolated from patients with cystic fibrosis can be reliably determined by disk diffusion or dilution methods but may need extended incubation for up to 24 hours before reporting as susceptible.
- (3) *P. aeruginosa* may develop resistance during prolonged therapy with all antimicrobial agents. Therefore, isolates that are initially susceptible may become resistant within 3 to 4 days after initiation of therapy. Testing of repeat isolates may be warranted.

These comments are found within the Table.

# TABLE 2

Table 2B-1. *Pseudomonas aeruginosa* (Continued)

Test/Report Group	Antimicrobial Agent	Disk Content	Interpretive Categories and Zone Diameter Breakpoints, nearest whole mm			Interpretive Categories and MIC Breakpoints, µg/mL			Comments
			S	I	R	S	I	R	
<b>PENICILLINS</b>									
O	Piperacillin	100 µg	≥ 21	15–20	≤ 14	≤ 16	32–64	≥ 128	(5) Breakpoints for piperacillin (alone or with tazobactam) are based on a piperacillin dosage regimen of at least 3 g <b>administered</b> every 6 h.
<b>β-LACTAM COMBINATION AGENTS</b>									
A	Piperacillin-tazobactam	100/10 µg	≥ 21	15–20	≤ 14	≤ 16/4	32/4–64/4	≥ 128/4	(6) Breakpoints for piperacillin (alone or with tazobactam) are based on a piperacillin dosage regimen of at least 3 g <b>administered</b> every 6 h.
B	Ceftazidime-avibactam	30/20 µg	≥ 21	–	≤ 20	≤ 4/4	–	≥ 16/4	(7) Breakpoints are based on a dosage regimen of 2.5 g (2 g ceftazidime + 0.5 g avibactam) <b>administered</b> every 8 h over 2 h.
B	Ceftolozane-tazobactam	30/10 µg	≥ 21	17–20	≤ 16	≤ 4/4	8/4	≥ 16/4	(8) Breakpoints are based on a dosage regimen of 1.5 g <b>administered</b> every 8 h.
O	Ticarcillin-clavulanate	75/10 µg	≥ 24	16–23	≤ 15	≤ 16/2	32/2–64/2	≥ 128/2	(9) Breakpoints for ticarcillin (alone or with clavulanate) are based on a ticarcillin dosage regimen of at least 3 g <b>administered</b> every 6 h.
<b>CEPHEMS (PARENTERAL) (Including cephalosporins I, II, III, and IV. Please refer to Glossary I.)</b>									
A	Ceftazidime	30 µg	≥ 18	15–17	≤ 14	≤ 8	16	≥ 32	(10) Breakpoints are based on a dosage regimen of 1 g <b>administered</b> every 6 h or 2 g <b>administered</b> every 8 h.
B	Cefepime	30 µg	≥ 18	15–17	≤ 14	≤ 8	16	≥ 32	(11) Breakpoints are based on a dosage regimen of 1 g <b>administered</b> every 8 h or 2 g <b>administered</b> every 12 h.

# TABLE 2

Inv.	Cefiderocol	–	–	–	–	≤4	8	≥16	<p>(12) Breakpoints are based on a dosage regimen of 2 g every 8 h administered over 3 h.</p> <p>(13) Testing cefiderocol requires iron-depleted CAMHB. Chelation is used for iron depletion, which also removes other cations (ie, calcium, magnesium, and zinc). Following this process, cations are added back to concentrations of calcium 20–25 mg/L, magnesium 10–12.5 mg/L, and zinc 0.5–1.0 mg/L.</p>
<p style="color: red; font-size: 1.2em;">Comment 13 from two slides ago</p>									
<b>MONOBACTAMS</b>									
B	Aztreonam	30 µg	≥ 22	16–21	≤ 15	≤ 8	16	≥ 32	<p>(14) Breakpoints are based on a dosage regimen of 1 g <b>administered</b> every 6 h or 2 g <b>administered</b> every 8 h.</p>

Table 2B-1  
*Pseudomonas aeruginosa*  
M02 and M07

# TABLE 2

## These exist for:

*Enterobacteriaceae*

*Pseudomonas aeruginosa*

*Acinetobacter* spp.

*Burkholderia cepacia* complex

*Stenotrophomonas maltophilia*

Other non-*Enterobacteriaceae*

*Staphylococcus* spp.

*Enterococcus* spp.

*Haemophilus influenzae*

*Haemophilus parainfluenzae*

*Neisseria gonorrhoeae*

*Streptococcus pneumoniae*

$\beta$ -hemolytic *Streptococcus*

viridans group *Streptococcus*

*Neisseria meningitidis*

Anaerobes

# CAVEATS

Table 2B-5. Other Non-*Enterobacteriaceae* (Continued)

Test/Report Group	Antimicrobial Agent	Disk Content	Interpretive Categories and Zone Diameter Breakpoints, nearest whole mm			Interpretive Categories and MIC Breakpoints, µg/mL		
			S	I	R	S	I	R
<b>PENICILLINS</b>								
O	Piperacillin	–	–	–	–	≤16	32–64	≥128
<b>β-LACTAM COMBINATION AGENTS</b>								
B	Piperacillin-tazobactam	–	–	–	–	≤16/4	32/4–64/4	≥128/4
O	Ticarcillin-clavulanate	–	–	–	–	≤16/2	32/2–64/2	≥128/2
<b>CEPHEMS (PARENTERAL) (Including cephalosporins I, II, III, and IV. Please refer to Glossary I.)</b>								
A	Ceftazidime	–	–	–	–	≤8	16	≥32
B	Cefepime	–	–	–	–	≤8	16	≥32
C	Cefotaxime	–	–	–	–	≤8	16–32	≥64
C	Ceftriaxone	–	–	–	–	≤8	16–32	≥64
O	Cefoperazone	–	–	–	–	≤16	32	≥64
O	Ceftizoxime	–	–	–	–	≤8	16–32	≥64
O	Moxalactam	–	–	–	–	≤8	16–32	≥64
<b>MONOBACTAMS</b>								
B	Aztreonam	–	–	–	–	≤8	16	≥32
<b>CARBAPENEMS</b>								
B	Imipenem	–	–	–	–	≤4	8	≥16
B	Meropenem	–	–	–	–	≤4	8	≥16
<b>AMINOGLYCOSIDES</b>								
A	Gentamicin	–	–	–	–	≤4	8	≥16
A	Tobramycin	–	–	–	–	≤4	8	≥16
B	Amikacin	–	–	–	–	≤16	32	≥64
O	Netilmicin	–	–	–	–	≤8	16	≥32
<b>TETRACYCLINES</b>								
(3) Organisms that are susceptible to tetracycline are also considered susceptible to doxycycline and minocycline. However, some organisms may be susceptible to doxycycline, minocycline, or both.								
U	Tetracycline	–	–	–	–	≤4	8	≥16
O	Doxycycline	–	–	–	–	≤4	8	≥16
O	Minocycline	–	–	–	–	≤4	8	≥16
<b>FLUOROQUINOLONES</b>								
B	Ciprofloxacin	–	–	–	–	≤1	2	≥4
B	Levofloxacin	–	–	–	–	≤1	2	≥4



3rd Edition

## M45

Methods for Antimicrobial Dilution and Disk Susceptibility Testing of Infrequently Isolated or Fastidious Bacteria

This guideline informs clinical, public health, and research laboratories on susceptibility testing of infrequently isolated or fastidious bacteria that are not included in CLSI documents M07, M07, or M100. Antimicrobial agent selection, test interpretation, and quality control are addressed.

A guideline for global application developed through the Clinical and Laboratory Standards Institute consensus process.



# FLUOROQUINOLONES

Organism	Method	Ciprofloxacin Previous			Ciprofloxacin New		
		S	I	R	S	I	R
<i>Enterobacteriaceae</i>	BMD	≤ 1	2	≥ 4	≤ 0.25	0.5	≥ 1
<i>P. aeruginosa</i>	BMD	≤ 1	2	≥ 4	≤ 0.5	1	≥ 2
<i>Enterobacteriaceae</i>	DD	≥ 21	16-20	≤ 15	≥ 26	22-25	≤ 21
<i>P. aeruginosa</i>	DD	≥ 21	16-20	≤ 15	≥ 25	19-24	≤ 18

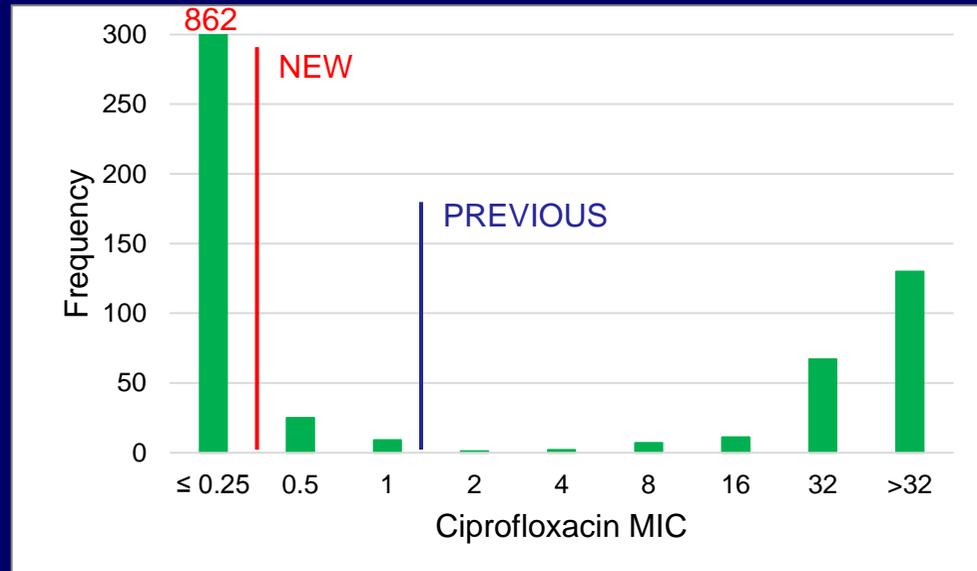
Organism	Method	Levofloxacin Previous			Levofloxacin New		
		S	I	R	S	I	R
<i>Enterobacteriaceae</i>	BMD	≤ 2	4	≥ 8	≤ 0.5	1	≥ 2
<i>P. aeruginosa</i>	BMD	≤ 2	4	≥ 8	≤ 1	2	≥ 4
<i>Enterobacteriaceae</i>	DD	≥ 17	14-16	≤ 13	≥ 21	17-20	≤ 16
<i>P. aeruginosa</i>	DD	≥ 17	14-16	≤ 13	≥ 22	15-21	≤ 14

# WISCONSIN SURVEILLANCE

Organism	n	Ciprofloxacin Previous			Ciprofloxacin New		
		%S	%I	%R	%S	%I	%R
<i>Escherichia coli</i>	1114	80.4	0.1	19.5	77.4	2.2	20.4
<i>Proteus mirabilis</i>	916	78.5	3.6	17.9	76.0	1.4	22.6
<i>Pseudomonas aeruginosa</i>	801	86.8	3.7	9.5	82.4	4.4	13.2

# WISCONSIN SURVEILLANCE

Organism	n	Ciprofloxacin Previous			Ciprofloxacin New		
		%S	%I	%R	%S	%I	%R
<i>Escherichia coli</i>	1114	80.4	0.1	19.5	77.4	2.2	20.4
<i>Proteus mirabilis</i>	916	78.5	3.6	17.9	76.0	1.4	22.6
<i>Pseudomonas aeruginosa</i>	801	86.8	3.7	9.5	82.4	4.4	13.2

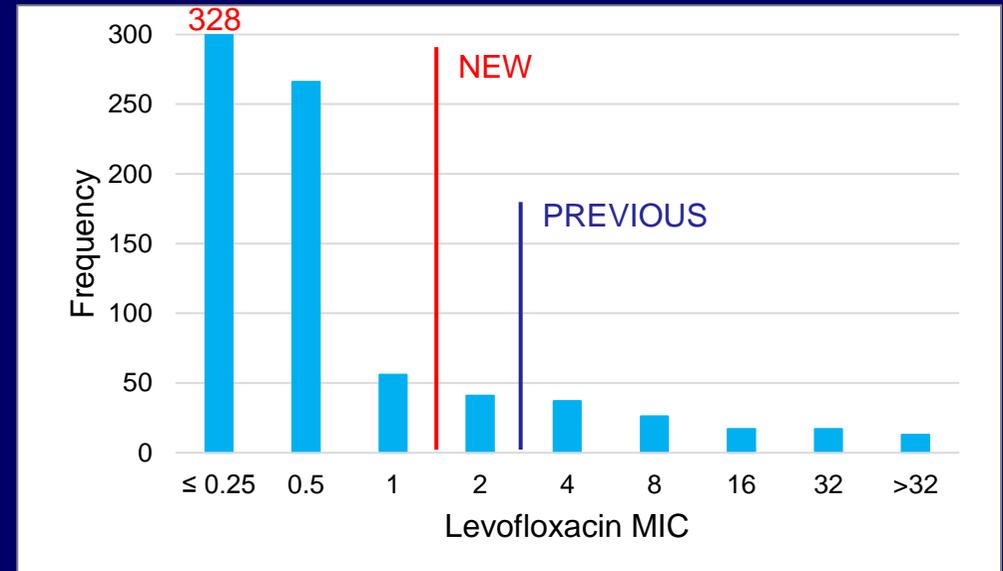
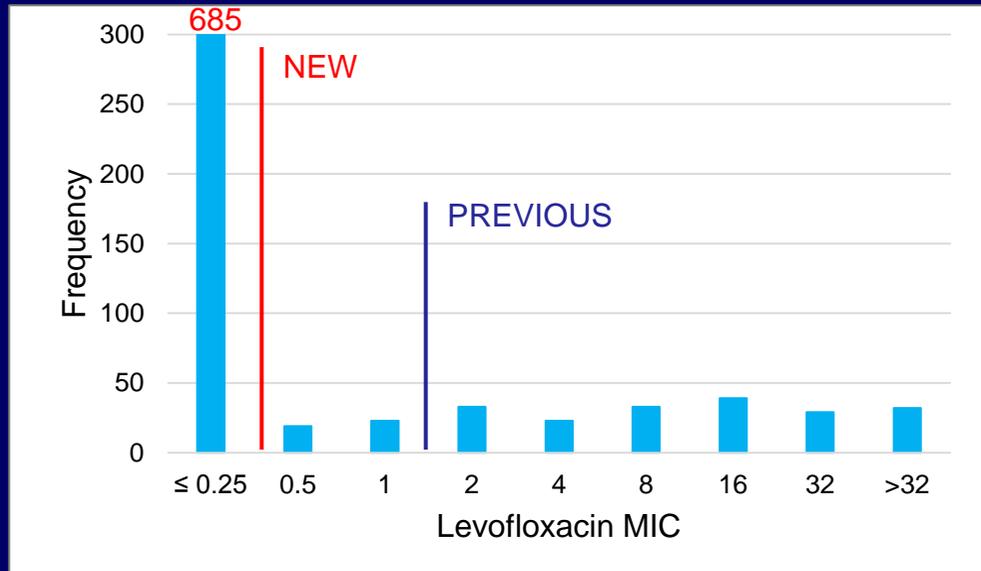


# WISCONSIN SURVEILLANCE

Organism	n	Levofloxacin Previous			Levofloxacin New		
		%S	%I	%R	%S	%I	%R
<i>Escherichia coli</i>	1114	80.6	0.4	18.9	79.5	0.9	19.6
<i>Proteus mirabilis</i>	916	83.0	2.5	14.5	76.9	2.5	20.6
<i>Pseudomonas aeruginosa</i>	801	86.3	4.6	9.1	81.1	5.1	13.7

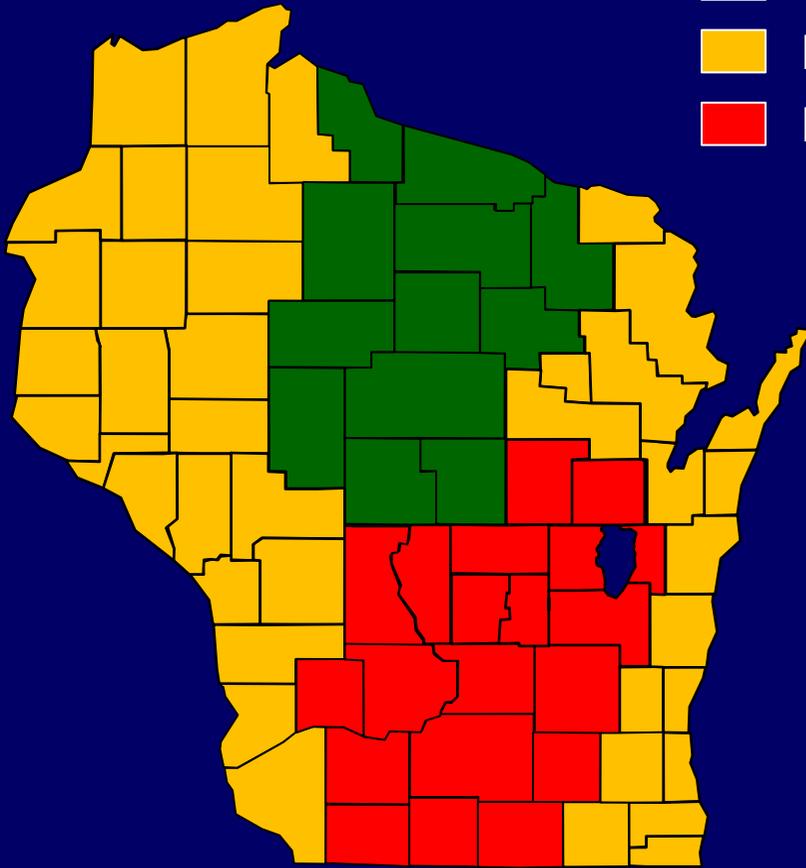
# WISCONSIN SURVEILLANCE

Organism	n	Levofloxacin Previous			Levofloxacin New		
		%S	%I	%R	%S	%I	%R
<i>Escherichia coli</i>	1114	80.6	0.4	18.9	79.5	0.9	19.6
<i>Proteus mirabilis</i>	916	83.0	2.5	14.5	76.9	2.5	20.6
<i>Pseudomonas aeruginosa</i>	801	86.3	4.6	9.1	81.1	5.1	13.7

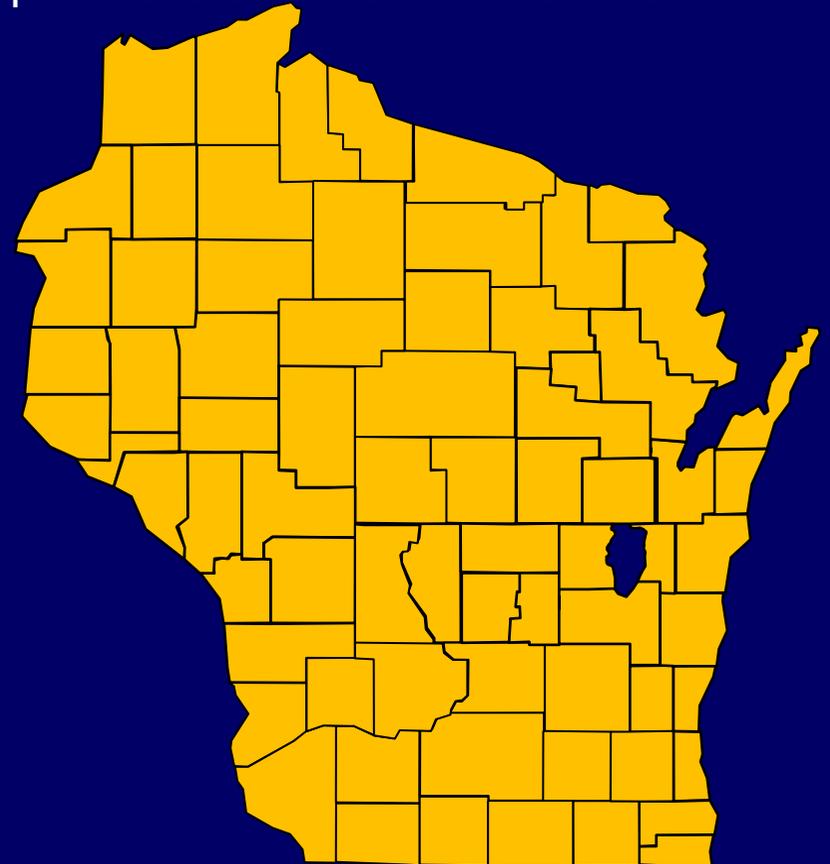


# *E. coli* / CIPROFLOXACIN

- Percentage susceptible 5% or more greater than state mean
- Percentage susceptible  $\pm 5\%$  of state mean
- Percentage susceptible 5% or more less than state mean



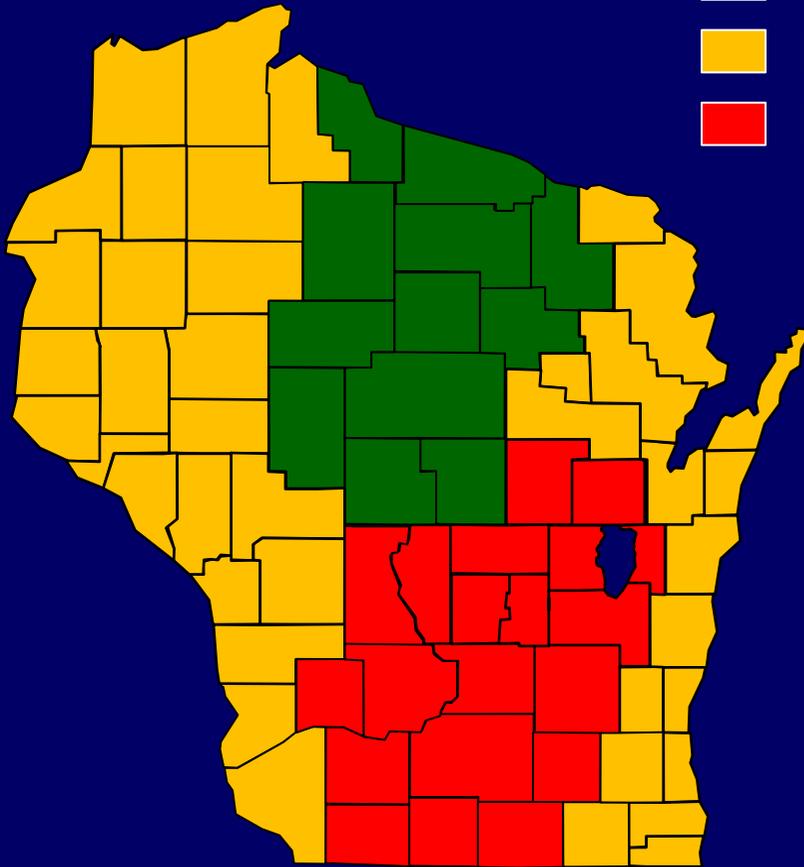
Previous  
state mean 80.4%



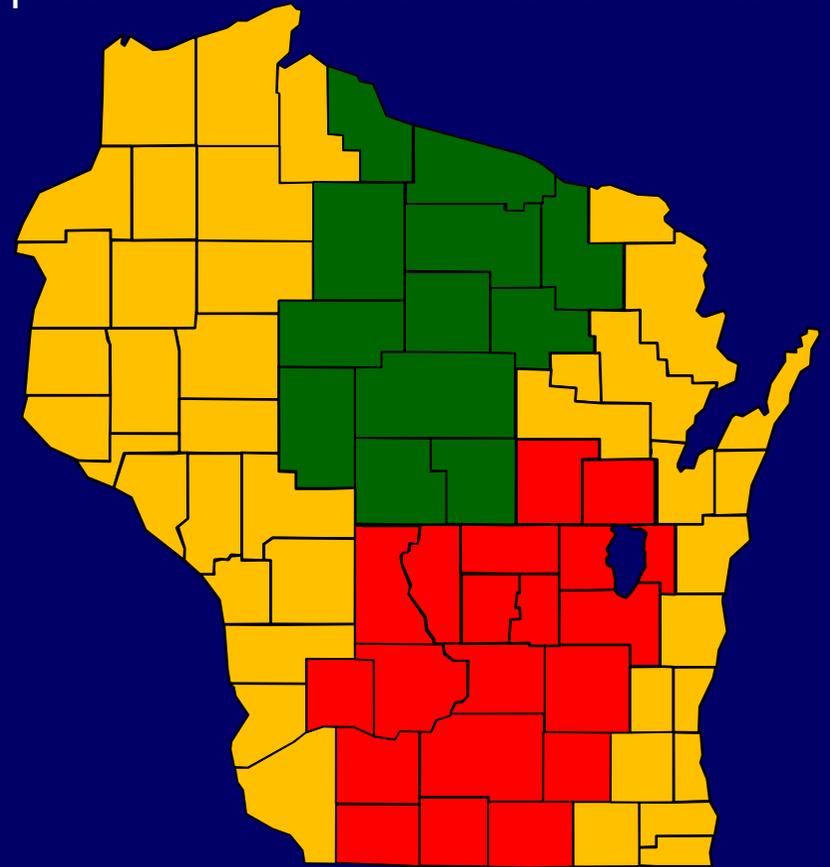
New  
state mean 77.4%

# *E. coli* / LEVOFLOXACIN

- Percentage susceptible 5% or more greater than state mean
- Percentage susceptible  $\pm 5\%$  of state mean
- Percentage susceptible 5% or more less than state mean



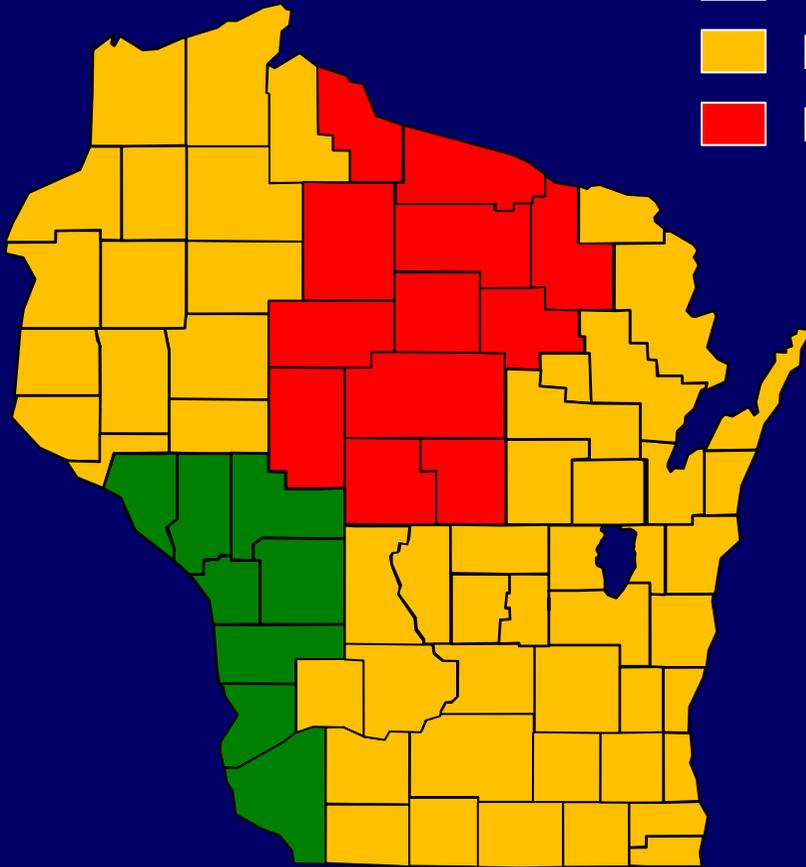
Previous  
state mean 80.6%



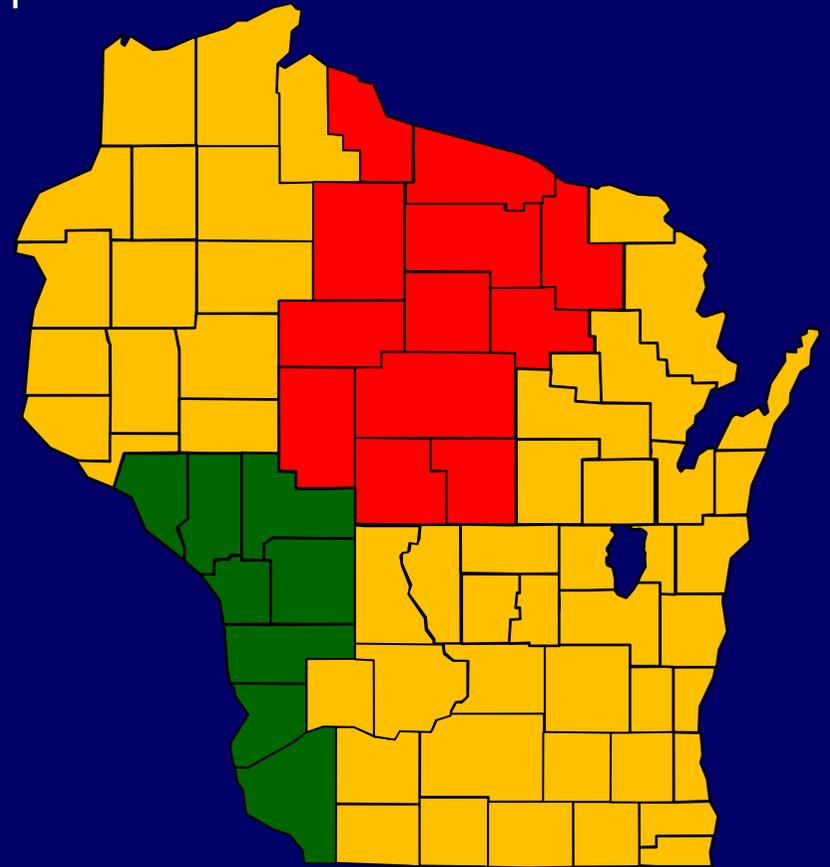
New  
state mean 79.5%

# *P. mirabilis* / CIPROFLOXACIN

- Percentage susceptible 5% or more greater than state mean
- Percentage susceptible  $\pm 5\%$  of state mean
- Percentage susceptible 5% or more less than state mean



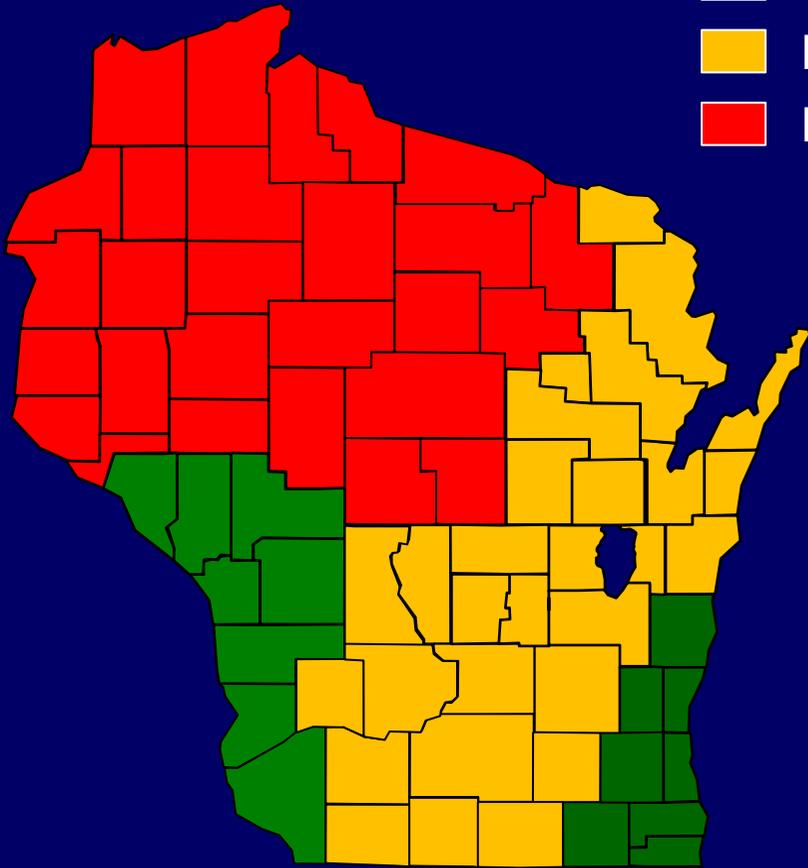
Previous  
state mean 78.5%



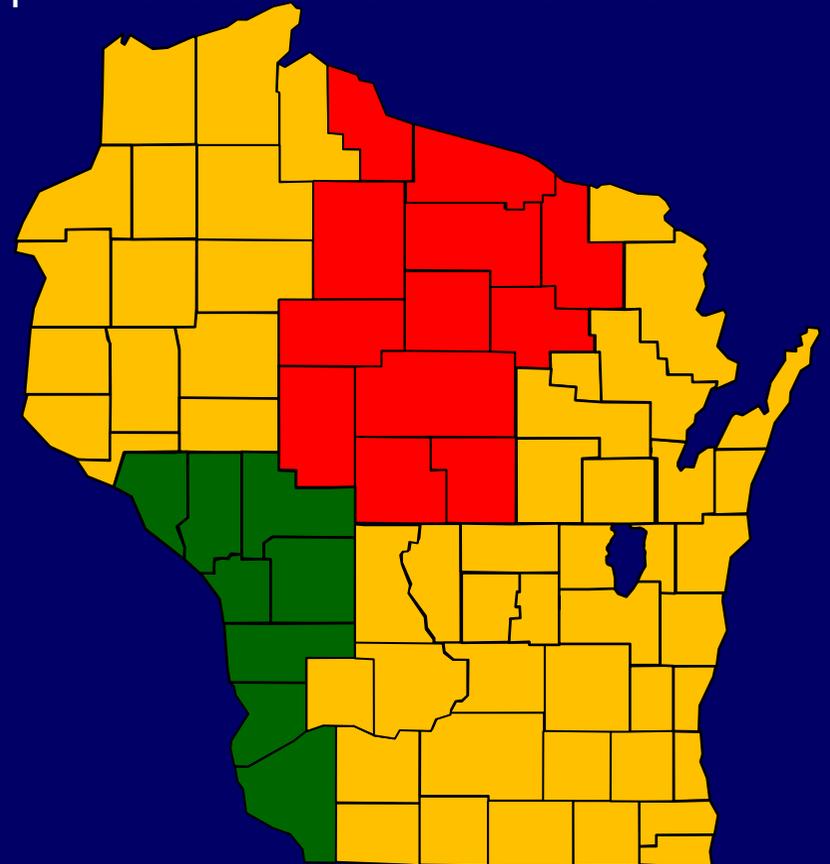
New  
state mean 76.0%

# *P. mirabilis* / LEVOFLOXACIN

- Percentage susceptible 5% or more greater than state mean
- Percentage susceptible  $\pm 5\%$  of state mean
- Percentage susceptible 5% or more less than state mean



Previous  
state mean 83.0%



New  
state mean 76.9%

# CEFTAROLINE

Organism	Method	Ceftaroline Previous			Ceftaroline New		
		S	I	R	S	SDD	R
<i>S. aureus</i> (incl. MRSA)	BMD	≤ 1	2	≥ 4	≤ 1	2-4	≥ 8
<i>S. aureus</i> (incl. MRSA)	DD	≥ 24	21-23	≤ 20	≥ 25	20-24	≤ 19

# CEFTAROLINE

Organism	Method	Ceftaroline Previous			Ceftaroline New		
		S	I	R	S	SDD	R
<i>S. aureus</i> (incl. MRSA)	BMD	≤ 1	2	≥ 4	≤ 1	2-4	≥ 8
<i>S. aureus</i> (incl. MRSA)	DD	≥ 24	21-23	≤ 20	≥ 25	20-24	≤ 19

MIC	Number of Wisconsin Isolates
≤ 0.12	126
0.25	133
0.5	49
1	1
2	0
4	1
8	0

# SUSCEPTIBLE DOSE DEPENDENT

- Intermediate
  - Approach attainable blood and tissue levels but have less clinical response than “susceptible”
  - Also implies clinical efficacy in sites where agents are physiologically concentrated
- Susceptible dose dependent (multiple regimens)
  - Implies that susceptibility of isolate is dependent on dosing regimen
  - Higher dose or more-frequent dosing results in higher drug exposure

# REVISED LANGUAGE

- Appendix E lists doses used for establishing SDD; drug label should be consulted for recommended doses and adjustment for organ function
- SDD category may be assigned when doses well above “susceptible” breakpoint are/have:
  - Supported by literature
  - Widely used clinically
  - Sufficient data to justify

# WISCONSIN SURVEILLANCE

Organism	n	% Cefepime Susceptible Dose Dependent (MIC 4-8 $\mu\text{g}/\text{mL}$ )
<i>Escherichia coli</i>	1114	1.5
<i>Proteus mirabilis</i>	916	0.4
<i>Enterobacter cloacae</i>	277	4.7
<i>Klebsiella pneumoniae</i>	304	0.0



# DAPTOMYCIN

Organism	Method	Daptomycin Previous			Daptomycin New		
		S	I	R	S	SDD	R
<i>Enterococcus</i> spp.	BMD	≤ 4			≤ 1	2-4	≥ 8

- Should not be reported for respiratory isolates
- SDD breakpoint based on dosage regimen of 8-12 mg/kg per day in adults
- Intended for serious *Enterococcus* spp. infections

# OTHER TABLE 2 ADDITIONS

- Follow-up MIC testing for *Enterobacteriaceae* / ceftazidime-avibactam zone sizes of 18-20 mm (disk diffusion may overcall resistance)

Organism	Acceptable Methods				
	Cefoxitin MIC	Cefoxitin disk diffusion	Oxacillin MIC	Oxacillin disk diffusion	Oxacillin salt agar
<i>S. aureus</i>	Yes	Yes	Yes	No	Yes
<i>S. lugdunensis</i>	Yes	Yes	Yes	No	No
<b><i>S. epidermidis</i></b>	<b>No</b>	<b>Yes</b>	<b>Yes</b>	<b>Yes</b>	<b>No</b>
<i>S. pseudintermedius</i>	No	No	Yes	Yes	No
<i>S. schleiferi</i>	No	No	Yes	Yes	No
<b>Other <i>Staphylococcus</i> spp. (not listed above)</b>	No	Yes	Yes*	No	No

\* For **other *Staphylococcus* spp.** with oxacillin MICs between 0.5–2 µg/mL, see comment (17) for recommendations on testing for *mecA* or for PBP2a.

Mechanisms of oxacillin resistance other than *mecA* are rare and include a novel *mecA* homologue, *mecC*.<sup>3</sup> MICs for strains with *mecC* are typically **cefoxitin resistant and oxacillin susceptible**; *mecC* resistance cannot be detected by tests directed at *mecA* or PBP2a.

# STAPHYLOCOCCI AND *mecA*

- Added *S. epidermidis*-specific breakpoints
  - Oxacillin disk diffusion
  - Oxacillin broth microdilution
  - Cefoxitin disk diffusion (cefoxitin MIC not reliable)
- Noteworthy revisions
  - Oxacillin disk diffusion not reliable for *S. aureus* and *S. lugdunensis*
  - Oxacillin MIC (0.5-2 µg/mL) may overcall resistance in **staphylococci other than...*S. aureus* *S. schleiferi* *S. pseudintermedius* *S. lugdunensis* *S. epidermidis***

# MORE TABLE 2 ADDITIONS

Table 2C. *Staphylococcus* spp. (Continued)

Test/Report Group	Antimicrobial Agent	<i>Staphylococcus</i> spp. Indications	Interpretive Categories and Zone Diameter Breakpoints, nearest whole mm			Interpretive Categories and MIC Breakpoints, µg/mL			Comments	
			SDD	I	R	S	SDD	I		R
<b>CEPHEMS (PARENTERAL)</b>										
B	Ceftaroline	<b><i>S. aureus</i> only, including MRSA</b>	—	—	≤19	≤1	2–4	—	≥8	(18) The breakpoint for susceptible is based on a dosage regimen of 600 mg administered every 12 h.  (19) The breakpoint for SDD is based on a dosage in adults of 600 mg every 8 h administered over 2 h.
<b>GLYCOPEPTIDES</b>										
(20) MIC tests should be performed to determine the susceptibility of all isolates of staphylococci to vancomycin. The disk test does not differentiate vancomycin-susceptible isolates of <i>S. aureus</i> from vancomycin-intermediate isolates, nor does the test differentiate among vancomycin-susceptible, -intermediate, and -resistant isolates of <i>Staphylococcus</i> spp. other than <i>S. aureus</i> , all of which give similar size zones of inhibition.										
B	Vancomycin	<b><i>S. aureus</i> only</b>	—	—	—	≤2	—	4–8	≥16	(21) For <i>S. aureus</i> , vancomycin-susceptible isolates may become vancomycin intermediate during the course of prolonged therapy.  (22) Send any <i>S. aureus</i> for which the vancomycin is ≥8 µg/mL to a referral laboratory. See Appendix A.  Also refer to Table 3F for <i>S. aureus</i> , Subchapter 3.12 in M07 <sup>2</sup> and Subchapter 3.9 in M02. <sup>1</sup>
		<b><i>Staphylococcus</i> spp. other than <i>S. aureus</i></b>	—	—	—	≤4	—	8–16	≥32	See comment (19).  (23) Send any <i>Staphylococcus</i> spp. for which the vancomycin MIC is ≥32 µg/mL to a referral laboratory. See Appendix A.  See also Subchapter 3.12 in M07 <sup>2</sup> and Subchapter 3.9 in M02. <sup>1</sup>
Inv.	Teicoplanin	<b>All staphylococci</b>	—	—	—	≤8	—	16	≥32	
<b>LIPOGLYCOPEPTIDES</b>										
C	Dalbavancin	<b><i>S. aureus</i> only, including MRSA</b>	—	—	—	≤0.25	—	—	—	
C	Oritavancin	<b><i>S. aureus</i> only, including MRSA</b>	—	—	—	≤0.12	—	—	—	
C	Telavancin	<b><i>S. aureus</i> only, including MRSA</b>	—	—	—	≤0.12	—	—	—	

# OTHER TABLE 2 DELETIONS

- Norfloxacin disk diffusion and MIC breakpoints

*Enterobacteriaceae*      *Pseudomonas aeruginosa*  
*Staphylococcus* spp.      *Enterococcus* spp.  
Non-*Enterobacteriaceae* (MIC breakpoints)

- Telrithromycin disk diffusion and MIC breakpoints

*Staphylococcus* spp.      *Haemophilus influenzae*  
*Haemophilus parainfluenzae*  
*Streptococcus pneumoniae*

# OTHER TABLE 2 DELETIONS

- *N. gonorrhoeae* disk diffusion and MIC breakpoints

Cefuroxime

Cefmetazole

Ceftazidime

Cefetamet

Enoxacin

Fleroxacin

Lomefloxacin

Ofloxacin

# TABLE 2 CLARIFICATIONS

- Organisms susceptible to tetracycline also susceptible to doxycycline and minocycline; **resistance to doxycycline and minocycline cannot be inferred by resistance to tetracycline**

*Haemophilus influenzae*

*Haemophilus parainfluenzae*

*Streptococcus pneumoniae*

$\beta$ -hemolytic *Streptococcus*

viridans group *Streptococcus*



## Table 3



# WHAT'S IN THERE?

- ESBL detection in *E. coli*, *K. pneumoniae*, *K. oxytoca*, *P. mirabilis*
- Carbapenemase production in *Enterobacteriaceae* and *Pseudomonas aeruginosa*

2010 breakpoints: CarbaNP, mCIM, eCIM, molecular  
Current breakpoints: may not need additional tests

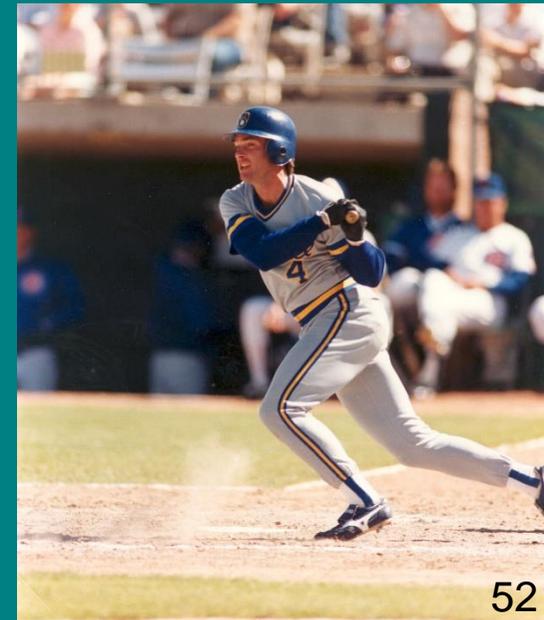
*No longer necessary to edit carbapenem results to resistant if a carbapenemase producer is detected*

# WHAT ELSE IS IN THERE?

- $\beta$ -lactamase detection *Staphylococcus* spp.
- Methicillin (oxacillin) resistance *Staphylococcus* spp.
- Vancomycin agar screen *Staphylococcus aureus* and *Enterococcus* spp.
- Inducible clindamycin resistance
- High-level mupirocin resistance *S. aureus*
- High-level aminoglycoside resist *Enterococcus* spp.



# Table 4



# TABLE 4

Table 4A-1  
Nonfastidious Disk Diffusion QC Excluding  $\beta$ -Lactam Combination Agents  
M02

*E. coli* ATCC 25922  
*P. aeruginosa* ATCC 27853  
*S. aureus* ATCC 25923

Table 4B  
Fastidious Disk Diffusion QC  
M02

*H. influenzae* ATCC 49247  
*H. influenzae* ATCC 49766  
*N. gonorrhoeae* ATCC 49226  
*S. pneumoniae* ATCC 49619

Table 4A-2  
Nonfastidious Disk Diffusion QC for  $\beta$ -Lactam Combination Agents  
M02

Same as left; add:

*E. coli* ATCC 35218  
*K. pneumoniae* ATCC 700603  
*E. coli* NCTC 13353  
*K. pneumoniae* ATCC BAA-1705  
*K. pneumoniae* ATCC BAA-2814  
*A. baumannii* NCTC 13304

# DISK DIFFUSION QC RANGES

- Noteworthy additions

  - ATCC 25922 and ATCC 27853 range for tebipenem

  - ATCC 49226 ranges for azithromycin, gepotidacin

  - Acinetobacter baumannii* NCTC 13304 as QC strain

  - Caveat for imipenem-relebactam (one manufacturer)

- Noteworthy deletions

  - Ranges for methicillin

  - Ranges for mezlocillin

  - Ranges for norfloxacin

# DISK DIFFUSION QC ADDED RANGES

<i>E. coli</i> ATCC 25922	cefepime-zidebactam imipenem-relebactam
<i>P. aeruginosa</i> ATCC 27853	cefepime-zidebactam imipenem-relebactam
<i>K. pneumoniae</i> ATCC 700603	cefepime; cefepime-zidebactam imipenem; imipenem-relebactam
<i>E. coli</i> NCTC 13353	cefepime; cefepime-zidebactam
<i>K. pneumoniae</i> ATCC BAA-1705	imipenem; imipenem-relebactam
<i>K. pneumoniae</i> ATCC BAA-2814	imipenem; imipenem-relebactam
<i>A. baumannii</i> NCTC 13304	cefepime; cefepime-zidebactam

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# Table 5



# TABLE 5

Table 5A-1  
Nonfastidious MIC QC Excluding  $\beta$ -Lactam Combination Agents  
M07

*E. coli* ATCC 25922  
*P. aeruginosa* ATCC 27853  
*S. aureus* ATCC 29213  
*E. faecalis* ATCC 29212

Table 5A-2  
Nonfastidious MIC QC for  $\beta$ -Lactam Combination Agents  
M07

Same as left; add:

*E. coli* ATCC 35218  
*K. pneumoniae* ATCC 700603  
*E. coli* NCTC 13353  
*K. pneumoniae* ATCC BAA-1705  
*K. pneumoniae* ATCC BAA-2814  
*A. baumannii* NCTC 13304

Table 5B  
Fastidious MIC QC Broth Dilution  
M07

*H. influenzae* ATCC 49247  
*H. influenzae* ATCC 49766  
*N. gonorrhoeae* ATCC 49226 (agar dilution)  
*S. pneumoniae* ATCC 49619

# MORE TABLE 5

Table 5D  
Anaerobe MIC QC  
Agar Dilution  
M11

Table 5E  
Anaerobe MIC QC  
Broth Microdilution  
M11

*B. fragilis* ATCC 25285

*B. thetaiotaomicron* ATCC 29741

*C. difficile* ATCC 700057

*E. lenta* ATCC 43055

# MIC QC RANGES

- Noteworthy additions

  - ATCC 27853 range for meropenem
  - QC ranges established for tebipenem

- Noteworthy revisions

  - ATCC 27853 range for ciprofloxacin
  - ATCC 35218 and NCTC 13353 for cefepime

- Noteworthy deletions

  - Ranges for methicillin, norfloxacin
  - Ranges for mezlocillin (including anaerobe agar dilution)

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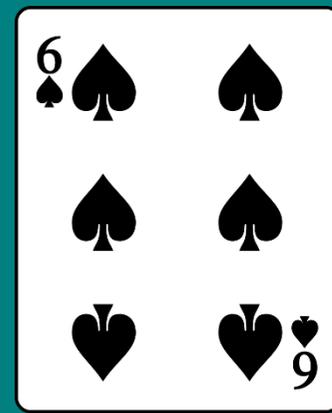
# MIC QC ADDED RANGES

<i>E. coli</i> ATCC 25922	meropenem-nacubactam; nacubactam
<i>P. aeruginosa</i> ATCC 27853	meropenem-nacubactam; nacubactam
<i>E. coli</i> ATCC 35218	cefpodoxime
<i>K. pneumoniae</i> ATCC 700603	cefpodoxime
<i>E. coli</i> NCTC 13353	cefpodoxime
<i>K. pneumoniae</i> ATCC BAA-2814	meropenem-nacubactam; nacubactam

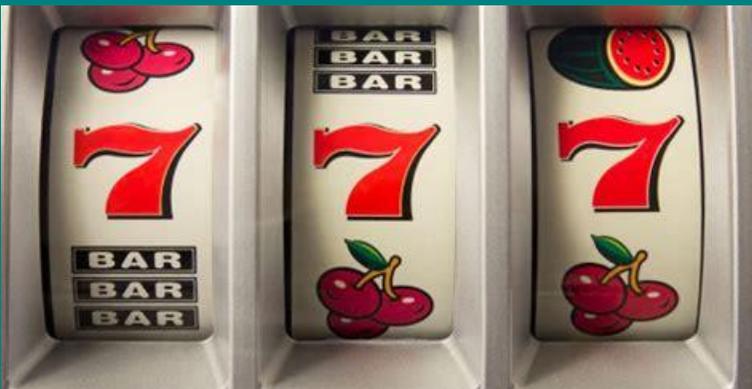
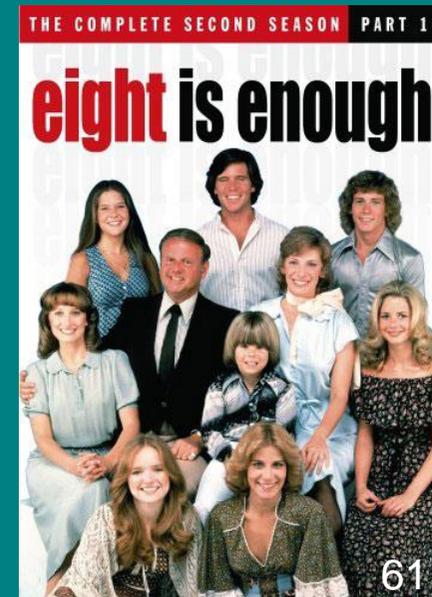
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**I'M  
RETIRED  
SO  
DO IT  
YOURSELF!**



## Tables 6-8



# TABLE 6 PREPARING STOCK SOLNS

- Added solvent and diluent information for:

Nacubactam

Tebipenem

Zidebactam

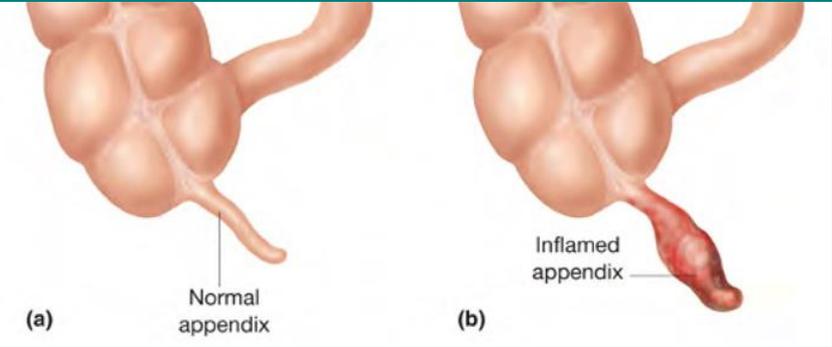
- Prep instructions for meropenem/nacubactam

- Deleted solvent and diluent information for:

Methicillin

Mezlocillin

Norfloxacin



# Appendices



# APPENDIX A; SUGGESTED CONFIRM

Appendix A. (Continued)		Occurrence and Significance of Resistance and Actions to Take Following Confirmation of Results <sup>a</sup>		
Organism or Organism Group	Resistance Phenotype Detected <sup>b</sup>	Category I	Category II	Category III
		Not reported or only rarely reported to date	Uncommon in most institutions	May be common but generally considered of epidemiological concern
<i>Stenotrophomonas maltophilia</i>	Trimethoprim-sulfamethoxazole – I or R		X	
<i>Haemophilus influenzae</i>	Carbapenem – NS Ceftaroline – NS Extended-spectrum cephalosporin <sup>c</sup> – NS Fluoroquinolone – NS	X		
	Amoxicillin-clavulanate – R Ampicillin – R and $\beta$ -lactamase negative		X	
<i>Neisseria gonorrhoeae</i>	<b>Azithromycin – NS</b>		X	
	Extended-spectrum cephalosporin <sup>c</sup> – NS		X	
	Fluoroquinolone – I or R			X
<i>Neisseria meningitidis</i>	Ampicillin or penicillin – R Extended-spectrum cephalosporin <sup>c</sup> – NS Meropenem – NS	X		
	Ampicillin or penicillin – I Azithromycin – NS Chloramphenicol – I or R Fluoroquinolone – I or R Minocycline – NS Rifampin – I or R		X	
<i>Enterococcus</i> spp.	Dalbavancin – NS Oritavancin – NS Telavancin – NS	X		
	Daptomycin – NS Linezolid – R Tedizolid – NS		X	
	High-level aminoglycoside – R Vancomycin – R			X

# APPENDIX B; INTRINSIC RESISTANT

## Appendix B. (Continued)

### B1. Enterobacteriaceae

Antimicrobial Agent Organism	Ampicillin	Amoxicillin-clavulanate	Ampicillin-sulbactam	Piperacillin	Ticarcillin	Cephalosporins I: Cefazolin, Cephalothin	Cephamycins: Cefoxitin, Cefotetan	Cephalosporin II: Cefuroxime	Imipenem	Tetracyclines	Tigecycline	Nitrofurantoin	Polymyxin B Colistin	Aminoglycosides
<i>Citrobacter freundii</i>	R	R	R			R	R	R						
<i>Citrobacter koseri</i> , <b><i>Citrobacter amalonaticus</i></b> <b>group<sup>a</sup></b>	R				R									
<i>Enterobacter cloacae</i> complex <sup>b</sup>	R	R	R			R	R							
<i>Escherichia coli</i>	There is no intrinsic resistance to $\beta$ -lactams in this organism.													
<i>Escherichia hermannii</i>	R				R									
<i>Hafnia alvei</i>	R	R	R			R	R							
<i>Klebsiella</i> (formerly <i>Enterobacter) aerogenes</i>	R	R	R			R	R							
<i>Klebsiella pneumoniae</i> , <b><i>Klebsiella oxytoca</i>, <i>Klebsiella</i></b> <b><i>variicola</i></b>	R				R									
<i>Morganella morganii</i>	R	R				R		R	°		R	R	R	
<i>Proteus mirabilis</i>	There is no intrinsic resistance to penicillins and cephalosporins in this organism.								°	R	R	R	R	
<i>Proteus penneri</i>	R					R		R	°	R	R	R	R	
<i>Proteus vulgaris</i>	R					R		R	°	R	R	R	R	
<i>Providencia rettgeri</i>	R	R				R			°	R	R	R	R	
<i>Providencia stuartii</i>	R	R				R			°	R	R	R	R	
<b><i>Raoultella</i> spp.<sup>e</sup></b>	<b>R</b>				<b>R</b>									<sup>d</sup>

# APPENDIX B; INTRINSIC RESISTANT

## B2. Non-Enterobacteriaceae

Antimicrobial Agent \ Organism	Ampicillin, Amoxicillin	Piperacillin	Ticarcillin	Ampicillin-sulbactam	Amoxicillin-clavulanate	Piperacillin-tazobactam	Cefotaxime	Ceftriaxone	Ceftazidime	Cefepime	Aztreonam	Imipenem	Meropenem	Ertapenem	Polymyxin B Colistin	Aminoglycosides	Tetracyclines/ Tigecycline	Trimethoprim	Trimethoprim- sulfamethoxazole	Chloramphenicol	Fosfomycin	
<i>Acinetobacter baumannii</i> / <i>Acinetobacter calcoaceticus</i> complex	R				R						R			R					R		R	R
<b><i>Burkholderia cepacia</i> complex<sup>a</sup></b>	<b>R</b>	<b>R</b>	<b>R</b>	<b>R</b>	<b>R</b>	<sup>a</sup>	<sup>a</sup>	<sup>a</sup>		<sup>a</sup>	<sup>a</sup>	<sup>a</sup>		<b>R</b>	<b>R</b>	<sup>a</sup>		<sup>a</sup>				R
<i>Pseudomonas aeruginosa</i>	R			R	R		R	R						R			R	R	R	R	R	
<i>Stenotrophomonas maltophilia</i>	R	R	R	R	R	R	R	R			R	R	R	R		R	<sup>b</sup>	R				R

Abbreviation: R, resistant.

\*\* <sup>a</sup> insufficient *in vitro* / *in vivo* correlation

Additional tables for *Staphylococcus* spp., *Enterococcus* spp., *Bacteroides* spp.;  
select *Clostridium* spp., *Fusobacterium* spp.

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# APPENDIX C; QC STRAINS

Appendix C. (Continued)					
QC Strains	Organism Characteristics	Disk Diffusion Tests	MIC Tests	Other Tests	Comments
<i>Escherichia coli</i> ATCC <sup>®</sup> 25922	<ul style="list-style-type: none"> <li>• <math>\beta</math>-lactamase negative</li> </ul>	<ul style="list-style-type: none"> <li>• Nonfastidious gram-negative bacteria</li> <li>• <i>Neisseria meningitidis</i></li> </ul>	<ul style="list-style-type: none"> <li>• Nonfastidious gram-negative bacteria</li> <li>• <i>N. meningitidis</i></li> </ul>		
<i>E. coli</i> ATCC <sup>®</sup> 35218 <sup>a,b,1,2</sup>	<ul style="list-style-type: none"> <li>• TEM-1</li> </ul>	<ul style="list-style-type: none"> <li>• <math>\beta</math>-lactam combination agents</li> </ul>	<ul style="list-style-type: none"> <li>• <math>\beta</math>-lactam combination agents</li> </ul>		
<i>E. coli</i> NCTC 13353 <sup>a,b,3</sup>	<ul style="list-style-type: none"> <li>• CTX-M-15 (ESBL)</li> </ul>	<ul style="list-style-type: none"> <li>• <math>\beta</math>-lactam combination agents</li> </ul>	<ul style="list-style-type: none"> <li>• <math>\beta</math>-lactam combination agents</li> </ul>		
<i>Haemophilus influenzae</i> ATCC <sup>®</sup> 10211					<ul style="list-style-type: none"> <li>• Assess each batch/lot of HTM for growth capabilities.</li> </ul>
<i>H. influenzae</i> ATCC <sup>®</sup> 49247	<ul style="list-style-type: none"> <li>• BLNAR</li> </ul>	<ul style="list-style-type: none"> <li>• <i>H. influenzae</i></li> <li>• <i>Haemophilus parainfluenzae</i></li> </ul>	<ul style="list-style-type: none"> <li>• <i>H. influenzae</i></li> <li>• <i>H. parainfluenzae</i></li> </ul>		
<i>H. influenzae</i> ATCC <sup>®</sup> 49766	<ul style="list-style-type: none"> <li>• Ampicillin susceptible</li> </ul>	<ul style="list-style-type: none"> <li>• <i>H. influenzae</i></li> <li>• <i>H. parainfluenzae</i></li> </ul>	<ul style="list-style-type: none"> <li>• <i>H. influenzae</i></li> <li>• <i>H. parainfluenzae</i></li> </ul>		<ul style="list-style-type: none"> <li>• More reproducible than <i>H. influenzae</i> ATCC<sup>®</sup> 49247 with selected <math>\beta</math>-lactam agents</li> </ul>
<i>Klebsiella pneumoniae</i> ATCC <sup>®</sup> 700603 <sup>a,b</sup>	<ul style="list-style-type: none"> <li>• SHV-18 (ESBL)<sup>1,2</sup></li> <li>• OXA-2</li> <li>• Mutations in OMPK35 and OMPK37</li> </ul>	<ul style="list-style-type: none"> <li>• <math>\beta</math>-lactam combination agents</li> </ul>	<ul style="list-style-type: none"> <li>• <math>\beta</math>-lactam combination agents</li> </ul>	<ul style="list-style-type: none"> <li>• ESBL tests</li> </ul>	
<i>K. pneumoniae</i> ATCC <sup>®</sup> BAA-1705 <sup>TM,a,b</sup>	<ul style="list-style-type: none"> <li>• KPC-2 (carbapenemase)</li> <li>• TEM</li> <li>• SHV</li> </ul>	<ul style="list-style-type: none"> <li>• <math>\beta</math>-lactam combination agents</li> </ul>	<ul style="list-style-type: none"> <li>• <math>\beta</math>-lactam combination agents</li> </ul>	<ul style="list-style-type: none"> <li>• Carbapenemase tests</li> </ul>	
<i>K. pneumoniae</i> ATCC <sup>®</sup> BAA-1706 <sup>TM</sup>	<ul style="list-style-type: none"> <li>• Resistant to carbapenems by noncarbapenemase mechanism</li> </ul>			<ul style="list-style-type: none"> <li>• Carbapenemase tests</li> </ul>	
<i>K. pneumoniae</i> ATCC <sup>®</sup> BAA-2146 <sup>TM</sup>	<ul style="list-style-type: none"> <li>• NDM</li> </ul>			<ul style="list-style-type: none"> <li>• Carbapenemase tests</li> </ul>	

# APPENDIX D; N ANTIBIOGRAM

Appendix D. Cumulative Antimicrobial Susceptibility Report for Anaerobic Organisms<sup>1</sup>

NOTE: Isolates collected from selected US hospitals from 1 January 2013 to 31 December 2016.<sup>a</sup>

Six US laboratories  
Agar dilution

D1. *Bacteroides* spp. and *Parabacteroides* spp.

Anaerobic Organisms	Number of Strains		Ampicillin-sulbactam		Number of Strains	Piperacillin-tazobactam		Number of Strains	Cefoxitin		Number of Strains	Ertapenem		Number of Strains	Imipenem		Number of Strains	Meropenem	
	%S	%R	%S	%R		%S	%R		%S	%R		%S	%R		%S	%R			
<b>Percent susceptible (% S) and percent resistant (% R)<sup>c</sup></b>																			
<b>Breakpoints, µg/mL</b>	≤ 8/4	≥ 32/16	≤ 16/4	≥ 128/4	≤ 16	≥ 64	≤ 4	≥ 16	≤ 4	≥ 16	≤ 4	≥ 16	≤ 4	≥ 16	≤ 4	≥ 16	≤ 4	≥ 16	
<i>B. fragilis</i>	129	84	2	1030	96	1	830	100	0	133	82	14	189	97	1	1505	93	5	
<i>B. thetaiotaomicron</i>	76	82	5	252	87	0	258	13	54	–	–	–	70	100	0	328	99	0	
<i>B. ovatus</i>	30	80	3	206	94	0	177	20	34	19 <sup>b</sup>	84 <sup>b</sup>	16 <sup>b</sup>	49	100	0	236	95	1	
<i>B. vulgatus</i>	20 <sup>b</sup>	45 <sup>b</sup>	15 <sup>b</sup>	168	92	0	153	73	14	–	–	–	35	97	0	171	96	4	
<i>B. uniformis</i>	19 <sup>b</sup>	84 <sup>b</sup>	0 <sup>b</sup>	78	96	0	72	85	10	–	–	–	19 <sup>b</sup>	100 <sup>b</sup>	0 <sup>b</sup>	93	100	0	
<i>Parabacteroides distasonis</i>	27 <sup>b</sup>	59 <sup>b</sup>	19 <sup>b</sup>	92	95	1	82	29	43	–	–	–	26 <sup>b</sup>	100 <sup>b</sup>	0	119	97	2	

# APPENDIX D; N ANTIBIOGRAM

## D2. Anaerobic Organisms Other Than *Bacteroides* spp. and *Parabacteroides* spp.

Anaerobic Organisms	Number of Strains	Ampicillin-sulbactam		Number of Strains	Piperacillin-tazobactam		Number of Strains	Imipenem		Number of Strains	Meropenem		Number of Strains	Penicillin	
		%S	%R		%S	%R		%S	%R		%S	%R		%S	%R
<b>Percent susceptible (% S) and percent resistant (% R)<sup>b</sup></b>		%S	%R		%S	%R		%S	%R		%S	%R		%S	%R
<b>Breakpoints, µg/mL</b>		≤ 8/4	≥ 32/16		≤ 32/4	≥ 128/4		≤ 4	≥ 16		≤ 4	≥ 16		≤ 0.5	≥ 2
<i>Prevotella</i> spp.	29 <sup>c</sup>	97 <sup>c</sup>	3 <sup>c</sup>	63	100	0	29 <sup>c</sup>	100	0	92	98	0	63	100	0
<i>Fusobacterium</i> spp.	20 <sup>c</sup>	100 <sup>c</sup>	0 <sup>c</sup>	55	96	2	75	95	4	20 <sup>c</sup>	100 <sup>c</sup>	0 <sup>c</sup>	— <sup>d</sup>	— <sup>d</sup>	— <sup>d</sup>
Anaerobic gram-positive cocci <sup>e</sup>	— <sup>d</sup>	— <sup>d</sup>	— <sup>d</sup>	1853	99	1	134	99	0	1647	100	0	1647	100	0
<i>Cutibacterium</i> (formerly <i>Propionibacterium</i> ) <i>acnes</i> <sup>f</sup>	— <sup>d</sup>	— <sup>d</sup>	— <sup>d</sup>	18 <sup>c</sup>	100 <sup>c</sup>	0 <sup>c</sup>	17 <sup>c</sup>	94 <sup>c</sup>	0 <sup>d</sup>	— <sup>d</sup>	— <sup>d</sup>	— <sup>d</sup>	— <sup>d</sup>	— <sup>d</sup>	— <sup>d</sup>
<i>Clostridium perfringens</i>	15 <sup>c</sup>	100 <sup>c</sup>	0	410	100	0	23 <sup>c</sup>	100 <sup>c</sup>	0 <sup>c</sup>	417	100	0	402	90	4
<i>Clostridioides</i> (formerly <i>Clostridium</i> ) <i>difficile</i> <sup>g</sup>	76	99	0	542	93	0	480	69	4	609	99	0	533	6	37
Other <i>Clostridium</i> spp.	— <sup>d</sup>	— <sup>d</sup>	— <sup>d</sup>	439	94	1	71	99	0	390	100	0	390	69	13

# APPENDIX E; S and SDD DOSING

## Appendix E. Dosage Regimens Used to Establish Susceptible or Susceptible-Dose Dependent Breakpoints

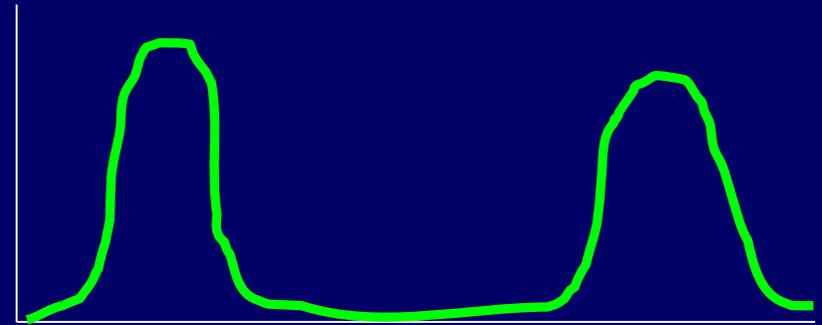
The evolving science of pharmacokinetics-pharmacodynamics has become increasingly important in recent years in determining minimal inhibitory concentration (MIC) breakpoints. Recently approved susceptible or susceptible-dose dependent (SDD) breakpoints for a number of agents have been based on a specific dosage regimen(s); these dosage regimens are listed in the table below. Proper application of the breakpoints necessitates drug exposure at the site of infection that corresponds to or exceeds the expected systemic drug exposure at the dose listed in adult patients with normal renal function. This information should be shared with pharmacists, infectious diseases staff, and others making dosing recommendations for the institution.

Antimicrobial Agent	Breakpoints and Interpretive Categories			
	Susceptible		SDD	
	MIC	Dose	MIC	Dose
<b>Table 2A. Enterobacteriaceae</b>				
Azithromycin ( <i>Salmonella</i> Typhi)	≤ 16 µg/mL	500 mg <b>administered</b> daily	N/A	
Aztreonam	≤ 4 µg/mL	1 g <b>administered</b> every 8 h	N/A	
Cefazolin	≤ 2 µg/mL	2 g <b>administered</b> every 8 h	N/A	
Ceftaroline	≤ 0.5 µg/mL	600 mg <b>administered</b> every 12 h	N/A	
Cefepime	≤ 2 µg/mL	1 g <b>administered</b> every 12 h	4 µg/mL	1 g <b>administered</b> every 8 h or 2 g <b>administered</b> every 12 h
			8 µg/mL or zone diameter: 19–24 mm	2 g <b>administered</b> every 8 h  (Because it is not possible to correlate specific zone diameters with specific MICs, an isolate with a zone diameter in the SDD range should be treated as if it might be an MIC of 8 µg/mL.)

Big susceptible dose dependent Q & A in Appendix F

# EPIDEMIOLOGICAL CUTOFF VALUE

- MIC values that separate bacterial populations into those that have (don't have) resistance mechanisms strictly on basis of phenotype
- Strictly *in vitro*; no clinical correlation
- Wild type            below ECV  
Non-wild type    above ECV (presumed resistant)



# ECV DETERMINATION

- Only within same species
- Use a CLSI reference method for testing
- $\geq 100$  unique strains;  $\geq 3$  laboratories
- Must be “on-scale”; lowest concentration tested cannot be the mode
- Data can be augmented with molecular profiles

# APPENDIX G; ECV

Table G1. ECVs for *Enterobacteriaceae*

Antimicrobial Agent	Disk Content	Zone Diameter ECV, mm		MIC ECV, µg/mL		Comments
		WT	NWT	WT	NWT	
Azithromycin <sup>1-6</sup>	15 µg	≥ 16	≤ 15	≤ 8	≥ 16	For use with <i>Shigella flexneri</i> .  See Table 2A for azithromycin and <i>Salmonella</i> spp.
	-	-	-	≤ 16	≥ 32	For use with <i>Shigella sonnei</i> .
Colistin	-	-	-	≤ 2	≥ 4	For use with <i>Klebsiella</i> (formerly <i>Enterobacter</i> ) <i>aerogenes</i> , <i>Enterobacter cloacae</i> , <i>Escherichia coli</i> , <i>Klebsiella pneumoniae</i> , and <i>Raoultella ornithinolytica</i> .  The only approved method for testing colistin is MIC by broth microdilution. Disk diffusion and gradient diffusion methods should not be used.  <b>The MICs obtained from testing colistin predict MICs for polymyxin B.</b>

Abbreviations: ECV, epidemiological cutoff value; MIC, minimal inhibitory concentration; NWT, non-wild-type; WT, wild-type.

Table G2. ECVs for Specific Anaerobic Species

Antimicrobial Agent	MIC ECV, µg/mL		Comments
	WT	NWT	
Vancomycin	≤ 2	≥ 4	For use with <i>Cutibacterium</i> (formerly <i>Propionibacterium</i> ) <i>acnes</i> <sup>1-4</sup> and <i>Clostridioides</i> (formerly <i>Clostridium</i> ) <i>difficile</i> . <sup>5,7</sup>

Abbreviations: ECV, epidemiological cutoff value; MIC, minimal inhibitory concentration; NWT, non-wild-type; WT, wild-type.

# APPENDIX H; MOLECULAR ASSAYS

- Strategies for reporting methicillin (oxacillin) results when using molecular/phenotypic AST methods for *S. aureus*
- Strategies for reporting vancomycin results when using molecular/phenotypic AST methods for *Enterococcus* spp.
- Reporting results from ESBL and carbapenemase molecular tests for *Enterobacteriaceae*

# GLOSSARIES

## Glossary I

### Glossary I (Part 1). $\beta$ -Lactams: Class and Subclass Designations and Generic Name

In the late 1990s, several authorities were consulted to construct the glossary. The intention was to include all agents that appeared in M100, along with related agents available for human use. Since that time, agents have been added to the glossary as they were introduced to CLSI, and they do not need to be FDA cleared to be included. It cannot be assumed that the list is exhaustive, and it should be noted that some agents are no longer available for human use.

Antimicrobial Class	Antimicrobial Subclass(es)		Agent(s) Included; Generic Name(s)
Penicillins	Penicillinase-labile penicillins <sup>a</sup>	Penicillin	Penicillin
		Aminopenicillins	Amoxicillin Ampicillin
		Carboxypenicillins	Carbenicillin Ticarcillin
		Ureidopenicillins	Azlocillin
	Penicillinase-stable penicillins <sup>b</sup>		Piperacillin Cloxacillin Dicloxacillin
Aminopenicillin		Nafcillin Oxacillin Medillinam	

## Additions

meropenem-nacubactam  
tebipenem  
imipenem-relebactam

## Deletions

methicillin  
mezlocillin  
norfloxacin

## Glossary II

### Glossary II. Antimicrobial Agent Abbreviation(s), Route(s) of Administration, and Drug Class

In the late 1990s, several authorities were consulted to construct the glossary. The intention was to include all agents that appeared in M100, along with related agents available for human use. Since that time, agents have been added to the glossary as they were introduced to CLSI, and they do not need to be FDA cleared to be included. It cannot be assumed that the list is exhaustive, and it should be noted that some agents are no longer available for human use.

Antimicrobial Agent	Abbreviation(s) <sup>a</sup>	Route(s) of Administration <sup>b</sup>				Drug Class or Subclass
		PO	IM	IV	Topical	
Amikacin	AN, AK, Ak, AMI, AMK		X	X		Aminoglycoside
Amikacin-fosfomicin	AKF	X <sup>c</sup>				Aminoglycoside-fosfomicin



Thank you for your attention.  
Have a good 2019.