



**Wisconsin State  
Laboratory of Hygiene**  
UNIVERSITY OF WISCONSIN-MADISON

# Laboratory-Based Surveillance Plan 2025-2026



**Surveillance program  
overviews, submission  
instructions and 2024-25  
data summaries**



# Welcome to the 2025-26 Season!

Thank you for participating in laboratory-based surveillance in Wisconsin! The participation of clinical laboratories across the state has helped Wisconsin develop one of the most robust surveillance networks in the country!

Supplies for collecting and shipping surveillance specimens, transport to the WSLH, and testing of surveillance specimens is available at **NO COST** when you send specimens using Purple Mountain Solutions. This booklet contains detailed information about the submission of clinical laboratory data and specimens for surveillance.

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## Updates for the 2025-26 Season

**Table 1. Updates to Data and Specimen Submission Requests for the 2025-26 Season**

Pathogen	Surveillance Type	What is Requested
<b>Updated Requests</b>		
<i>Enterobacterales</i> , <i>Pseudomonas aeruginosa</i> and <i>Acinetobacter baumannii</i>	Specimens—Antimicrobial resistance monitoring	Changes to submission criteria (pg.21-22)
<i>Pseudomonas aeruginosa</i>	Specimens—Antimicrobial resistance monitoring	Updated AST profile information (pg. 21-22)
<i>Blastomyces</i>	Specimens-Other Pathogens of Public Health Significance	All positive isolates (pg. 26)
<i>Hepatitis C Virus</i>	Specimens-Other Pathogens of Public Health Significance	All new positives for molecular typing (NGS) (pg. 26)
<b>No Longer Requested</b>		
<i>Influenza</i> , <i>SARS-CoV-2</i> , <i>Rotavirus</i> and <i>RSV</i>	Antigen Testing Data	Number tested and number positive no longer needed (pg. 6)
<i>Aeromonas</i> , <i>Plesiomonas shigelloides</i> , <i>E. coli</i> O157, <i>Entamoeba histolytica</i>	PCR Testing Data	Number tested and number positive no longer needed (pg.6)

## Biosafety Considerations



### Evaluate biosafety at your testing site using a risk assessment:

Review your testing practices and lab environment and consider performing a risk assessment to minimize potential staff exposures when performing testing.

- Use personal protective equipment (e.g. safety glasses, gloves, lab coat)
- An online laboratory risk assessment tool is available here: [www.cdc.gov/safelabs/resources-tools/biosafety-resources-and-tools.html](http://www.cdc.gov/safelabs/resources-tools/biosafety-resources-and-tools.html)



NEW

# New WSLH General Requisition Form!!

NEW

The Communicable Disease Division has recently combined Form A & Form B into an updated **General Requisition Form**. Combining test offerings to a single form and other updates are intended to enhance the accuracy, efficiency, and ease of the test submission process. Please contact WSLH at 1-800-862-1013 for customized requisitions forms.

## WSLH Test Catalog:

Please refer to the Online Test Catalog for any questions regarding test availability, as test list offerings may have changed or been updated. <https://www.slh.wisc.edu/TestDirectory>



## Summary of Requisition Form Updates:

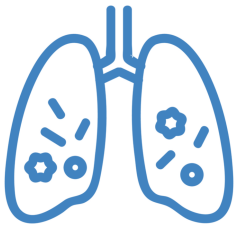
- Combine CDD Req Form A and CDD Req Form B
- Emphasize required information
- Reorganized and defined section grouping
- Reduce field area for less-utilized sections
- Combined Coding & Billing section
- Simplify symptoms

Wisconsin State Laboratory of Hygiene UNIVERSITY OF WISCONSIN-MADISON		Erin C. Rider, Ph.D., D(ABMM), M(ASCP)CM Director of Clinical Laboratory Services 2601 Agriculture Dr, Madison, WI 53718 <a href="http://www.slh.wisc.edu">http://www.slh.wisc.edu</a> Version 1 (6/2025)	CDD Customer Service PHONE: 800-862-1013 FAX: 844-390-6233 KIT/SUPPLIES: 800-862-1088	Communicable Disease Division General Requisition Form 2 - SUBMITTING INSTITUTION
<b>1 - PATIENT INFORMATION*</b> (*) REQUIRED INFORMATION FOR ALL TESTS				
LAST NAME*		FIRST NAME*	MI	DATE OF BIRTH* MM/DD/YYYY
SEX* <input type="checkbox"/> MALE (M) <input type="checkbox"/> FEMALE (F)				
RACE AT BIRTH <input type="checkbox"/> American Indian <input type="checkbox"/> Asian <input type="checkbox"/> Black or African American <input type="checkbox"/> Pacific Islander <input type="checkbox"/> White <input type="checkbox"/> Other				
ETHNICITY <input type="checkbox"/> Hispanic OR Latino <input type="checkbox"/> Not Hispanic or Latino				
PATIENT ADDRESS		CITY	STATE	ZIP
CHART/PATIENT ID		CLINICIAN*	NPI #*	<input type="checkbox"/> HOSPITALIZED <input type="checkbox"/> POSTMORTEM <input type="checkbox"/> PREGNANT
<b>4 - SPECIMEN COLLECTION TYPE/SOURCE*</b>				
<input type="checkbox"/> Blood, Whole <input type="checkbox"/> Cerebrospinal Fluid (CSF) <input type="checkbox"/> Isolate		<input type="checkbox"/> Stool, Cary-Blair/ETM <input type="checkbox"/> Stool, Other <input type="checkbox"/> Swab <input type="checkbox"/> STool PRESERVATIVE		
COLLECTION DATE/TIME*		ISOLATE MEDIA/CONTAINER ISOLATE SOURCE <input type="checkbox"/> Serum <input type="checkbox"/> ACUTE <input type="checkbox"/> CONValesCENT <input type="checkbox"/> Slide/Smear <input type="checkbox"/> Sputum <input type="checkbox"/> SLIDE/SMear SOURCE <input type="checkbox"/> Urine <input type="checkbox"/> Wash/Aspirate <input type="checkbox"/> WASH/ASPIRATE SOURCE		
SUBMITTER SPECIMEN ID*		SWAB SOURCE <input type="checkbox"/> LESION SWABBED? <input type="checkbox"/> YES <input type="checkbox"/> NO <input type="checkbox"/> Tissue <input type="checkbox"/> TISSUE SOURCE		
OTHER TYPE/SOURCE NOT LISTED		PATIENT SIGNATURE		
<b>5 - CLINICAL &amp; EPIDEMIOLOGICAL INFORMATION</b>				
DATE OF ONSET		DATE OF EXPOSURE		DATE OF BIRTH
SYMPTOMS <input type="checkbox"/> Fever <input type="checkbox"/> Headache <input type="checkbox"/> Encephalitis <input type="checkbox"/> Arthralgia <input type="checkbox"/> Myalgia <input type="checkbox"/> Other (specify)		VACCINATION <input type="checkbox"/> Relevant Vaccination <input type="checkbox"/> VACCINATION TYPE <input type="checkbox"/> VACCINATION DATE		TRAVEL <input type="checkbox"/> YES <input type="checkbox"/> NO <input type="checkbox"/> TRAVEL LOCATION <input type="checkbox"/> TRAVEL DATES
DATE OF EXPOSURE		VACCINATION DATE		TRAVEL DATES
<b>6 - TEST SELECTION*</b> SEE THE TEST DIRECTORY FOR FULL LIST & REQUIREMENTS: <a href="https://www.slh.wisc.edu/TestDirectory">https://www.slh.wisc.edu/TestDirectory</a> TEST DIRECTORY QR CODE →				
<input type="checkbox"/> H. Influenzae PCR MP00651 <input type="checkbox"/> Mycobacteria (AFB) Smear AND Culture MH00250 <input type="checkbox"/> Measles Virus PCR - DPH APPROVAL REQUIRED VR01713		<input type="checkbox"/> Mumps Virus PCR VR01714		
<input type="checkbox"/> H. meningitidis PCR MP00561 <input type="checkbox"/> Mycobacteria Isolate Identification MH00253 <input type="checkbox"/> Mumps IgM - Immune Status S502125		<input type="checkbox"/> Rubella Virus PCR - DPH APPROVAL REQUIRED VR01725		
<input type="checkbox"/> Bordetella PCR (pertussis/pseudotuberculosis/tuberculosis) MP00311 <input type="checkbox"/> M. tuberculosis complex (MAC) Susceptibility MH00202 <input type="checkbox"/> Rubella IgG - Immune Status S502135		<input type="checkbox"/> Varicella Zoster Virus PCR & Typing VR01737		
<input type="checkbox"/> Shiga Toxin PCR MP00545 <input type="checkbox"/> M. tuberculosis complex Susceptibility: sst Line Drugs MH00204 <input type="checkbox"/> Measles IgM AND IgG Ab - Diagnostic S502120		<input type="checkbox"/> Measles IgG - Immune Status S502125		
<input type="checkbox"/> Legionella Diagnostic PCR MP00441 <input type="checkbox"/> M. tuberculosis complex PCR DECONTAMINATED? <input type="checkbox"/> YES <input type="checkbox"/> NO S502095 <input type="checkbox"/> Mumps IgM AND IgG Ab - Diagnostic S502130		<input type="checkbox"/> Mumps IgG - Immune Status S502135		
<input type="checkbox"/> Legionella Culture, Diagnostic MP00420 <input type="checkbox"/> M. avium complex (MAC) PCR DECONTAMINATED? <input type="checkbox"/> YES <input type="checkbox"/> NO S502096 <input type="checkbox"/> Rubella IgG - Immune Status S502175		<input type="checkbox"/> Rubella IgG - Immune Status S502165		
<input type="checkbox"/> Legionella Culture, Surveillance MP00420 <input type="checkbox"/> HIV-1/HIV-2 Diagnostic Algorithm S500095 <input type="checkbox"/> HIV-1 Proviral DNA PCR - CALL PRIOR TO SENDING S500091 <input type="checkbox"/> Q Fever IgG Ab S502032		<input type="checkbox"/> Rickettsia IgG Ab (RMSF & Typhus) S502042		
ID & CHARACTERIZATION <input type="checkbox"/> ORGANISM SUSPECTED <input type="checkbox"/> CDT: <input type="checkbox"/> YES <input type="checkbox"/> NO <input type="checkbox"/> ID METHOD: <input type="checkbox"/> Stool Culture, Routine <input type="checkbox"/> Bacterial ID, Enteric <input type="checkbox"/> Bacterial ID, Non-Enteric		HEPATITIS <input type="checkbox"/> Hepatitis A IgM AND IgG Ab - Diagnostic S500020 <input type="checkbox"/> Hepatitis A IgG - Immune Status S500065 <input type="checkbox"/> Hepatitis B Diagnostic Panel (sAb/sAg/sAb) S500083 <input type="checkbox"/> Hepatitis B Immune Status Panel (sAb/sAg) S500085 <input type="checkbox"/> Hepatitis B surface Ab Post-Vaccine (sAb) S500075 <input type="checkbox"/> Hepatitis C Ab w/ reflex to TMA S500069 <input type="checkbox"/> Hepatitis C Virus TMA, Quantitative S500053		
PRE-APPROVAL REQUIRED - CALL 800-862-1013		PATIENT HISTORY OF SYPHILIS: <input type="checkbox"/> YES <input type="checkbox"/> NO <input type="checkbox"/> SYPHILIS Diagnostic Algorithm S502016 <input type="checkbox"/> SYPHILIS VDRL ONLY - Post Treatment S502019 <input type="checkbox"/> SYPHILIS CSF VDRL ONLY (Neurosyphilis) S502018		
SELECT AGENT <input type="checkbox"/> ORGANISM SUSPECTED <input type="checkbox"/> Suspect Select Agent		OTHER TESTS (SPECIFY):		
PARASITOLOGY <input type="checkbox"/> ORGANISM/PARASITE SUSPECTED <input type="checkbox"/> Cryptosporidium/Giardia DFA MP00802 <input type="checkbox"/> Parasite, Surveillance <input type="checkbox"/> Parasite, Blood Smear MP00880		COMPLETE SECTION 5 - SYMPTOMS, ONSET, & TRAVEL <input type="checkbox"/> Arbovirus RT-PCR (DENV/CHIKV/ZIKV) S502110 <input type="checkbox"/> Arbovirus IgM Antibody Panel S502201 <input type="checkbox"/> Dengue Fever IgM, Serology S502272		

# Laboratory Testing Data Submissions

In collaboration with other public health partners, the WSLH has developed data-driven statewide respiratory and gastrointestinal pathogen surveillance programs in Wisconsin. Weekly reporting of diagnostic testing data to WSLH is important so that public health partners know what communicable diseases are impacting community health in Wisconsin.

## Wisconsin Respiratory Surveillance Program:



The aim of this program is to gain situational awareness of the respiratory pathogens circulating in Wisconsin, including geographic spread and positivity rates. **All Sites** in Wisconsin performing PCR/molecular and/or antigen testing for respiratory pathogens are asked to submit their testing data on a **weekly basis** (number tested, number positive) **all year round**. Refer to Table 2 for a list of pathogens for which testing data is requested.

## Wisconsin Acute Diarrheal Illness Surveillance Program:



The aim of this program is to gain awareness of the gastrointestinal pathogens effecting community health in Wisconsin. The WSLH requests that clinical labs submit gastropathogen PCR testing data for bacterial, parasitic and viral pathogens on a **weekly basis** (number tested, number positive), **all year round**. Refer to Table 2 for a list of pathogens for which testing data is requested.

## Why Submit Data?

### To determine geographic spread:

Testing numbers from around the state are broken down by region to monitor trends for respiratory pathogens in each part of the state. Regional graphs are updated weekly and can be found on the WSLH website.

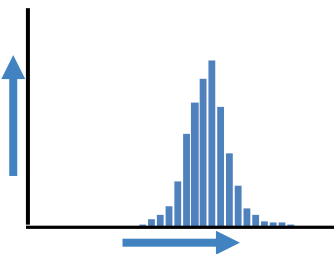
[www.slh.wisc.edu/wcln-surveillance/surveillance/virology-surveillance/](http://www.slh.wisc.edu/wcln-surveillance/surveillance/virology-surveillance/)



### To provide situational awareness:

Testing data is used to determine when the season begins, peaks and ends for a variety of pathogens. The data can be used to identify outbreaks, detect an issue with test methods, inform medical provider differential diagnosis, and inform clinical lab staffing needs for seasonal and outbreak testing. Graphs of surveillance data are updated weekly and can be found on the WSLH website.

[www.slh.wisc.edu/wcln-surveillance/surveillance/](http://www.slh.wisc.edu/wcln-surveillance/surveillance/)



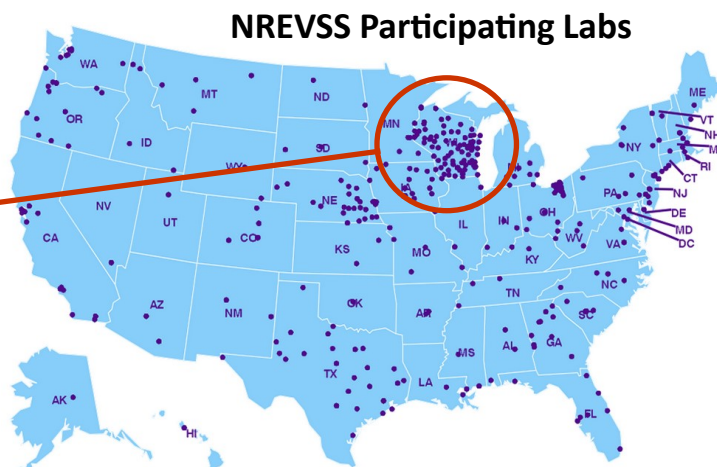
## Why Submit Data?

### To participate in the National Respiratory and Enteric Virus Surveillance System (NREVSS)

WSLH collates all testing data collected from Wisconsin and submits it to NREVSS on a weekly basis. NREVSS is a laboratory-based system that monitors temporal and geographic circulation of a variety of viral pathogens. NREVSS data can be viewed on the CDC website at:

[www.cdc.gov/surveillance/nrevss/](http://www.cdc.gov/surveillance/nrevss/)

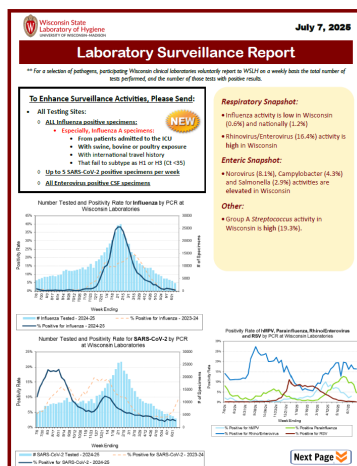
Wisconsin leads the country for the number of labs participating in surveillance activities.



### To participate in national influenza surveillance:

Influenza testing data is submitted to the CDC for inclusion into the national influenza surveillance program. Influenza data can be found on CDC's FLUVIEW website at: [www.cdc.gov/fluview/](http://www.cdc.gov/fluview/)

## Wisconsin Surveillance Data Distribution:



### WSLH Website:

Aggregate data for a variety of pathogens can be found on the WSLH website at: [www.slh.wisc.edu/wcln-surveillance/surveillance/](http://www.slh.wisc.edu/wcln-surveillance/surveillance/)

### WSLH Bi-Weekly Surveillance Report:

The WSLH aggregates all respiratory and enteric pathogen data and provides summary reports in the bi-weekly Laboratory Surveillance Report, which is distributed to over 600 recipients across the state. Email [WCLN@slh.wisc.edu](mailto:WCLN@slh.wisc.edu) to sign up for this report.

### WCLN Laboratory Messages:

The WSLH provides laboratory guidance and general updates through Wisconsin Clinical Laboratory Network (WCLN) Laboratory Messages. If you would like to receive these messages, please email [wcln@slh.wisc.edu](mailto:wcln@slh.wisc.edu) and include "WCLN Lab Message Sign-up" in the subject line.

### WDHS Respiratory Illness Dashboard

A data summary of respiratory pathogens is also sent to the Wisconsin Department of Health Services for inclusion into their interactive **Respiratory Illness Dashboard**. [www.dhs.wisconsin.gov/disease/respiratory-data.htm](http://www.dhs.wisconsin.gov/disease/respiratory-data.htm)



# Data Submission Requests

The WSLH requests that **all clinical labs** in Wisconsin performing PCR/molecular and/or rapid antigen testing for the following bacterial, parasitic and viral pathogens submit testing data (number tested and number positive) on a **weekly basis, all year round**.

**Table 2. Laboratory Testing Data Requests**

## Antigen Detection

**Rapid Strep (Group A *Streptococcus*)**

## Respiratory Pathogens - PCR/Molecular Detection

<b>Influenza A/B</b>	<b>SARS-CoV-2</b>	<b>RSV</b>
<b>Seasonal Coronaviruses</b>	<b>Human Metapneumovirus</b>	<b>Human Parainfluenza virus</b>
<b>Rhinovirus/Enterovirus</b>	<b>Adenovirus</b>	<b><i>B. pertussis</i> and <i>parapertussis</i></b>
<b>Group A <i>Streptococcus</i></b>		

## Gastrointestinal Pathogens - PCR/Molecular Detection

<b><i>Campylobacter</i></b>	<b><i>Clostridioides difficile</i></b>	<b><i>Salmonella</i></b>
<b><i>Shigella</i>/ Enteroinvasive <i>E.coli</i> (EIEC)</b>	<b>Shiga-like toxin-producing <i>E. coli</i> (STEC)</b>	<b><i>Vibrio</i></b>
<b><i>Yersinia enterocolitica</i></b>	<b>Adenovirus 40/41</b>	<b>Astrovirus</b>
<b>Norovirus</b>	<b>Rotavirus</b>	<b>Sapovirus</b>
<b><i>Cryptosporidium</i></b>	<b><i>Cyclospora cayetanensis</i></b>	<b><i>Giardia lamblia</i></b>

# Submission of Laboratory Testing Data

- **Access** the web reporting webpage:
  - Go to the WSLH website <http://www.slh.wisc.edu/wcln-surveillance/surveillance/> then click on ***“Click here to report Wisconsin Test Data”*** in the center of the page.



**OR**

For more information regarding reportable diseases, please see the following:

- Go directly to <http://www.surveymoz.com/s3/389222/Wisconsin-Laboratory-Surveillance-Reporting>
- **Enter** your laboratory’s identification number (***“Institution ID”***); ***this is a required field.***
  - Your ***“Institution ID”*** is a series of letters (must be capitalized) and numbers (eg. LRN789). It has been added to your customized requisition form that is included with this mailing.
    - If you cannot find your ***“Institution ID”***, please contact us at [WCLN@slh.wisc.edu](mailto:WCLN@slh.wisc.edu) or call 800-862-1013.
  - Your institution’s name, address, city and telephone number will be entered automatically.
- **Review** the institution name, address and telephone number for accuracy.
  - If any of the information is not accurate, enter the correct information and check the box at the bottom of the form ***“Check here if any pre-filled information on this page was changed”***.

## Wisconsin Laboratory Surveillance Reporting

Survey Gizmo - Wisconsin Laboratory Surveillance Report - Antigen Detection

Please select the reporting week, the number of specimens tested, the number positive, and the test used for the agents listed below. Click "Finished? Submit your Survey" when finished. Press tab to move between fields. Please email [wcln@slh.wisc.edu](mailto:wcln@slh.wisc.edu) with questions or corrections.

**Institution ID \***

LRN000

### Institution Information

**Institution Name**

Wisconsin State Lab

**Street Address**

2601 Agriculture Dr

**City**

Madison

**State**

WI

**Zip Code**



## Submission of Laboratory Testing Data (cont'd)

- Select the “**Week Ending Date**” for which you wish to report data.
  - Click on the date in the drop-down list. ***This is a required field.***

*It is critical that you select the correct week ending date!*

### Week Ending - **2025/2026 Season** (Saturday)

Reporting week is Sunday to Saturday

-- Please Select --

- Check either “Antigen Detection” or “PCR / Molecular” for the testing method for the data you would like to enter and click “Next”.

Select the method below to enter data; you must also select “Next”.

- ☐ Antigen Detection
- ☐ PCR / Molecular

Back Next

17%

### If you chose Antigen Detection:

- **Report** the number tested and number positive for each of the listed agents for which you perform testing on-site.
- **If you do not perform a test on-site and/or refer specimens to another laboratory**, skip that agent/test section without entering any data.
- **If you normally perform that testing on site, but did not test any specimens** that week, enter zero “0” for the number tested. If the “number tested” is “0”, you can skip the “number positive” field.
  - ***If you discontinue testing for a season (summer), please notify us so that our data accurately reflects testing in Wisconsin.***
- **Review** for accuracy the test(s) that have been pre-marked for your institution.

Strep Test Used: Please check all that apply

- |  |  |
|--|--|
| <input type="checkbox"/> Acceava Strep A             | <input type="checkbox"/> OSOM Ultra Strep A        |
| <input type="checkbox"/> BD Chek Group A Strep       | <input type="checkbox"/> QuickVue Dipstick Strep A |
| <input type="checkbox"/> BinaxNOW Strep A            | <input type="checkbox"/> QuickVue In-Line Strep A  |
| <input type="checkbox"/> Clearview Strep A Exact II  | <input type="checkbox"/> QuickVue+ Strep A         |
| <input type="checkbox"/> CONSULT Strep A Dipstick    | <input type="checkbox"/> SAS StrepAlert            |
| <input type="checkbox"/> Directigen EZ Group A Strep | <input type="checkbox"/> Signify Strep A Cassette  |
| <input type="checkbox"/> ICON SC Strep A             | <input type="checkbox"/> Signify Strep A Dipstick  |
| <input type="checkbox"/> ImmunoCard STAT! Strep A    | <input type="checkbox"/> Sure-Vue Strep A          |
| <input type="checkbox"/> McKesson Strep A Twist      | <input type="checkbox"/> Other (specify):          |
| <input type="checkbox"/> OSOM Strep A                | <input type="text"/>                               |

## Submission of Laboratory Testing Data (cont'd)

- If the marked test is **NOT** the test your facility used, click on the marked test to “un-check” it, then click on the correct test.
  - Please check the box at the bottom of the form “**Check the box below if any pre-filled information on this page was changed.**”

Check the box below if any pre-filled information on this page was changed.

☐ Pre-filled information was changed.

- Check “PCR / Molecular” to enter more data or check “Finished entering data” to finish, and then click “Next”.

Select the method below to continue entering data or select “Finished entering data” if done; you must also select “Next”.

- ☐ PCR / Molecular
- ☐ Finished entering data

Back

Next

33%

### If you chose PCR/Molecular :

- Report** the number tested and number positive for each of the listed agents for which you provide testing on-site.
- If you do not perform a test on-site and/or refer specimens to another laboratory**, skip that agent/test section without entering any data.
- If you normally perform that testing on site, but did not test any specimens** that week, enter zero “0” for the number tested. If the “number tested” is “0”, you can skip the “number positive” field.

### Respiratory Pathogens PCR testing

Please report the number of specimens tested and the number positive.

	Number Tested	Number Positive
Adenovirus	<input type="text"/>	<input type="text"/>
Bocavirus	<input type="text"/>	<input type="text"/>
Chlamydia pneumonia	<input type="text"/>	<input type="text"/>
Coronavirus 229E	<input type="text"/>	<input type="text"/>
Coronavirus HKU1	<input type="text"/>	<input type="text"/>
Coronavirus NL63	<input type="text"/>	<input type="text"/>



## Submission of Laboratory Testing Data (cont'd)

- For each molecular/PCR testing that your laboratory performs, **“check”** the test(s) your laboratory used.

Gastrointestinal Pathogen PCR Test Used: Please check all that apply

- 
- |   |  |
|---|--|
| <input type="checkbox"/> FilmArray Gastrointestinal Panel (BioFire) | <input type="checkbox"/> xTAG Gastrointestinal Panel PCR (GPP) (Luminex) |
| <input type="checkbox"/> MAX Enteric Bacteria Panel (BD)            | <input type="checkbox"/> Verigene Enteric Pathogen Test (Nanosphere)     |
| <input type="checkbox"/> Prodesse ProGastro SSCS (Hologic/GenProbe) | <input type="checkbox"/> Other (specify):                                |

- Check “Antigen Detection” to enter more data or check “Finished entering data” to finish.
  - Click “Next”.

**Select the method below to continue entering data or select “Finished entering data” if done; you must also select “Next”.**

- ☐ Antigen Detection  
☐ Finished entering data

Back Next

50%

### If you chose “Finished entering data”:

- To save and submit your data, click on “Submit”. The data you entered will not be saved or transmitted until you click “Submit”.

Wisconsin Laboratory Surveillance Reporting  
Thank you!

Thank You!


Thank you for your report!

If you have any questions or updates, please email [wcln@slh.wisc.edu](mailto:wcln@slh.wisc.edu)

To go to WSLH Web site: <http://www.slh.wisc.edu/>

Back Submit

86%



- If you want to report data for another week, return to <http://www.slh.wisc.edu/wcln-surveillance/surveillance/> and repeat the data entry process for the new week, starting with entering your Institution ID.

**If you have questions please email [WCLN@slh.wisc.edu](mailto:WCLN@slh.wisc.edu) or call  
WSLH customer service at 800-862-1013**

# Respiratory Pathogen Surveillance

Laboratory-based surveillance for influenza, SARS-CoV-2, and other respiratory pathogens is coordinated by the Wisconsin State Laboratory of Hygiene (WSLH), in collaboration with the Wisconsin Division of Public Health and the Centers for Disease Control and Prevention (CDC). This surveillance network utilizes a combination of laboratory testing data and respiratory specimens submitted to the WSLH for additional surveillance testing and characterization. Requests for influenza positive specimens are modified as the level of influenza activity changes and as other circumstances require. These changes will be communicated in the bi-weekly “Laboratory Surveillance Report” which is posted at the WSLH website (<http://www.slh.wisc.edu/wcln-surveillance/surveillance/>). Requests for other respiratory pathogens are static throughout the year and can be found in Table 3 on page 14.

The following is a description of the contributing elements of the laboratory-based respiratory surveillance plan for Wisconsin:

## Data Submissions:



**All Sites** in Wisconsin performing PCR/molecular testing for respiratory pathogens are asked to submit their testing data on a weekly basis (number tested, number positive) as described in Table 2 found on page 6.

## Specimen Submissions:



**All Sites:** Any Clinical Laboratory in Wisconsin performing testing for respiratory pathogens is requested to submit influenza, SARS-CoV-2, *Legionella* and TB positive specimens to the WSLH! The WSLH requests influenza and SARS-CoV-2 positive specimens from clinical laboratories performing testing using antigen and/or molecular PCR methods as described in Table 3. In addition, these sites are asked to submit isolates or lower respiratory specimens (BAL, Sputum) from patients that test positive for *Legionella* by culture or urine antigen testing, and all TB isolates.



**Enrolled Sentinel Surveillance Sites:** 17 labs throughout the 5 public health regions of Wisconsin are enrolled as Sentinel Surveillance sites. This surveillance network is designed to provide a consistent and randomized supply of respiratory specimens from all areas of the state. These sites provide the first 3 specimens per week from patients presenting with respiratory symptoms (regardless of initial test results) as described in Table 3 (page 14).



**University Health Clinics:** This surveillance program is used to monitor respiratory pathogens impacting student health, including influenza, SARS-CoV-2, and adenovirus. University Health sites are requested to submit the first 3 specimens per week from patients presenting with respiratory symptoms (regardless of initial test results) as described in Table 3 (page 14). These specimens are tested at WSLH with an enhanced respiratory pathogen panel.

## What Do We Do With These Specimens?

Respiratory surveillance specimens submitted to the WSLH for viral surveillance are tested with the CDC Influenza/SARS-CoV-2 multiplex RT-PCR assay.

### Influenza Positive Specimens:

- WSLH performs the CDC influenza A subtyping and CDC B lineage PCRs on all influenza positive specimens received. Subtyping of influenza specimens from around Wisconsin monitors for the emergence of avian and novel influenza in humans.
- Whole genome sequencing is performed on a selection of influenza positive specimens received at the WSLH.
- WSLH submits a selection of influenza positive specimens from around the state to the national influenza surveillance pipeline. Viral culture and whole genome sequencing is performed on these specimens before they are submitted to the CDC.

#### Detect novel influenza viruses!

Subtyping is important for the detection of any variant or emerging influenza viruses with pandemic potential.



#### Participate in national influenza surveillance!

CDC uses influenza positive specimens to inform vaccine strain selection and provide vaccine candidates.



### SARS-CoV-2 Positive Specimens:

- Whole genome sequencing is performed on SARS-CoV-2 positive specimens received at the WSLH.
- WSLH also submits specimens to the National SARS-CoV-2 Strain Surveillance (NS3) program to enhance national surveillance efforts. This program provides information guides anti-viral recommendations and vaccine selection.

#### Perform genomic surveillance!



Sequencing is useful to detect novel viral variants, monitor antiviral resistance, and aid in outbreak investigations.

### Specimens Negative for Influenza and SARS-CoV-2:

- An enhanced respiratory pathogen panel is performed on a selection of surveillance specimens. This data is submitted to NREVSS and can aid public health personnel in determining which respiratory pathogens are circulating throughout the year.

#### Monitor for other respiratory pathogens!

Testing with an enhanced respiratory panel provides positivity data for less-commonly tested for respiratory pathogens.



### Respiratory Specimens or Isolates from Patients with Legionella:

- These specimens/isolates are cultured and their identity confirmed with PCR. Confirmed positive isolates may undergo whole genome sequencing that can help identify the source of the outbreak.

## Respiratory Specimen Submission Instructions:

For each viral surveillance specimen submitted, fill out a copy of your customized respiratory surveillance requisition form (included with your mailing packet). Please be sure to check the appropriate reason for submission on the requisition form.

For *Legionella* submissions, please fill out a WSLH General requisition form and check “*Legionella* Culture, Surveillance” (MP00420).

### Specimen Collection:

All upper and lower respiratory specimens are acceptable for viral surveillance testing. Upper respiratory specimens should be collected with Dacron or polyester tipped plastic shafted swabs and placed in virus transport medium (VTM).

- If the swab was placed in virus transport medium or saline before a portion was removed for testing, submit remaining swab in virus transport medium.
- If the swab was collected and immersed in test reagent, collect a second sample for surveillance testing.

When selecting specimens to submit for influenza surveillance, please prioritize specimens from patients who are hospitalized, have international travel history or have had recent contact with cattle, swine or poultry.



Any clinical specimen that produces a positive result for influenza A by PCR, but fails to subtype as H3 or (H1N1)pdm09, has the potential to be a novel strain of influenza virus. **Repeat Testing is recommended in these situations.** If the influenza A specimen fails to subtype after repeat testing **AND** the influenza A Ct is <35, please send the sample to the WSLH for further testing.

### Order:

Additional specimen requisition forms and supplies for collecting and shipping surveillance specimens are provided at **NO COST** and can be requested at 800-862-1088.

### Ship:

Transport to the WSLH and testing of surveillance specimens is available at **NO COST** when you send specimens using **Purple Mountain Solutions** (PH: 800-990-9668). Detailed shipping instructions can be found on pages 27-28 of this packet.



## Respiratory Specimen Submission Instructions:

**Table 3: Respiratory Specimen Submission Requests**

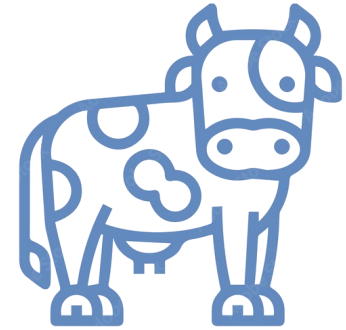
Pathogen Specific Respiratory Surveillance		
Influenza Surveillance		
Submitter Testing Method:	Season	
	Off Season (June-October)	Influenza Season (Winter/Spring*)
PCR/Molecular OR Antigen	<u>ALL</u> influenza positive specimens	One influenza-related hospitalization per week AND Unsubtypable influenza A positives (If subtyping is attempted, Ct < 35) AND Influenza A positive specimens with: <ul style="list-style-type: none"><li>• International travel history</li><li>• Bovine, swine or avian exposure</li></ul>
SARS-CoV-2 Surveillance		
PCR/Molecular OR Antigen	Five positive SARS-CoV-2 samples per week for genomic surveillance	
Legionella Surveillance		
Culture, PCR OR Urine Antigen	Specimens from all <i>Legionella</i> positive patients: <ul style="list-style-type: none"><li>• Sputum or BAL from Urine Antigen positive patients (NOT URINE)</li><li>• Isolates</li></ul>	
Mycobacteria Surveillance		
The WSLH requests that clinical laboratories submit one <i>Mycobacterium tuberculosis</i> complex (MTBC) isolate from each patient identified as having a MTBC infection.		
Site Specific Respiratory Surveillance		
University Health	The first 3 respiratory specimens per week from symptomatic patients (regardless of initial test results, all year round)	
Sentinel Surveillance	The first 3 respiratory specimens per week from symptomatic patients (regardless of initial test results, all year round)	

\*Refer to the WSLH bi-weekly "Laboratory Surveillance Report" for current influenza activity in Wisconsin

# Avian Influenza Update

In the spring of 2022, H5N1 avian influenza emerged in wild birds worldwide, including in North America. This outbreak continues, and has affected many wild birds and poultry farms in Wisconsin and across the US.

On March 25, 2024, H5N1 avian influenza was detected in a dairy cow in Texas. Since that first detection, H5N1 avian influenza has been found in almost 1,000 cattle herds across 16 states. On April 1, 2024, the Centers for Disease Control and Prevention (CDC) announced that H5N1 avian influenza had been detected in a person with exposure to dairy cattle in Texas. Since then, an additional 70 human cases of H5N1 have been detected in the U.S. including one fatality.



Although CDC's Risk Assessment for the general population remains LOW, additional precautions are being taken to enhance influenza surveillance across the country. In Wisconsin, there have been no documented dairy cattle cases of avian influenza. People who have close contact with poultry or dairy cattle infected with H5N1 avian influenza are at increased risk for infection.

## Clinicians who suspect influenza virus infection in symptomatic patients with known exposure to the following should contact the **Wisconsin Division of Public Health (WDPH)**

- **Ill or dying animals or their products**
- **Confirmed H5N1 positive flock/herd**
- **Person with a probable or confirmed case of H5N1**

- ◆ ***Weekdays during business hours — call 608-267-9003***
- ◆ ***After-hours — call 608-258-0099 and ask for “the communicable disease epidemiologist on-call”***

### **Specimen Collection and Submission:**

- Collect **one nasopharyngeal (NP) swab** and place in a tube of viral transport medium (VTM).
- For patients with **conjunctivitis**, additionally, obtain a conjunctival swab and place in a tube of VTM.
  - Use swabs with a Dacron or polyester tip with an aluminum or plastic shaft.
- For patients with lower respiratory illness, collect a lower respiratory tract specimen (e.g. BAL).
- After gaining approval from WDPH, an “Enhanced Surveillance” requisition form should be requested from the WSLH, and filled out for each submitted specimen.
  - **Please check “Avian Influenza Suspect” under reason for submission.**

## Avian Influenza Update (Cont'd)

### Shipping of Avian Influenza Suspect Specimens to WSLH

- Arrange transport so that specimens arrive at the WSLH within 24 hours of collection.
  - Avian influenza suspects can be shipped Category B.
  - Specimens can be transported to the WSLH by a carrier of your choice, or at NO COST when using Purple Mountain Solutions (PH: 800-990-9668).
  - Detailed shipping instructions can be found on pages 27-28 of this packet.
- Testing is usually completed within 1-2 business days of the specimen receipt at WSLH. There is no charge for this testing.
- Please contact the **WSLH Customer Service Department** at 800-862-1013 if you have questions regarding laboratory testing.



### Suspect Avian Influenza Testing at the WSLH

- Suspect avian influenza specimens that are submitted to the WSLH for testing **will first be tested with the CDC influenza A/influenza B/SARS-CoV-2 multiplex PCR assay.**
- If a specimen is positive for influenza A, it will also be **tested with a seasonal influenza A subtyping PCR.**
- If a specimen is **positive for influenza A and “unable to subtype”** with the seasonal influenza A subtyping PCR, then it will be tested with the **influenza A – H5 specific subtyping PCR.**
- Visit the WSLH Website for additional information:  
<https://www.slh.wisc.edu/clinical/diseases/h5n1-highly-pathogenic-avian-influenza-update/>

## Influenza A Swine Variant Virus Testing

Influenza viruses from swine normally do not infect humans; however, cases do occur sporadically in humans and are typically associated with close contact with infected pigs. There have been no reports of sustained human transmission of these variant viruses as of June 2025.

**Clinicians who suspect influenza virus infection in patients with close contact to swine should contact the Wisconsin Division of Public Health at 608-258-0099 for subtype-specific PCR testing at WSLH.**

### Specimen Submission:

- Specimen requirements for Influenza A swine variant suspects are the same as described on page 13.
- After gaining approval from WDPH, an “Enhanced Surveillance” requisition form should be requested from the WSLH, and filled out for each submitted specimen.
  - **Please check “Swine Contact” under reason for submission.**
- Please contact the **WSLH Customer Service Department** at 800-862-1013 if you have questions regarding laboratory testing.





# Wisconsin Mycobacteriology Surveillance

WSLH leads the state-wide surveillance and antimicrobial susceptibility testing program for *Mycobacterium tuberculosis* complex (MTBC) specimens identified in Wisconsin, with the support of the WI Mycobacteriology Laboratory Network (WMLN). The WMLN was developed in 1998 with a goal to assure excellence in TB laboratory testing, support TB control efforts by the WI Tuberculosis Program through the state Division of Public Health, and drive surveillance for non-tuberculous mycobacteria (NTM) identified among WI residents. The WMLN provides a laboratory-based mechanism for monitoring the incidence of *M. tuberculosis* isolation, drug resistance rates and works to ensure that mycobacteriology testing in Wisconsin is meeting the needs of Wisconsin residents.



## Mycobacterium Tuberculosis Complex (MTBC) Surveillance:

- The WSLH requests that clinical laboratories submit one MTBC isolate from each patient identified as having a MTBC infection.
  - WSLH performs rapid rifampin susceptibility testing and phenotypic susceptibility testing for first-line MTBC antimicrobials on one isolate from each patient.
  - In conjunction with the WI TB Program, WSLH ensures that any additional susceptibility testing necessary will be performed at the Centers for Disease Control and Prevention or one of our other national partner laboratories.
  - WSLH performs genotyping on one isolate from each MTBC-positive patient. Genotype data, coupled with the CDC Tuberculosis Genotyping Information Management System (TB-GIMS), helps identify state- and nation-wide TB transmission links.
  - WSLH provides a state repository for MTBC isolates for the purpose of universal drug susceptibility testing and genotyping.

## Non-Tuberculous Mycobacteria Surveillance:

- WSLH organizes surveillance for non-tuberculous mycobacteria (NTM) identified from Wisconsin residents.
  - WSLH requests that all clinical laboratories performing mycobacteria testing (AFB smear/culture, mycobacteria-specific NAAT testing, mycobacterial identification) submit NTM isolation **data** to the WMLN Coordinator annually.
  - WSLH generates an annual NTM isolation report which is distributed to network members and state and local public health departments.
  - Clinical laboratories interested in participating in NTM surveillance or receiving the annual WMLN Mycobacteria Surveillance Report can contact WSLH at 800-862-1013 for more information.



## Specimen Submissions:

### Specimen submission instructions:

WSLH requests that clinical laboratories submit MTBC isolates for universal drug susceptibility testing and genotyping. **MTBC isolates MUST be submitted using Category A packaging and shipping requirements.**

- Fill out a WSLH General Requisition Form with each specimen submitted
  - For isolates for which susceptibility testing **has not been performed** at another reference laboratory, please select test *M. tuberculosis* Susceptibility: 1<sup>st</sup> - Line Drugs [MM00204].
  - For isolates for which susceptibility testing **has already been performed and reported to WI DHS/WI TB Program**, please select test *M. tuberculosis* genotyping [MM02881].
- Specimen requisition forms and supplies for shipping surveillance specimens are provided at **NO COST** and can be requested at 800-862-1088.
- Transport to the WSLH is available at **NO COST** when you send specimens using **Purple** Mountain Solutions (PH: 800-990-9668). Detailed shipping instructions can be found on pages 27-28.



MYCOBACTERIOLOGY	<input type="checkbox"/> Mycobacteria (AFB) Smear <b>AND</b> Culture	MM00250
	<input type="checkbox"/> Mycobacteria Isolate Identification	MM00253
	<input type="checkbox"/> <i>M. avium</i> complex (MAC) Susceptibility	MM00202
	<input type="checkbox"/> <i>M. tuberculosis</i> complex Genotyping	MM02881
	<input type="checkbox"/> <i>M. tuberculosis</i> complex Susceptibility: 1st Line Drugs	MM00204
	<input type="checkbox"/> <i>M. tuberculosis</i> complex PCR DECONTAMINATED? <input type="checkbox"/> YES <input type="checkbox"/> NO SMEAR RESULT:	MM00256
	<input type="checkbox"/> <i>M. avium</i> complex (MAC) PCR DECONTAMINATED? <input type="checkbox"/> YES <input type="checkbox"/> NO SMEAR RESULT:	MM00260

**Wisconsin State Laboratory of Hygiene**  
UNIVERSITY OF WISCONSIN-MADISON

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KITS/SUPPLIES: 800-862-1088

Communicable Disease Division  
General Requisition Form  
2 - SUBMITTING INSTITUTION

1 - PATIENT INFORMATION\* (\* REQUIRED INFORMATION FOR ALL TESTS)

LAST NAME\* FIRST NAME\* MI DATE OF BIRTH\* MM/DD/YYYY  
SEX\* ☐ MALE (M) ☐ FEMALE (F)  
AT BIRTH  
RACE/ETHNICITY ☐ American Indian ☐ Asian ☐ Black or African American ☐ Pacific Islander ☐ White ☐ OTHER, RACE

PATIENT ADDRESS CITY STATE ZIP ☐ HOSPITALIZED ☐ POSTMORTEM ☐ PRENATAL  
CHART/PATIENT ID CLINICIAN\* NPI #

4 - SPECIMEN COLLECTION TYPE/SOURCE\*  
COLLECTION DATE/TIME\*  
SUBMITTER SPECIMEN ID\*  
OTHER TYPE/SOURCE NOT LISTED

5 - CLINICAL & EPIDEMIOLOGICAL INFORMATION  
DATE OF EXPOSURE  
SYMPTOMS  
DATE OF ONSET  
DATE OF EXPOSURE  
SYMPTOMS  
DATE OF ONSET

6 - TEST SELECTION  
SEE THE TEST DIRECTORY ON THE RIGHT  
Mycobacteria (AFB) Smear AND Culture  
Mycobacteria Isolate Identification  
M. avium complex (MAC) Susceptibility  
M. tuberculosis complex Genotyping  
M. tuberculosis complex Susceptibility: 1st Line Drugs  
M. tuberculosis complex PCR DECONTAMINATED? ☐ YES ☐ NO SMEAR RESULT:  
M. avium complex (MAC) PCR DECONTAMINATED? ☐ YES ☐ NO SMEAR RESULT:

7 - CODING & BILLING  
ICD-10 CODES ARE REQUIRED FOR THIRD-PARTY PAYERS BILLING  
ICD-10 (A) ICD-10 (B) ICD-10 (C) ICD-10 (D)  
ICD-10 (E) ICD-10 (F) ICD-10 (G) ICD-10 (H) ICD-10 (I) ICD-10 (J) ICD-10 (K) ICD-10 (L) ICD-10 (M) ICD-10 (N) ICD-10 (O) ICD-10 (P) ICD-10 (Q) ICD-10 (R) ICD-10 (S) ICD-10 (T) ICD-10 (U) ICD-10 (V) ICD-10 (W) ICD-10 (X) ICD-10 (Y) ICD-10 (Z)  
ICD-10 (A) ICD-10 (B) ICD-10 (C) ICD-10 (D) ICD-10 (E) ICD-10 (F) ICD-10 (G) ICD-10 (H) ICD-10 (I) ICD-10 (J) ICD-10 (K) ICD-10 (L) ICD-10 (M) ICD-10 (N) ICD-10 (O) ICD-10 (P) ICD-10 (Q) ICD-10 (R) ICD-10 (S) ICD-10 (T) ICD-10 (U) ICD-10 (V) ICD-10 (W) ICD-10 (X) ICD-10 (Y) ICD-10 (Z)

TEST DIRECTORY OR CODE  
Mycobacteria (AFB) Smear AND Culture  
Mycobacteria Isolate Identification  
M. avium complex (MAC) Susceptibility  
M. tuberculosis complex Genotyping  
M. tuberculosis complex Susceptibility: 1st Line Drugs  
M. tuberculosis complex PCR DECONTAMINATED? ☐ YES ☐ NO SMEAR RESULT:  
M. avium complex (MAC) PCR DECONTAMINATED? ☐ YES ☐ NO SMEAR RESULT:

8 - OTHER TESTS (SPECIAL)  
HIV-1/2 Diagnostic Algorithm  
HIV-1 Proviral DNA PCR - CALL PRIOR TO SENDING  
Chlamydia AND Gonorrhea TMA  
Chlamydia trachomatis ONLY  
Neisseria gonorrhoeae ONLY  
Trichomonas vaginalis TMA  
Mycoplasma genitalium TMA  
Herpes Simplex Virus 1 & 2 TMA - LESION  
PATIENT HISTORY OF SYPHILIS ☐ YES ☐ NO  
Syphilis Diagnostic Algorithm  
Syphilis VDRL ONLY - Post Treatment  
Syphilis CSF VDRL ONLY (Neurosyphilis)

9 - COMPLETE SECTION 5 - SYMPTOMS, ONSET, & TRAVEL  
Arbovirus RT-PCR (JEV/CHIKV/DENV)  
Arbovirus IgM Antibody Panel  
Dengue Fever IgM, Serology

# Wisconsin Enteric Pathogen Surveillance

The WSLH requests that clinical laboratories submit isolates, enrichment broths, or stools in enteric transport medium that were positive using a culture-independent diagnostic test (CIDT). These isolates and samples undergo testing such as whole-genome sequencing at the WSLH to generate data that is critical to the ability to recognize and respond to clusters and outbreaks of gastropathogens in Wisconsin. The resulting laboratory data is used by epidemiologists at the WDPH to rapidly determine linkage to potential food and environmental point sources.



## Specimen submission instructions:

WSLH requests that clinical laboratories submit the pathogens listed in Table 4 **As Detected**.

- Fill out a **WSLH General requisition form** with each specimen submitted:
  - For all enteric submissions, indicate the **Organism(s) Suspected**, whether a CIDT (Culture independent diagnostic test) was used and the ID method performed.
    - For isolates, select **Bacterial ID, Enteric**
    - For Stool, select **Stool Culture, Routine**
- Specimen requisition forms and supplies for shipping surveillance specimens are provided at **NO COST** and can be requested at 800-862-1088.
- Transport to the WSLH is available at **NO COST** when you send specimens using **Purple Mountain Solutions** (PH: 800-990-9668). Detailed shipping instructions can be found on pages 27-28.



Wisconsin State  
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Communicable Disease Division  
General Requisition Form  
2 - SUBMITTING INSTITUTION

BACTERIOLOGY

ID & CHARACTERIZATION

- ☐ Shiga Toxin PCR MP00545
- ☐ Shiga Toxin EIA MP00549
- ☐ Legionella Diagnostic PCR MP00441
- ☐ Legionella Culture, Diagnostic MP00420
- ☐ Legionella Culture, Surveillance

ORGANISM SUSPECTED

CIDT: ☐ YES ☐ NO ID METHOD:

- ☐ Stool Culture, Routine
- ☐ Bacterial ID, Enteric
- ☐ Bacterial ID, Non-Enteric

### 1 - PATIENT INFORMATION\* (\*) REQUIRED INFORMATION FOR ALL TESTS

LAST NAME\* FIRST NAME\* MI DATE OF BIRTH\* MM/DD/YYYY  
SEX\* ☐ MALE (M) ☐ FEMALE (F)  
RACE ☐ American Indian ☐ Asian ☐ Black or African American ☐ Pacific Islander ☐ White ☐ OTHER, RACE  
ETHNICITY ☐ Hispanic OR Latino ☐ Not Hispanic or Latino

PATIENT ADDRESS CITY STATE ZIP  
CHART/PATIENT ID CLINICIAN\* NPI #\* ☐ HOSPITALIZED ☐ POST-INTENSIVE ☐ PRESIDENT

4 - SPECIMEN COLLECTION TYPE/SOURCE\*  
Blood, Whole ANTI-CLONALANT Stool, Cary-Blair/ETM  
Cerebrospinal Fluid (CSF) Isolate Stool, Other  
Isolate SWAB SOURCE STool PRESERVATIVE  
ISOLATE MEDIA/CONTAINER ISOLATE SOURCE  
SUBMITTER SPECIMEN ID\* ☐ Serum ☐ ACUTE ☐ Tissue  
☐ Other Type/Source (specify) ☐ CONVALESCENT ☐ Urine  
OTHER TYPE/SOURCE NOT LISTED ☐ Sputum ☐ SLIDE/SWAB SOURCE ☐ Wash/Aspirate ☐ WASH/ASPIRATE SOURCE

5 - CLINICAL & EPIDEMIOLOGICAL INFORMATION  
Fever ☐ Diarrhea ☐ Nausea/Discharge ☐ Relevant Vaccination ☐ Travel: ☐ YES ☐ NO  
Headache ☐ Vomiting ☐ Conjunctivitis ☐ Sore Throat  
Rash ☐ Antralgia ☐ Lesion ☐ Cough  
Myalgia ☐ Other (specify) ☐ VACCINATION TYPE ☐ TRAVEL LOCATION ☐ OUTBREAK NAME/ID  
DATE OF ONSET DATE OF EXPOSURE VACCINATION DATE TRAVEL DATES

### 6 - TEST SELECTION\* SET THE TEST DIRECTORY FOR FULL LIST & REQUIREMENTS: <https://www.wslh.wisc.edu/TestDirectory> TEST DIRECTORY OR CODE ->

☐ Influenza PCR NP00051 ☐ Mycobacteria (M) Smear AND Culture HM00250 ☐ Measles Virus PCR - IGH APPROVAL REQUIRED VR01713

☐ A. meningitidis PCR NP00052 ☐ Mycobacteria Isolate Identification HM00253 ☐ Mumps Virus PCR VR01714

☐ Bordetella PCR NP00053 ☐ M. avium complex (MAC) Susceptibility HM00252 ☐ Rubella Virus PCR - IGH APPROVAL REQUIRED VR01725

☐ Shiga Toxin PCR NP00054 ☐ M. tuberculosis complex Genotyping HM00251 ☐ Varicella Zoster Virus PCR & Typing VR01737

☐ Shiga Toxin EIA NP00055 ☐ M. tuberculosis complex Susceptibility HM00254 ☐ Measles IgM AND IgG Ab - Diagnostic SS00120

☐ Legionella Diagnostic PCR NP00056 ☐ M. tuberculosis complex PCR HM00255 ☐ Measles IgG - Immune Status SS00125

☐ Legionella Culture, Diagnostic NP00057 ☐ M. tuberculosis complex PCR HM00256 ☐ Mumps IgM AND IgG Ab - Diagnostic SS00130

☐ Legionella Culture, Surveillance NP00058 ☐ M. avium complex (MAC) PCR HM00257 ☐ Mumps IgG - Immune Status SS00135

☐ Stool Culture, Routine NP00059 ☐ M. avium complex (MAC) PCR HM00258 ☐ Rubella IgG - Immune Status SS00175

☐ Bacterial ID, Enteric NP00060 ☐ M. tuberculosis complex PCR HM00259 ☐ Varicella IgG - Immune Status SS00165

☐ Bacterial ID, Non-Enteric NP00061 ☐ M. tuberculosis complex PCR HM00260 ☐ Brucella Ab SS00202

☐ Stool Culture, Routine NP00062 ☐ M. tuberculosis complex PCR HM00261 ☐ Q Fever IgG Ab SS00203

☐ Bacterial ID, Enteric NP00063 ☐ M. tuberculosis complex PCR HM00262 ☐ Rickettsia IgG Ab (RMSF & Typhus) SS00204

☐ Bacterial ID, Non-Enteric NP00064 ☐ M. tuberculosis complex PCR HM00263 ☐ Measles IgM AND IgG Ab - Diagnostic SS00120

☐ Stool Culture, Routine NP00065 ☐ M. tuberculosis complex PCR HM00264 ☐ Measles IgG - Immune Status SS00125

☐ Bacterial ID, Enteric NP00066 ☐ M. tuberculosis complex PCR HM00265 ☐ Mumps IgM AND IgG Ab - Diagnostic SS00130

☐ Bacterial ID, Non-Enteric NP00067 ☐ M. tuberculosis complex PCR HM00266 ☐ Mumps IgG - Immune Status SS00135

☐ Stool Culture, Routine NP00068 ☐ M. tuberculosis complex PCR HM00267 ☐ Rubella IgG - Immune Status SS00175

☐ Bacterial ID, Enteric NP00069 ☐ M. tuberculosis complex PCR HM00268 ☐ Varicella IgG - Immune Status SS00165

☐ Bacterial ID, Non-Enteric NP00070 ☐ M. tuberculosis complex PCR HM00269 ☐ Brucella Ab SS00202

☐ Stool Culture, Routine NP00071 ☐ M. tuberculosis complex PCR HM00270 ☐ Q Fever IgG Ab SS00203

☐ Bacterial ID, Enteric NP00072 ☐ M. tuberculosis complex PCR HM00271 ☐ Rickettsia IgG Ab (RMSF & Typhus) SS00204

☐ Bacterial ID, Non-Enteric NP00073 ☐ M. tuberculosis complex PCR HM00272 ☐ Measles IgM AND IgG Ab - Diagnostic SS00120

☐ Stool Culture, Routine NP00074 ☐ M. tuberculosis complex PCR HM00273 ☐ Measles IgG - Immune Status SS00125

☐ Bacterial ID, Enteric NP00075 ☐ M. tuberculosis complex PCR HM00274 ☐ Mumps IgM AND IgG Ab - Diagnostic SS00130

☐ Bacterial ID, Non-Enteric NP00076 ☐ M. tuberculosis complex PCR HM00275 ☐ Mumps IgG - Immune Status SS00135

☐ Stool Culture, Routine NP00077 ☐ M. tuberculosis complex PCR HM00276 ☐ Rubella IgG - Immune Status SS00175

☐ Bacterial ID, Enteric NP00078 ☐ M. tuberculosis complex PCR HM00277 ☐ Varicella IgG - Immune Status SS00165

☐ Bacterial ID, Non-Enteric NP00079 ☐ M. tuberculosis complex PCR HM00278 ☐ Brucella Ab SS00202

☐ Stool Culture, Routine NP00080 ☐ M. tuberculosis complex PCR HM00279 ☐ Q Fever IgG Ab SS00203

☐ Bacterial ID, Enteric NP00081 ☐ M. tuberculosis complex PCR HM00280 ☐ Rickettsia IgG Ab (RMSF & Typhus) SS00204

☐ Bacterial ID, Non-Enteric NP00082 ☐ M. tuberculosis complex PCR HM00281 ☐ Measles IgM AND IgG Ab - Diagnostic SS00120

☐ Stool Culture, Routine NP00083 ☐ M. tuberculosis complex PCR HM00282 ☐ Measles IgG - Immune Status SS00125

☐ Bacterial ID, Enteric NP00084 ☐ M. tuberculosis complex PCR HM00283 ☐ Mumps IgM AND IgG Ab - Diagnostic SS00130

☐ Bacterial ID, Non-Enteric NP00085 ☐ M. tuberculosis complex PCR HM00284 ☐ Mumps IgG - Immune Status SS00135

☐ Stool Culture, Routine NP00086 ☐ M. tuberculosis complex PCR HM00285 ☐ Rubella IgG - Immune Status SS00175

☐ Bacterial ID, Enteric NP00087 ☐ M. tuberculosis complex PCR HM00286 ☐ Varicella IgG - Immune Status SS00165

☐ Bacterial ID, Non-Enteric NP00088 ☐ M. tuberculosis complex PCR HM00287 ☐ Brucella Ab SS00202

☐ Stool Culture, Routine NP00089 ☐ M. tuberculosis complex PCR HM00288 ☐ Q Fever IgG Ab SS00203

☐ Bacterial ID, Enteric NP00090 ☐ M. tuberculosis complex PCR HM00289 ☐ Rickettsia IgG Ab (RMSF & Typhus) SS00204

☐ Bacterial ID, Non-Enteric NP00091 ☐ M. tuberculosis complex PCR HM00290 ☐ Measles IgM AND IgG Ab - Diagnostic SS00120

☐ Stool Culture, Routine NP00092 ☐ M. tuberculosis complex PCR HM00291 ☐ Measles IgG - Immune Status SS00125

☐ Bacterial ID, Enteric NP00093 ☐ M. tuberculosis complex PCR HM00292 ☐ Mumps IgM AND IgG Ab - Diagnostic SS00130

☐ Bacterial ID, Non-Enteric NP00094 ☐ M. tuberculosis complex PCR HM00293 ☐ Mumps IgG - Immune Status SS00135

☐ Stool Culture, Routine NP00095 ☐ M. tuberculosis complex PCR HM00294 ☐ Rubella IgG - Immune Status SS00175

☐ Bacterial ID, Enteric NP00096 ☐ M. tuberculosis complex PCR HM00295 ☐ Varicella IgG - Immune Status SS00165

☐ Bacterial ID, Non-Enteric NP00097 ☐ M. tuberculosis complex PCR HM00296 ☐ Brucella Ab SS00202

☐ Stool Culture, Routine NP00098 ☐ M. tuberculosis complex PCR HM00297 ☐ Q Fever IgG Ab SS00203

☐ Bacterial ID, Enteric NP00099 ☐ M. tuberculosis complex PCR HM00298 ☐ Rickettsia IgG Ab (RMSF & Typhus) SS00204

☐ Bacterial ID, Non-Enteric NP00100 ☐ M. tuberculosis complex PCR HM00299 ☐ Measles IgM AND IgG Ab - Diagnostic SS00120

☐ Stool Culture, Routine NP00101 ☐ M. tuberculosis complex PCR HM00300 ☐ Measles IgG - Immune Status SS00125

☐ Bacterial ID, Enteric NP00102 ☐ M. tuberculosis complex PCR HM00301 ☐ Mumps IgM AND IgG Ab - Diagnostic SS00130

☐ Bacterial ID, Non-Enteric NP00103 ☐ M. tuberculosis complex PCR HM00302 ☐ Mumps IgG - Immune Status SS00135

☐ Stool Culture, Routine NP00104 ☐ M. tuberculosis complex PCR HM00303 ☐ Rubella IgG - Immune Status SS00175

☐ Bacterial ID, Enteric NP00105 ☐ M. tuberculosis complex PCR HM00304 ☐ Varicella IgG - Immune Status SS00165

☐ Bacterial ID, Non-Enteric NP00106 ☐ M. tuberculosis complex PCR HM00305 ☐ Brucella Ab SS00202

☐ Stool Culture, Routine NP00107 ☐ M. tuberculosis complex PCR HM00306 ☐ Q Fever IgG Ab SS00203

☐ Bacterial ID, Enteric NP00108 ☐ M. tuberculosis complex PCR HM00307 ☐ Rickettsia IgG Ab (RMSF & Typhus) SS00204

☐ Bacterial ID, Non-Enteric NP00109 ☐ M. tuberculosis complex PCR HM00308 ☐ Measles IgM AND IgG Ab - Diagnostic SS00120

☐ Stool Culture, Routine NP00110 ☐ M. tuberculosis complex PCR HM00309 ☐ Measles IgG - Immune Status SS00125

☐ Bacterial ID, Enteric NP00111 ☐ M. tuberculosis complex PCR HM00310 ☐ Mumps IgM AND IgG Ab - Diagnostic SS00130

☐ Bacterial ID, Non-Enteric NP00112 ☐ M. tuberculosis complex PCR HM00311 ☐ Mumps IgG - Immune Status SS00135

☐ Stool Culture, Routine NP00113 ☐ M. tuberculosis complex PCR HM00312 ☐ Rubella IgG - Immune Status SS00175

☐ Bacterial ID, Enteric NP00114 ☐ M. tuberculosis complex PCR HM00313 ☐ Varicella IgG - Immune Status SS00165

☐ Bacterial ID, Non-Enteric NP00115 ☐ M. tuberculosis complex PCR HM00314 ☐ Brucella Ab SS00202

☐ Stool Culture, Routine NP00116 ☐ M. tuberculosis complex PCR HM00315 ☐ Q Fever IgG Ab SS00203

☐ Bacterial ID, Enteric NP00117 ☐ M. tuberculosis complex PCR HM00316 ☐ Rickettsia IgG Ab (RMSF & Typhus) SS00204

☐ Bacterial ID, Non-Enteric NP00118 ☐ M. tuberculosis complex PCR HM00317 ☐ Measles IgM AND IgG Ab - Diagnostic SS00120

☐ Stool Culture, Routine NP00119 ☐ M. tuberculosis complex PCR HM00318 ☐ Measles IgG - Immune Status SS00125

☐ Bacterial ID, Enteric NP00120 ☐ M. tuberculosis complex PCR HM00319 ☐ Mumps IgM AND IgG Ab - Diagnostic SS00130

☐ Bacterial ID, Non-Enteric NP00121 ☐ M. tuberculosis complex PCR HM00320 ☐ Mumps IgG - Immune Status SS00135

☐ Stool Culture, Routine NP00122 ☐ M. tuberculosis complex PCR HM00321 ☐ Rubella IgG - Immune Status SS00175

☐ Bacterial ID, Enteric NP00123 ☐ M. tuberculosis complex PCR HM00322 ☐ Varicella IgG - Immune Status SS00165

☐ Bacterial ID, Non-Enteric NP00124 ☐ M. tuberculosis complex PCR HM00323 ☐ Brucella Ab SS00202

☐ Stool Culture, Routine NP00125 ☐ M. tuberculosis complex PCR HM00324 ☐ Q Fever IgG Ab SS00203

☐ Bacterial ID, Enteric NP00126 ☐ M. tuberculosis complex PCR HM00325 ☐ Rickettsia IgG Ab (RMSF & Typhus) SS00204

☐ Bacterial ID, Non-Enteric NP00127 ☐ M. tuberculosis complex PCR HM00326 ☐ Measles IgM AND IgG Ab - Diagnostic SS00120

☐ Stool Culture, Routine NP00128 ☐ M. tuberculosis complex PCR HM00327 ☐ Measles IgG - Immune Status SS00125

☐ Bacterial ID, Enteric NP00129 ☐ M. tuberculosis complex PCR HM00328 ☐ Mumps IgM AND IgG Ab - Diagnostic SS00130

☐ Bacterial ID, Non-Enteric NP00130 ☐ M. tuberculosis complex PCR HM00329 ☐ Mumps IgG - Immune Status SS00135

☐ Stool Culture, Routine NP00131 ☐ M. tuberculosis complex PCR HM00330 ☐ Rubella IgG - Immune Status SS00175

☐ Bacterial ID, Enteric NP00132 ☐ M. tuberculosis complex PCR HM00331 ☐ Varicella IgG - Immune Status SS00165

☐ Bacterial ID, Non-Enteric NP00133 ☐ M. tuberculosis complex PCR HM00332 ☐ Brucella Ab SS00202

☐ Stool Culture, Routine NP00134 ☐ M. tuberculosis complex PCR HM00333 ☐ Q Fever IgG Ab SS00203

☐ Bacterial ID, Enteric NP00135 ☐ M. tuberculosis complex PCR HM00334 ☐ Rickettsia IgG Ab (RMSF & Typhus) SS00204

☐ Bacterial ID, Non-Enteric NP00136 ☐ M. tuberculosis complex PCR HM00335 ☐ Measles IgM AND IgG Ab - Diagnostic SS00120

☐ Stool Culture, Routine NP00137 ☐ M. tuberculosis complex PCR HM00336 ☐ Measles IgG - Immune Status SS00125

☐ Bacterial ID, Enteric NP00138 ☐ M. tuberculosis complex PCR HM00337 ☐ Mumps IgM AND IgG Ab - Diagnostic SS00130

☐ Bacterial ID, Non-Enteric NP00139 ☐ M. tuberculosis complex PCR HM00338 ☐ Mumps IgG - Immune Status SS00135

☐ Stool Culture, Routine NP00140 ☐ M. tuberculosis complex PCR HM00339 ☐ Rubella IgG - Immune Status SS00175

☐ Bacterial ID, Enteric NP00141 ☐ M. tuberculosis complex PCR HM00340 ☐ Varicella IgG - Immune Status SS00165

☐ Bacterial ID, Non-Enteric NP00142 ☐ M. tuberculosis complex PCR HM00341 ☐ Brucella Ab SS00202

☐ Stool Culture, Routine NP00143 ☐ M. tuberculosis complex PCR HM00342 ☐ Q Fever IgG Ab SS00203

☐ Bacterial ID, Enteric NP00144 ☐ M. tuberculosis complex PCR HM00343 ☐ Rickettsia IgG Ab (RMSF & Typhus) SS00204

☐ Bacterial ID, Non-Enteric NP00145 ☐ M. tuberculosis complex PCR HM00344 ☐ Measles IgM AND IgG Ab - Diagnostic SS00120

☐ Stool Culture, Routine NP00146 ☐ M. tuberculosis complex PCR HM00345 ☐ Measles IgG - Immune Status SS00125

☐ Bacterial ID, Enteric NP00147 ☐ M. tuberculosis complex PCR HM00346 ☐ Mumps IgM AND IgG Ab - Diagnostic SS00130

☐ Bacterial ID, Non-Enteric NP00148 ☐ M. tuberculosis complex PCR HM00347 ☐ Mumps IgG - Immune Status SS00135

☐ Stool Culture, Routine NP00149 ☐ M. tuberculosis complex PCR HM00348 ☐ Rubella IgG - Immune Status SS00175

☐ Bacterial ID, Enteric NP00150 ☐ M. tuberculosis complex PCR HM00349 ☐ Varicella IgG - Immune Status SS00165

☐ Bacterial ID, Non-Enteric NP00151 ☐ M. tuberculosis complex PCR HM00350 ☐ Brucella Ab SS00202

☐ Stool Culture, Routine NP00152 ☐ M. tuberculosis complex PCR HM00351 ☐ Q Fever IgG Ab SS00203

☐ Bacterial ID, Enteric NP00153 ☐ M. tuberculosis complex PCR HM00352 ☐ Rickettsia IgG Ab (RMSF & Typhus) SS00204

☐ Bacterial ID, Non-Enteric NP00154 ☐ M. tuberculosis complex PCR HM00353 ☐ Measles IgM AND IgG Ab - Diagnostic SS00120

☐ Stool Culture, Routine NP00155 ☐ M. tuberculosis complex PCR HM00354 ☐ Measles IgG - Immune Status SS00125

☐ Bacterial ID, Enteric NP00156 ☐ M. tuberculosis complex PCR HM00355 ☐ Mumps IgM AND IgG Ab - Diagnostic SS00130

☐ Bacterial ID, Non-Enteric NP00157 ☐ M. tuberculosis complex PCR HM00356 ☐ Mumps IgG - Immune Status SS00135

☐ Stool Culture, Routine NP00158 ☐ M. tuberculosis complex PCR HM00357 ☐ Rubella IgG - Immune Status SS00175

☐ Bacterial ID, Enteric NP00159 ☐ M. tuberculosis complex PCR HM00358 ☐ Varicella IgG - Immune Status SS00165

☐ Bacterial ID, Non-Enteric NP00160 ☐ M. tuberculosis complex PCR HM00359 ☐ Brucella Ab SS00202

☐ Stool Culture, Routine NP00161 ☐ M. tuberculosis complex PCR HM00360 ☐ Q Fever IgG Ab SS00203

☐ Bacterial ID, Enteric NP00162 ☐ M. tuberculosis complex PCR HM00361 ☐ Rickettsia IgG Ab (RMSF & Typhus) SS00204

☐ Bacterial ID, Non-Enteric NP00163 ☐ M. tuberculosis complex PCR HM00362 ☐ Measles IgM AND IgG Ab - Diagnostic SS00120

☐ Stool Culture, Routine NP00164 ☐ M. tuberculosis complex PCR HM00363 ☐ Measles IgG - Immune Status SS00125

☐ Bacterial ID, Enteric NP00165 ☐ M. tuberculosis complex PCR HM00364 ☐ Mumps IgM AND IgG Ab - Diagnostic SS00130

☐ Bacterial ID, Non-Enteric NP00166 ☐ M. tuberculosis complex PCR HM00365 ☐ Mumps IgG - Immune Status SS00135

☐ Stool Culture, Routine NP00167 ☐ M. tuberculosis complex PCR HM00366 ☐ Rubella IgG - Immune Status SS00175

☐ Bacterial ID, Enteric NP00168 ☐ M. tuberculosis complex PCR HM00367 ☐ Varicella IgG - Immune Status SS00165

☐ Bacterial ID, Non-Enteric NP00169 ☐ M. tuberculosis complex PCR HM00368 ☐ Brucella Ab SS00202

☐ Stool Culture, Routine NP00170 ☐ M. tuberculosis complex PCR HM00369 ☐ Q Fever IgG Ab SS00203

☐ Bacterial ID, Enteric NP00171 ☐ M. tuberculosis complex PCR HM00370 ☐ Rickettsia IgG Ab (RMSF & Typhus) SS00204

☐ Bacterial ID, Non-Enteric NP00172 ☐ M. tuberculosis complex PCR HM00371 ☐ Measles IgM AND IgG Ab - Diagnostic SS00120

☐ Stool Culture, Routine NP00173 ☐ M. tuberculosis complex PCR HM00372 ☐ Measles IgG - Immune Status SS00125

☐ Bacterial ID, Enteric NP00174 ☐ M. tuberculosis complex PCR HM00373 ☐ Mumps IgM AND IgG Ab - Diagnostic SS00130

☐ Bacterial ID, Non-Enteric NP00175 ☐ M. tuberculosis complex PCR HM00374 ☐ Mumps IgG - Immune Status SS00135

☐ Stool Culture, Routine NP00176 ☐ M. tuberculosis complex PCR HM00375 ☐ Rubella IgG - Immune Status SS00175

☐ Bacterial ID, Enteric NP00177 ☐ M. tuberculosis complex PCR HM00376 ☐ Varicella IgG - Immune Status SS00165

☐ Bacterial ID, Non-Enteric NP00178 ☐ M. tuberculosis complex PCR HM00377 ☐ Brucella Ab SS00202

☐ Stool Culture, Routine NP00179 ☐ M. tuberculosis complex PCR HM00378 ☐ Q Fever IgG Ab SS00203

☐ Bacterial ID, Enteric NP00180 ☐ M. tuberculosis complex PCR HM00379 ☐ Rickettsia IgG Ab (RMSF & Typhus) SS00204

☐ Bacterial ID, Non-Enteric NP00181 ☐ M. tuberculosis complex PCR HM00380 ☐ Measles IgM AND IgG Ab - Diagnostic SS00120

☐ Stool Culture, Routine NP00182 ☐ M. tuberculosis complex PCR HM00381 ☐ Measles IgG - Immune Status SS00125

☐ Bacterial ID, Enteric NP00183 ☐ M. tuberculosis complex PCR HM00382 ☐ Mumps IgM AND IgG Ab - Diagnostic SS00130

☐ Bacterial ID, Non-Enteric NP00184 ☐ M. tuberculosis complex PCR HM00383 ☐ Mumps IgG - Immune Status SS00135

☐ Stool Culture, Routine NP00185 ☐ M. tuberculosis complex PCR HM00384 ☐ Rubella IgG - Immune Status SS00175

☐ Bacterial ID, Enteric NP00186 ☐ M. tuberculosis complex PCR HM00385 ☐ Varicella IgG - Immune Status SS00165

☐ Bacterial ID, Non-Enteric NP00187 ☐ M. tuberculosis complex PCR HM00386 ☐ Brucella Ab SS00202

☐ Stool Culture, Routine NP00188 ☐ M. tuberculosis complex PCR HM00387 ☐ Q Fever IgG Ab SS00203

☐ Bacterial ID, Enteric NP00189 ☐ M. tuberculosis complex PCR HM00388 ☐ Rickettsia IgG Ab (RMSF & Typhus) SS00204

☐ Bacterial ID, Non-Enteric NP00190 ☐ M. tuberculosis complex PCR HM00389 ☐ Measles IgM AND IgG Ab - Diagnostic SS00120

☐ Stool Culture, Routine NP00191 ☐ M. tuberculosis complex PCR HM00390 ☐ Measles IgG - Immune Status SS00125

☐ Bacterial ID, Enteric NP00192 ☐ M. tuberculosis complex PCR HM00391 ☐ Mumps IgM AND IgG Ab - Diagnostic SS00130

☐ Bacterial ID, Non-Enteric NP00193 ☐ M. tuberculosis complex PCR HM00392 ☐ Mumps IgG - Immune Status SS00135

☐ Stool Culture, Routine NP00194 ☐ M. tuberculosis complex PCR HM00393 ☐ Rubella IgG - Immune Status SS00175

☐ Bacterial ID, Enteric NP00195 ☐ M. tuberculosis complex PCR HM00394 ☐ Varicella IgG - Immune Status SS00165

☐ Bacterial ID, Non-Enteric NP00196 ☐ M. tuberculosis complex PCR HM00395 ☐ Brucella Ab SS00202

☐ Stool Culture, Routine NP00197 ☐ M. tuberculosis complex PCR HM00396 ☐ Q Fever IgG Ab SS00203

☐ Bacterial ID, Enteric NP00198 ☐ M. tuberculosis complex PCR HM00397 ☐ Rickettsia IgG Ab (RMSF & Typhus) SS00204

☐ Bacterial ID, Non-Enteric NP00199 ☐ M. tuberculosis complex PCR HM00398 ☐ Measles IgM AND IgG Ab - Diagnostic SS00120

☐ Stool Culture, Routine NP00200 ☐ M. tuberculosis complex PCR HM00399 ☐ Measles IgG - Immune Status SS00125

☐ Bacterial ID, Enteric NP00201 ☐ M. tuberculosis complex PCR HM00400 ☐ Mumps IgM AND IgG Ab - Diagnostic SS00130

☐ Bacterial ID, Non-Enteric NP00202 ☐ M. tuberculosis complex PCR HM00401 ☐ Mumps IgG - Immune Status SS00135

☐ Stool Culture, Routine NP00203 ☐ M. tuberculosis complex PCR HM00402 ☐ Rubella IgG - Immune Status SS00175

☐ Bacterial ID, Enteric NP00204 ☐ M. tuberculosis complex PCR HM00403 ☐ Varicella IgG - Immune Status SS00165

☐ Bacterial ID, Non-Enteric NP00205 ☐ M. tuberculosis complex PCR HM00404 ☐ Brucella Ab SS00202

☐ Stool Culture, Routine NP00206 ☐ M. tuberculosis complex PCR HM00405 ☐ Q Fever IgG Ab SS00203

☐ Bacterial ID, Enteric NP00207 ☐ M. tuberculosis complex PCR HM00406 ☐ Rickettsia IgG Ab (RMSF & Typhus) SS00204

☐ Bacterial ID, Non-Enteric NP00208 ☐ M. tuberculosis complex PCR HM00407 ☐ Measles IgM AND IgG Ab - Diagnostic SS00120

☐ Stool Culture, Routine NP00209 ☐ M. tuberculosis complex PCR HM00408 ☐ Measles IgG - Immune Status SS00125

☐ Bacterial ID, Enteric NP00210 ☐ M. tuberculosis complex PCR HM00409 ☐ Mumps IgM AND IgG Ab - Diagnostic SS00130

☐ Bacterial ID, Non-Enteric NP00211 ☐ M. tuberculosis complex PCR HM00410 ☐ Mumps IgG - Immune Status SS00135

☐ Stool Culture, Routine NP00212 ☐ M. tuberculosis complex PCR HM00411 ☐ Rubella IgG - Immune Status SS00175

☐ Bacterial ID, Enteric NP0

## Wisconsin Enteric Pathogen Surveillance Submissions

**Table 4. Enteric Pathogen Specimen Submissions\***

Pathogen	Specimen Type	Testing Performed at WSLH
<b><i>Campylobacter</i> species</b>	Isolates or stool	Identification, antimicrobial susceptibility testing and molecular subtyping (WGS) will be performed as necessary
<b>Enterohemorrhagic/Shiga Toxin-Producing <i>E. coli</i> (EHEC/STEC)</b>	Isolates, stool or enrichment broth	Identification, serotyping and molecular subtyping (WGS)
<b><i>Salmonella</i> species</b>	Isolates or stool	Identification, serotyping, antimicrobial susceptibility testing and molecular subtyping (WGS)
<b><i>Shigella</i> species and Enteroinvasive <i>E. coli</i> (EIEC)</b>	Isolates or stool	<i>Shigella</i> identification, serogrouping, antimicrobial susceptibility testing, and molecular subtyping
<b><i>Vibrio</i> Species</b>	Isolates or stool	Identification and referral to CDC
<b><i>Yersinia</i> species</b>	Isolates or stool	Identification
<b><i>Cryptosporidium</i> species</b>	Stool	Identification** and genotyping (and/or referral to CDC)
<b><i>Cyclospora cayetanensis</i></b>	Stool	Molecular subtyping and/or referral to CDC
<b>Rotavirus</b>	Stool	<u>One positive per week</u> for molecular subtyping/genotyping (performed at CDC)

\*Consult with the Wisconsin Division of Public Health Foodborne Disease Epidemiologists to inquire about testing of any other organisms that are suspected of being in a cluster or outbreak of public health significance.

\*\*Stool specimens positive for *Cryptosporidium* by PCR-based methods will not be confirmed.

# Antimicrobial Resistance Monitoring



The WSLH is the Midwest Regional Laboratory for the CDC-coordinated Antimicrobial Resistance Laboratory Network (AR Lab Network). The overarching goal of AR Lab Network testing is rapid identification and containment of resistant pathogens. Data compiled from the AR Lab Network is shared with CDC and Wisconsin Clinical Laboratory Network partners. A summary can be found on pages 35-38.

## Specimen Submission instructions:

WSLH requests that clinical laboratories submit the isolated pathogens listed in Table 5.

- One isolate per patient with the same organism and mechanism will be tested per 12 month period.
- If more than one isolate per patient is received at the same time, the more invasive isolate will be tested.
- If your facility performs carbapenemase testing (e.g., mCIM, Carba-NP®, NG CARBA 5®, Ce-  
pheid CARBA-R®) on Enterobacterales and *Pseudomonas aeruginosa* isolates, only submit isolates that test positive for a carbapenemase.

Please fill out a **WSLH ARLN Requisition form** (included with your mailing packet) with each isolate submitted and include:

- AST results and any relevant phenotypic or molecular test results.
- Specimen requisition forms and supplies for shipping surveillance specimens are provided at **NO COST** and can be requested at 800-862-1088.
- Transport to the WSLH is available at **NO COST** when you send specimens using **Purple Mountain Solutions** (PH: 800-990-9668). Detailed shipping instructions can be found on pages 27-28.

WISCONSIN STATE LABORATORY OF HYGIENE		2601 Agriculture Dr Madison, WI 53718 800-862-1013		Antimicrobial Resistant Isolate Characterization Version 2 (06/2024)	
<b>Patient Information</b>				<b>Submitter Information</b>	
Name (Last, First):					
Address:					
City: State: Zip:					
Date of Birth:		Gender: M F		(Telephone Number)	
Your Patient ID Number:				Laboratory Contact Name	
Your Specimen ID Number:				Submitter CLIA or NPI Number (facility):	
OC Result Report to:				WSLH Use Only Study: ARLN CRE/CRPA/CANDIDA	
<b>Originating Facility (where specimen was collected):</b>					
Name: _____					
City and Zip Code _____					
<b>Results and Tests Performed in Your Laboratory:</b>					
<input type="checkbox"/> OXA	<input type="checkbox"/> Negative	<input type="checkbox"/> Positive	<input type="checkbox"/> Carba-NP	<input type="checkbox"/> Negative	<input type="checkbox"/> Positive
<input type="checkbox"/> IMP	<input type="checkbox"/> Negative	<input type="checkbox"/> Positive	<input type="checkbox"/> mCIM	<input type="checkbox"/> Negative	<input type="checkbox"/> Positive
<input type="checkbox"/> KPC	<input type="checkbox"/> Negative	<input type="checkbox"/> Positive	<input type="checkbox"/> Other (specify): _____		
<input type="checkbox"/> VIM	<input type="checkbox"/> Negative	<input type="checkbox"/> Positive	<input type="checkbox"/> Other (specify): _____		
<input type="checkbox"/> NDM	<input type="checkbox"/> Negative	<input type="checkbox"/> Positive	<input type="checkbox"/> Negative <input type="checkbox"/> Positive		
<b>Organism (Your Test Results):</b>					
<input type="checkbox"/> Acinetobacter baumannii			<input type="checkbox"/> Klebsiella pneumoniae		
<input type="checkbox"/> Candida (species: _____)			<input type="checkbox"/> Klebsiella oxytoca		
<input type="checkbox"/> Enterobacter (species: _____)			<input type="checkbox"/> Pseudomonas aeruginosa		
<input type="checkbox"/> Escherichia coli			<input type="checkbox"/> Other (specify): _____		
Date Collected:		Specimen Type:			
Time Collected:		<input type="checkbox"/> Isolate, source: _____			
<b>Please include AST Results performed at your laboratory with the specimen</b>					
WSLH Test Request:					
<input type="checkbox"/> MP00580 Bacterial Characterization			<input type="checkbox"/> MP00900 Fungal Characterization		
<input type="checkbox"/> MP00696 Aztreonam-Avibactam (ExAST)					
(Prior approval required)					
WISCONSIN STATE LABORATORY OF HYGIENE USE ONLY					

## Antimicrobial Resistance Specimen Submissions

Table 5. Antimicrobial Resistance Specimen Submissions

Pathogen	Resistance Traits	Testing Performed at WSLH
<b>Pan-resistant organisms</b>	Resistant to all drugs tested in your laboratory	Identification, antimicrobial susceptibility testing, AR-targeted PCR and referral to CDC as necessary
<b><i>Candida auris</i></b>	N/A	<ul style="list-style-type: none"> <li>• Identification</li> <li>• Molecular subtyping</li> <li>• Antimicrobial susceptibility testing (performed on invasive and urine isolates. Other isolates upon request)</li> </ul>
<b><i>Enterobacterales</i></b>	Resistant to any carbapenems (CRE)**	Identification, antimicrobial susceptibility testing, carbapenemase screen, AR-targeted PCR and molecular subtyping
<b><i>Staphylococcus aureus</i></b>	Non-susceptible to vancomycin (VRSA)	Identification, antimicrobial susceptibility testing and referral to CDC as necessary
<b><i>Pseudomonas aeruginosa</i> ***</b>	<ul style="list-style-type: none"> <li>• Resistant to carbapenems other than ertapenem</li> </ul> <p style="text-align: center;"><b>AND</b></p> <ul style="list-style-type: none"> <li>• Non-susceptible to cefepime and/or ceftazidime</li> </ul> <p style="text-align: center;"><b>OR</b></p> <ul style="list-style-type: none"> <li>• Resistant to ceftolozane/tazobactam</li> </ul>	Identification, antimicrobial susceptibility testing, carbapenemase screen, AR-targeted PCR and molecular subtyping
<b><i>Acinetobacter baumannii</i></b>	Resistant to any carbapenems (CRAB)	Identification, antimicrobial susceptibility testing, AR-targeted PCR and molecular subtyping
<b><i>Aspergillus fumigatus</i></b> isolates from invasive infections	N/A	Isolates will be forwarded to the Maryland Department of Health for surveillance of azole resistance

\*\* Exception: *Proteus* spp., *Providencia* spp., and *Morganella* spp., that are resistant to imipenem only but are not resistant to meropenem or doripenem. These isolates may have elevated imipenem MICs by mechanisms other than production of carbapenemases. These isolates will also be susceptible to other beta-lactams.

\*\*\*Exception: Do not submit resistant *Pseudomonas aeruginosa* isolates from cystic fibrosis patients or mucoid isolates due to difficulties in testing.

# Wisconsin Invasive Pathogen Surveillance

The Wisconsin Invasive Pathogen Surveillance program is a partnership between WSLH and other public health and clinical partners, and monitors invasive infections in the state. The WSLH requests the submission of isolates and specimens of invasive pathogens listed in Table 6 **As Detected**.



**Specimen submission instructions:**

Isolates should be from **sterile body sites** (CSF, Blood, etc...).

- In the absence of an isolate, please submit CSF specimens which have been determined to contain these pathogens by a culture-independent diagnostic test (CIDT).
- If the specimen is an isolate, please indicate the source (CSF, Blood, etc...).
- For **invasive bacterial** submissions, Please fill out a WSLH **General requisition** with each specimen submitted and indicate the pathogen isolated/detected.
  - Place a check mark next to the pathogen detected or isolated.

[illegible]

<input type="checkbox"/> <i>H. influenzae</i> PCR	MP00651
<input type="checkbox"/> <i>N. meningitidis</i> PCR	MP00561

- For *Listeria monocytogenes* isolates, please select **Bacterial ID, Non-Enteric,**

**ID & CHARACTERIZATION**

---

**ORGANISM SUSPECTED**

CIDT: ☐ YES ☐ NO    ID METHOD:


☐ Stool Culture, Routine

☐ Bacterial ID, Enteric

☐ Bacterial ID, Non-Enteric

- For ***Streptococcus pneumoniae*** specimens, please fill out the *Streptococcus pneumoniae* Requisition form (included with your mailing packet). Attach a copy of the susceptibility results if available.
  - Check the appropriate specimen type:
  - If submitting an isolate, indicate the source (must be a sterile body site)

**Specimen Type:**  
☐ CSF  
☐ Isolate  
**Source:** ☐ Blood   ☐ CSF   ☐ Other \_\_\_\_\_

 <b>WSHL</b> <small>WILSON STATE</small> <b>LABORATORY OF HYGIENE</b>		<i>Streptococcus pneumoniae</i> Registration version 2 (06/2024)	
<b>Patient Information</b>		<b>Submitter Information</b>	
Name (Last, First):		(Your Institution's Agency Number- If Known and CLIA Number)	
Address:		(Your Institution's Name)	
City: State: Zip:		(Your Institution's Address)	
Date of Birth:		(City, State, Zip Code)	
Gender: M: F:		(Telephone Number)	
Your Patient ID Number:		Laboratory Contact Name:	
Your Specimen ID Number:		REFL Use Only Study: <b>AREN</b> Specimen	
<b>Vaccination History:</b> Was patient vaccinated? <input type="checkbox"/> Yes <input type="checkbox"/> No <input type="checkbox"/> Unknown If Yes, Date of Last Vaccination:                      /                      /			
Children: <input type="checkbox"/> PCV15 <input type="checkbox"/> PCV20 Adults: <input type="checkbox"/> PCV15 <input type="checkbox"/> DMS2323		<div style="border: 2px solid red; padding: 10px; margin: 10px;"> <b>Specimen Type:</b>  <input type="checkbox"/> CSF  <input type="checkbox"/> Isolate  <b>Source:</b> <input type="checkbox"/> Blood <input type="checkbox"/> CSF <input type="checkbox"/> Other _____         </div>	
<b>Date Collected:</b>			
<b>Time Collected:</b>			
Criteria for testing (must meet one of the following): <input type="checkbox"/> CSF (isolation only) <input type="checkbox"/> CSF (PCR)			
Include isolates that are: <input type="checkbox"/> Resistant to clinically relevant antibiotic* <input type="checkbox"/> Suspected vaccine failure <input type="checkbox"/> Suspected treatment failure* <input type="checkbox"/> Suspected outbreak			
<b>*Attach copy of susceptibility results if available</b> <b>WSHL Test Request:</b> <input type="checkbox"/> MP00461 S. pneumoniae PCR (CSF only) <input type="checkbox"/> MP00689 S. pneumoniae serotyping			

## Wisconsin Invasive Pathogen Surveillance Submissions

- Select one or more criteria for testing (REQUIRED)

Criteria for testing (must meet one of the following):

- ☐ CSF (identification only)
- ☐ CSF isolate

Invasive isolates that are:

- ☐ Resistant to clinically relevant antibiotic\*
- ☐ Suspected vaccine failure
- ☐ Suspected treatment failure\*
- ☐ Suspected outbreak

- At the bottom of the page, indicate the testing being requested.
- Specimen requisition forms and supplies for shipping surveillance specimens are provided at **NO COST** and can be requested at 800-862-1088 .
- Transport to the WSLH is available at **NO COST** when you send specimens using **Purple Mountain Solutions**. Detailed shipping instructions can be found on pages 27-28.

**Table 6. Invasive Pathogen Specimen Submission Requests**

Pathogen	Specimen Type	Testing Performed at WSLH
<i>Haemophilus influenzae</i>	Isolates or CSF	Identification and serotyping
<i>Listeria monocytogenes</i>	Isolates	Identification and molecular subtyping (WGS)
<i>Neisseria meningitidis</i>	Isolates or CSF	Identification, antimicrobial susceptibility testing and serogrouping
<i>Streptococcus pneumoniae</i>	Isolates or CSF	Identification, antimicrobial susceptibility testing and serotyping performed on: <ul style="list-style-type: none"> <li>CSF (Identification PCR only)</li> <li>CSF isolates</li> <li>Non-CSF invasive isolates that are:               <ul style="list-style-type: none"> <li>Non-susceptible to clinically relevant antibiotics</li> <li>Suspected vaccine failure</li> <li>Suspected treatment failure</li> <li>Outbreak related isolates</li> </ul> </li> </ul>



# Vector-borne Pathogen Surveillance

The WSLH works with WDPH to monitor cases of vector-borne diseases in our state. WSLH will confirm species identification of key blood borne pathogens and submit specimens to CDC to monitor for drug resistance and emergence of novel pathogens. Please submit the organisms listed in Table 7 **As Detected**.



## Specimen Submission instructions:

- Please fill out a WSLH General requisition form with each specimen submitted.
  - Indicate the pathogen detected in the “Other Tests” field in the bottom right.
- Specimen requisition forms and supplies for shipping surveillance specimens are provided at **NO COST** and can be requested at 800-862-1088.
- Transport to the WSLH is available at **NO COST** when you send specimens using **Purple Mountain Solutions**. Detailed shipping instructions can be found on pages 27-28.

**Table 7. Vector-borne Pathogen Specimen Submission Requests**

Pathogen	Specimen Type	Testing Performed at WSLH
<b>Malaria</b>	Positive thick and thin blood smears or residual EDTA blood	Species confirmation via microscopy and PCR.
<b>Babesia</b>	Positive thick and thin blood smears or residual EDTA blood	Confirmation of <i>B. microti</i> by PCR. Unknown species forwarded to CDC for confirmation
<b>Ehrlichia (species unknown)</b>	Residual blood and/or nucleic acid	Forwarded to CDC for species identification (if speciation not available at your lab)



## Other Pathogens of Public Health Significance

WSLH also requests the following specimens for state-wide surveillance purposes. These specimens help to monitor circulating strains, and identify potential outbreaks or emerging pathogens.

### Specimen Submission instructions:



- Please fill out a WSLH General requisition form with each specimen submitted.
  - Indicate the pathogen detected in the “Other Tests” field in the bottom right.
- Specimen requisition forms and supplies for shipping surveillance specimens are provided at **NO COST** and can be requested at 800-862-1088.
- Transport to the WSLH is available at **NO COST** when you send specimens using **Purple Mountain Solutions**. Detailed shipping instructions can be found on pages 27-28.

**Table 8. Other Pathogens of Public Health Significance Submission Requests**

Pathogen	Specimen Type	Testing Performed at WSLH
Hepatitis C Virus	Serum	All new positives, molecular subtyping (WGS)
<i>Cronobacter</i> spp.	Isolates from infants	Identification and molecular subtyping (WGS)
Enterovirus	CSF	Molecular typing
<i>Blastomyces</i>	Isolates	Species identification
Other organisms suspected of being part of a cluster or outbreak of public health significance	Isolates or specimens	Consult with Wisconsin Division of Public Health Epidemiologists to inquire about testing (608-267-9003)

# Shipment of Specimens

## Order Supplies:

Specimen collection and shipping supplies including:

- Requisition sheets
- Specimen collection kits
- Mailers and ice packs



Supplies are available at **NO COST** for surveillance submissions. Please contact the WSLH Clinical Orders department at **800-862-1088** or to order supplies.

## Shipping Resources

Guidance and training materials for the shipping of category B biological substances can be found on the WSLH website: [www.slh.wisc.edu/clinical/diseases/packaging-and-shipping/](http://www.slh.wisc.edu/clinical/diseases/packaging-and-shipping/)

**Mycobacterium tuberculosis complex (MTBC) isolates MUST be submitted using Category A packaging and shipping requirements.** Guidance for shipping Category A can be found on the Department of Transportation website: [www.phmsa.dot.gov/](http://www.phmsa.dot.gov/)

## Category B Specimen Packaging:

If classified as Category B, specimens should be triple packaged and shipped as:

***“Biological substance, Category B / UN 3373”***

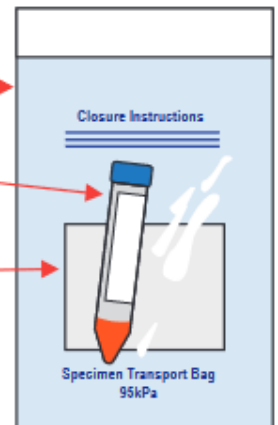
- Label each specimen with patient name (First and Last) and date of birth, and the specimen source.
- Ensure the specimen cap is on tight and the specimen is not leaking.
- Securely tape the cap of the specimen container, and wrap specimen with absorbent material.
- Place the specimen vial into a biohazard bag.

### Secondary Packaging

Biohazard or 95kPa specimen bag

Primary Receptacle

Absorbent material



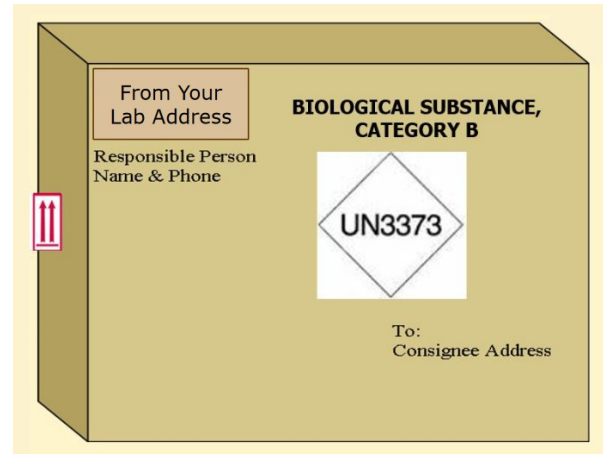
- Place the completed requisition form into the **outer** sleeve of the bag.
- Place the bagged specimen and form in the Styrofoam mailer with a **frozen** cold pack and cushioning.
- Replace lid on the Styrofoam box. Close and securely tape the cardboard box shut.

## Shipment of Specimens (Cont'd)

- Attach the WSLH address label to the package:

**Wisconsin State Lab of Hygiene**  
**Specimen Receiving, Communicable Disease Division**  
**2601 Agriculture Drive**  
**Madison, WI 53718**

- Attach the “*Biological substance, Category B / UN 3373*” label to the package.
- Attach your return address label
- **Include the name and telephone number** of the person who knows the content of the package (**REQUIRED**)



### Shipping Arrangements:

Transport of surveillance specimens or other specimens requested by WSLH is available at **NO COST** when you send specimens using **Purple Mountain Solutions** (<https://purplemountainsolutions.com/>)

- The WSLH has a contract with Purple Mountain Solutions for shipment of specimens to the WSLH, with charges billed to the WSLH.
- This account is for surveillance specimens or others requested by WSLH. Funding is not available for transport of other samples.
- You are not required to ship via Purple Mountain Solutions, only if you wish to have the transport charges billed to the WSLH.
- Specimens will be picked up during regular working hours, but you must confirm the time with Purple Mountain Solutions.
- Specimens will be delivered to the WSLH the following day. **Ensure to include enough coolant to preserve items during shipment.**
- All package preparation should be completed before the courier arrives.
- Contact Purple Mountain Solutions directly to arrange for a pick-up. Orders must be placed by noon for same day pickup.

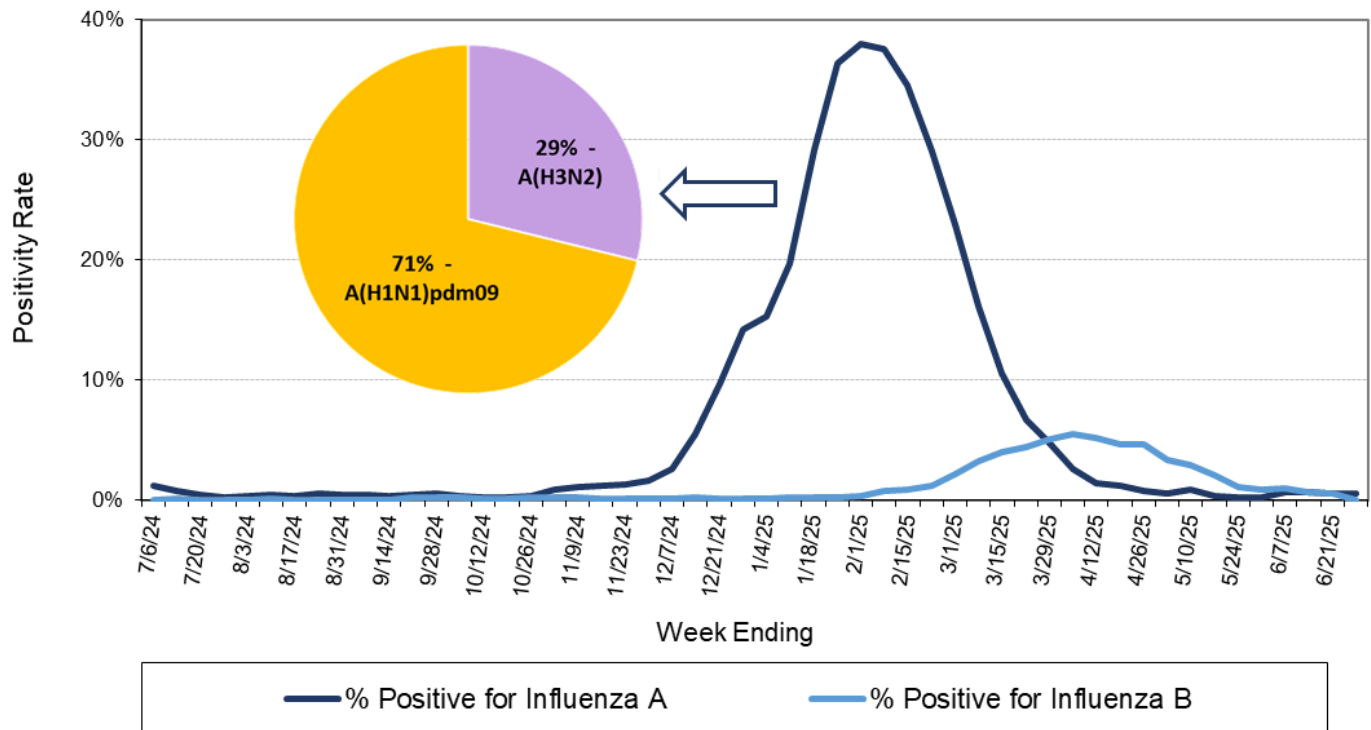
**Phone #: 800-990-9668**



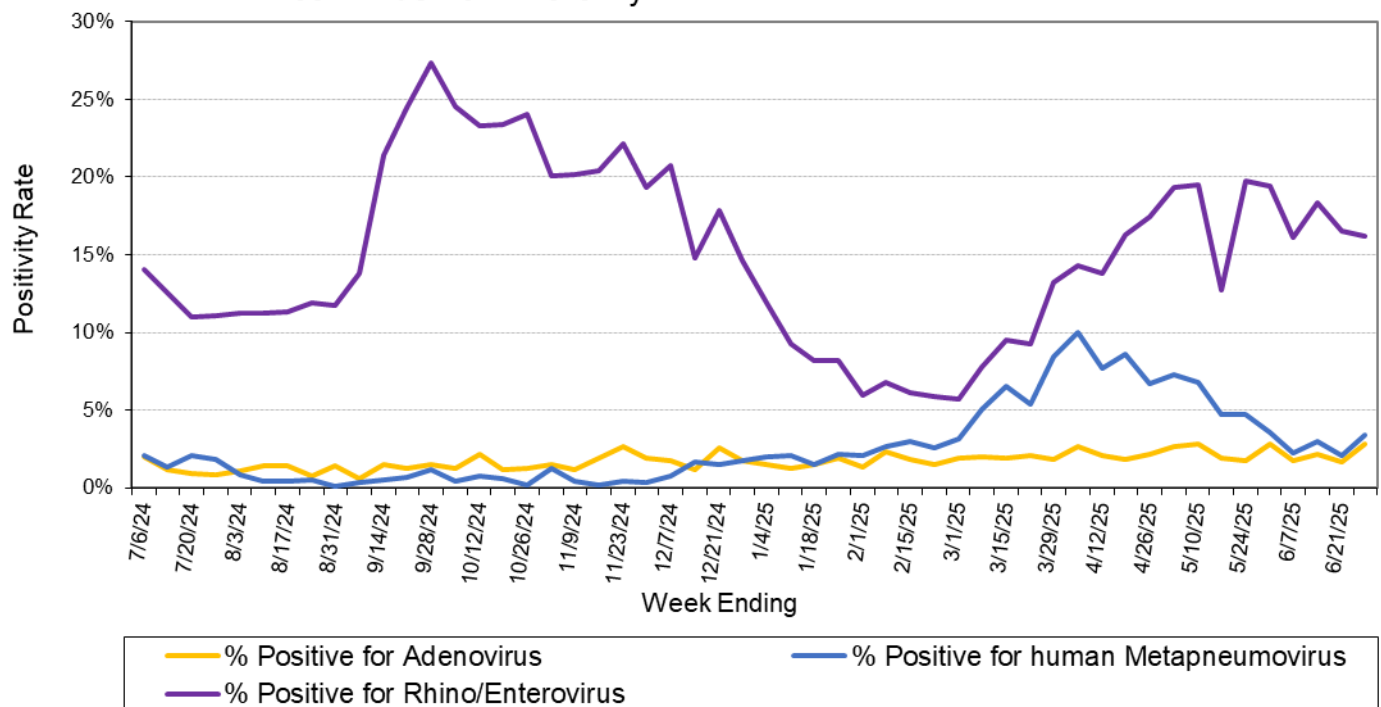
- Same day delivery may be available for specific high priority specimens. However, **pre-approval from WSLH is necessary for same day delivery.**

# Surveillance Graphs for 2024-25 Season

Positivity Rate and Subtyping for **Influenza** by PCR at Wisconsin Laboratories

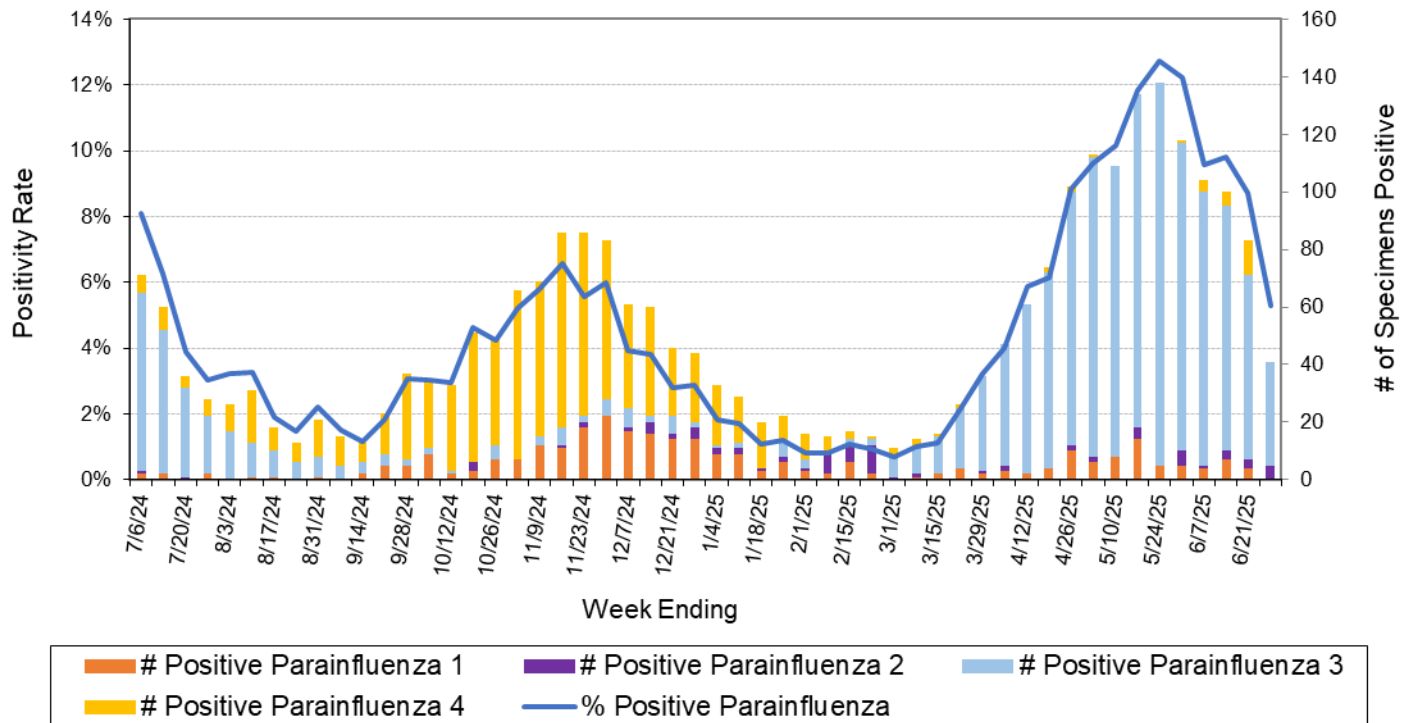


Positivity Rate of **Adenovirus, hMPV and Rhino/Enterovirus** by PCR at Wisconsin Laboratories

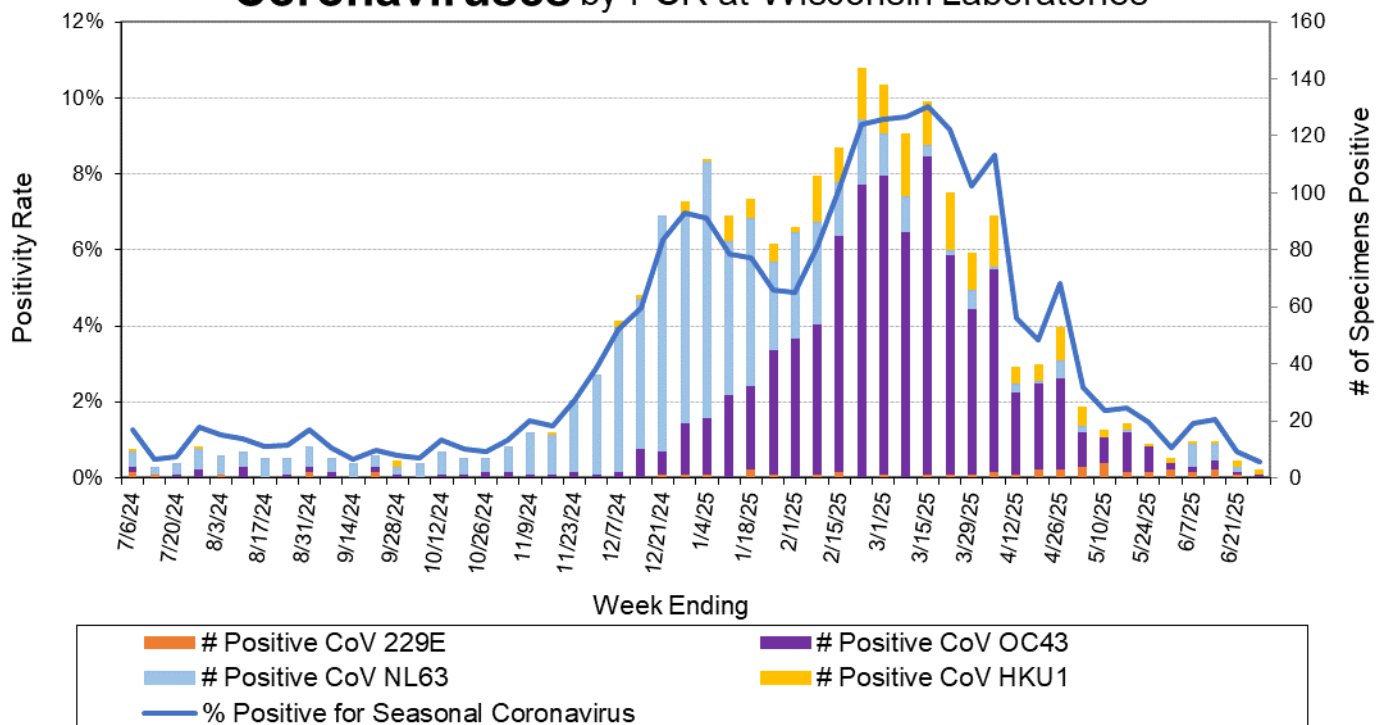


## Surveillance Graphs for 2024-25 Season

Positivity Rate and Number of Specimens Positive for  
**Parainfluenzavirus** by PCR at Wisconsin Laboratories

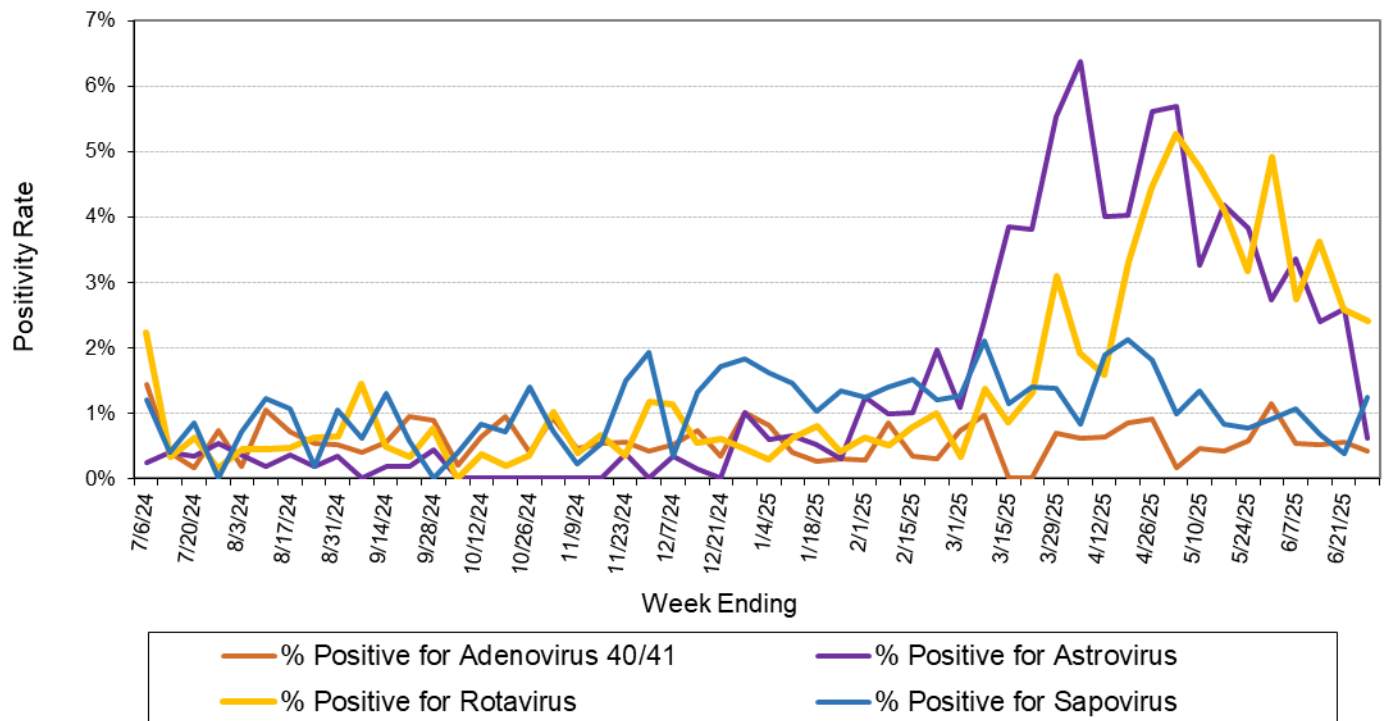


Positivity Rate and Number of Specimens Positive for **Seasonal Coronaviruses** by PCR at Wisconsin Laboratories



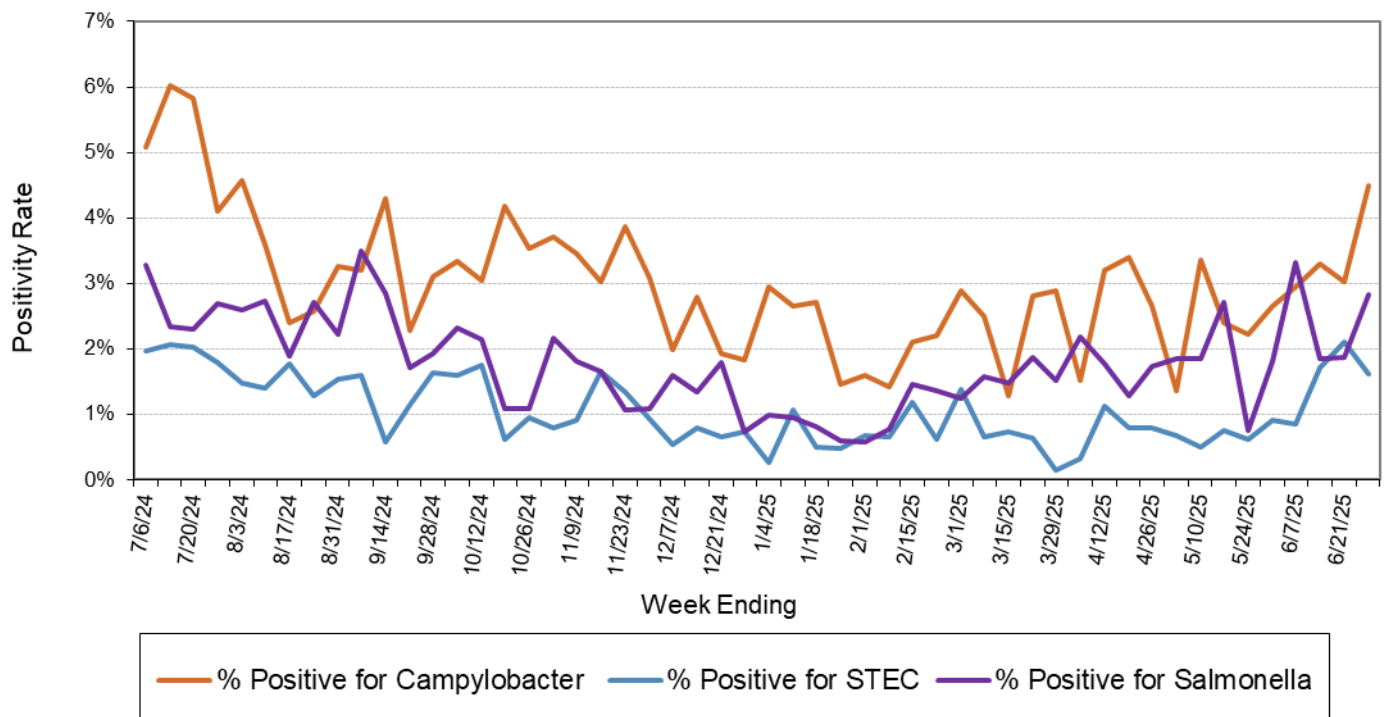
## Surveillance Graphs for 2024-25 Season

### Positivity Rate of **Viral Enteric Pathogens** by PCR at Wisconsin Laboratories



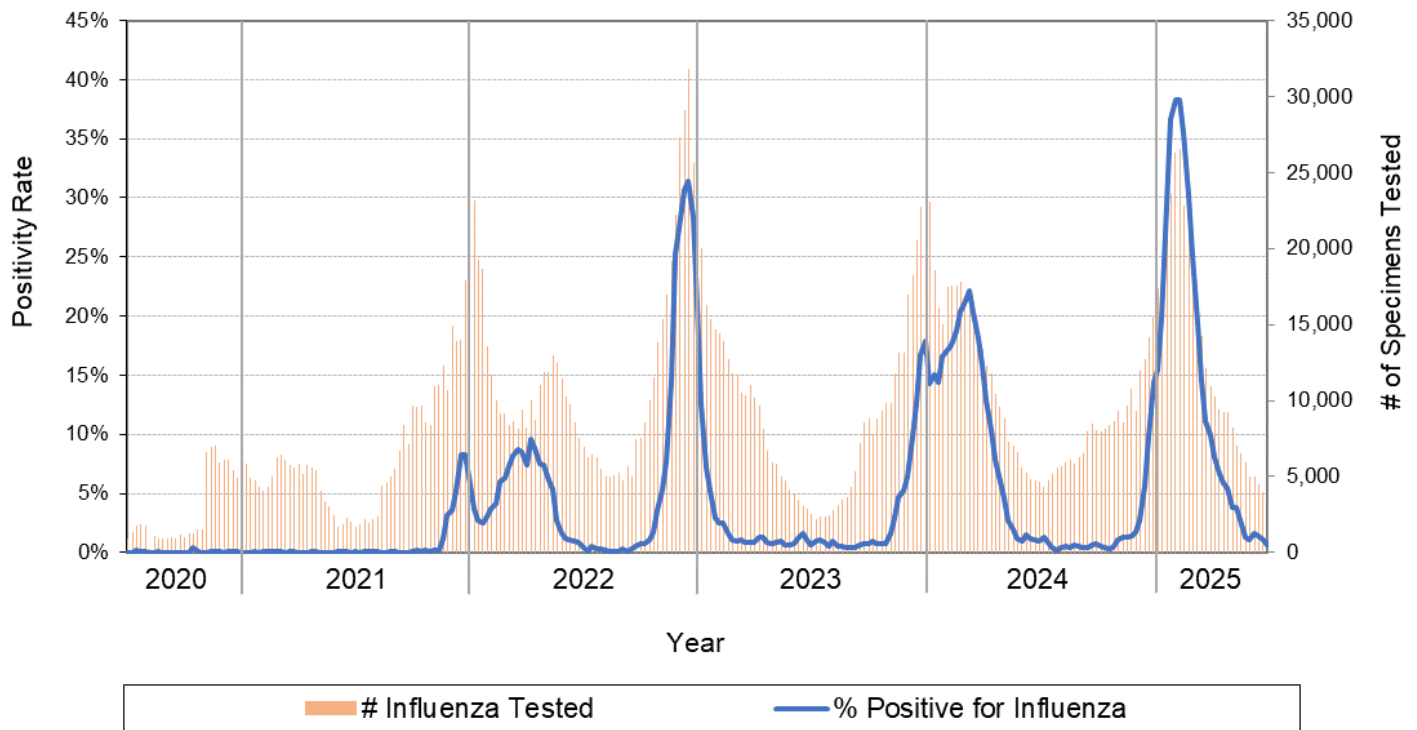
\* Norovirus data is included on page 34.

### Positivity Rate of **Bacterial Enteric Pathogens** by PCR at Wisconsin Laboratories

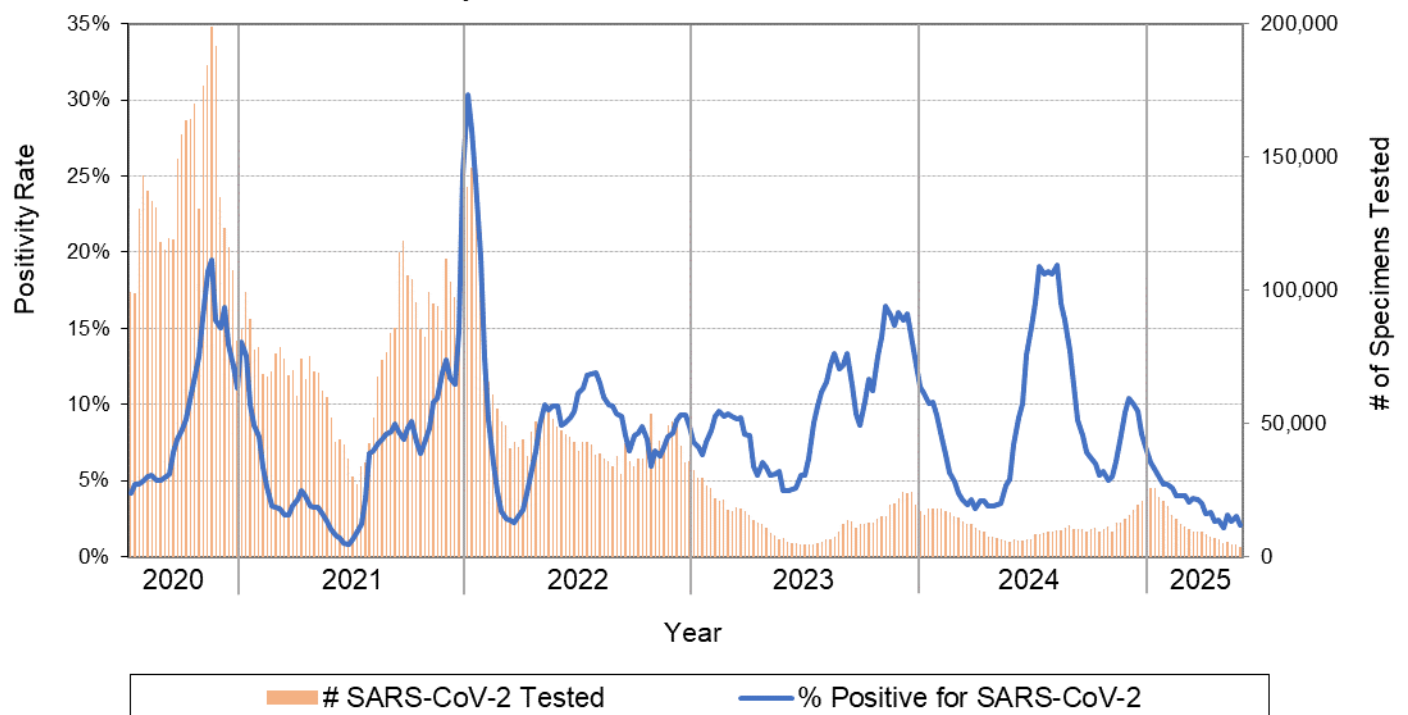


## Surveillance Graphs - 5 Year Summary

Positivity Rate and Number of Specimens Tested for **Influenza** by PCR at Wisconsin Laboratories from 2020-25



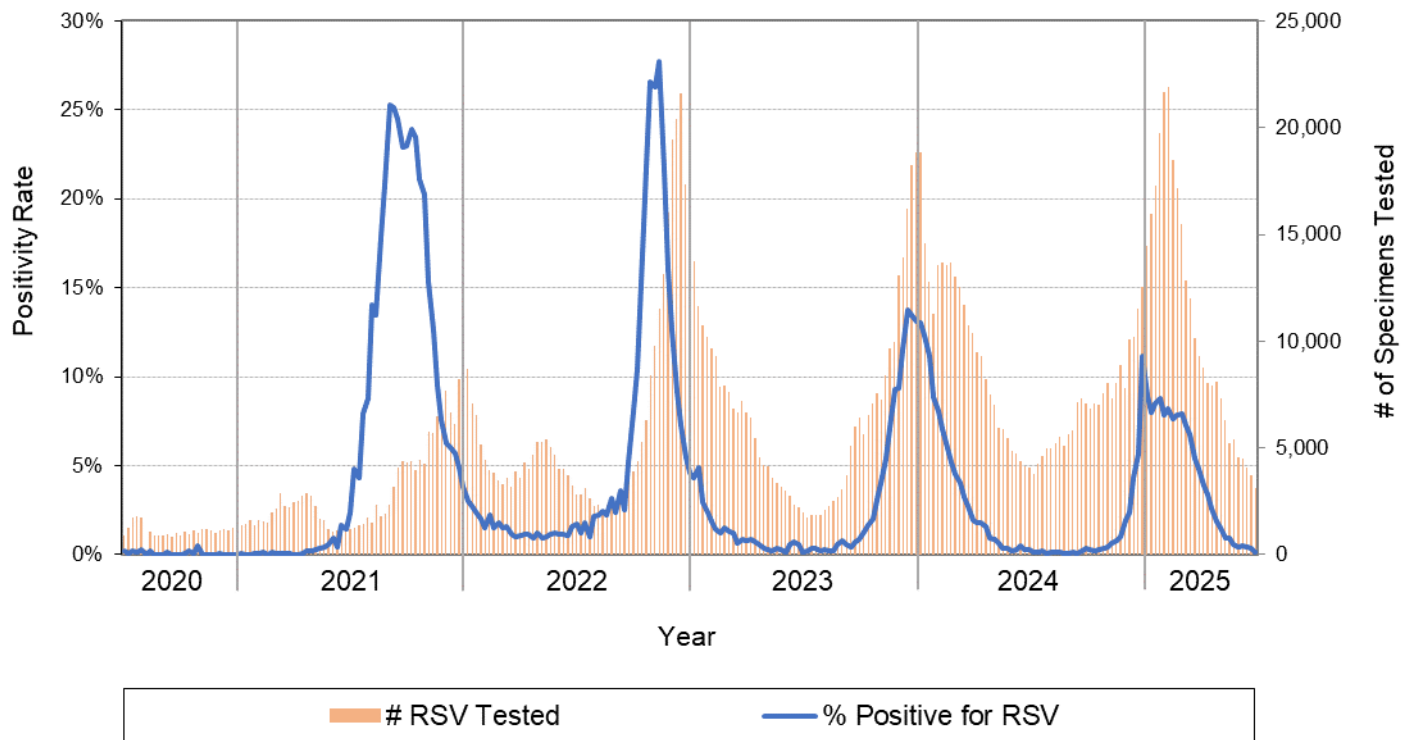
Positivity Rate and Number of Specimens Tested for **SARS-CoV-2** by PCR at Wisconsin Laboratories from 2020-25



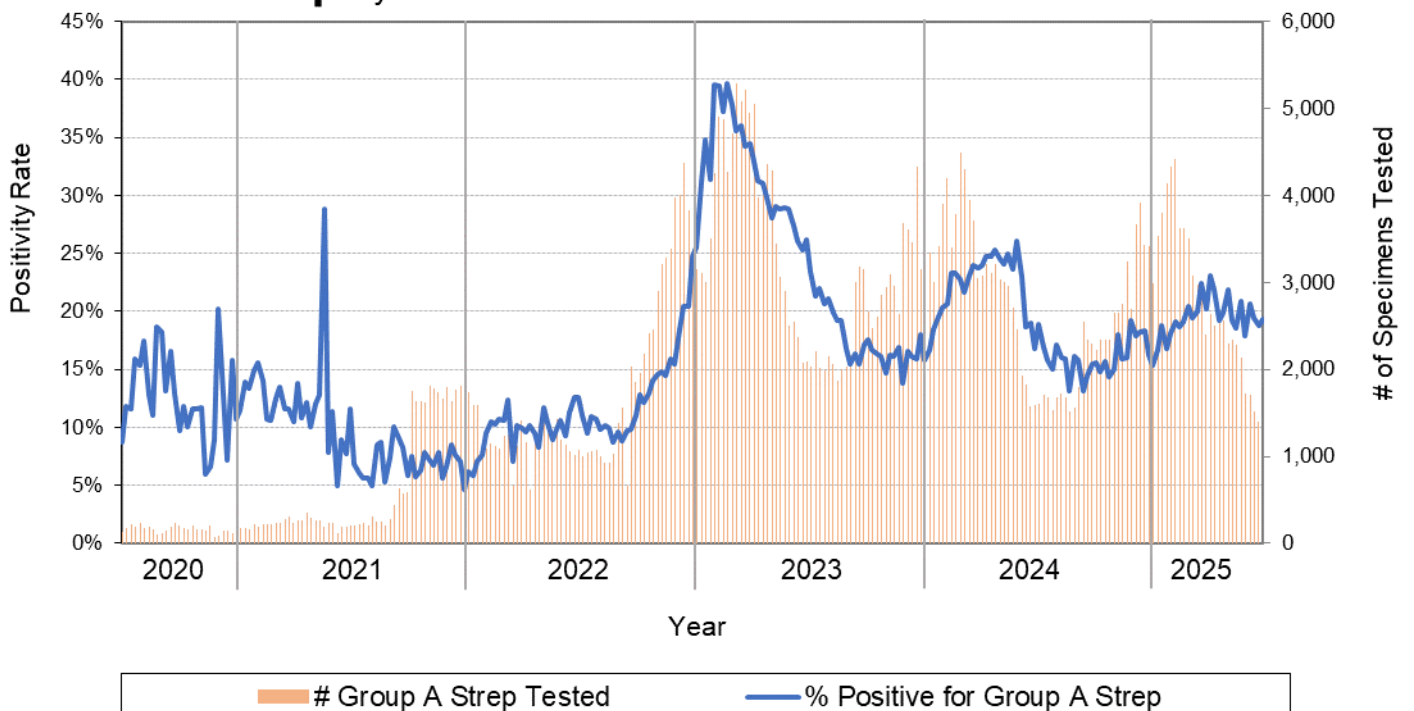


## Surveillance Graphs - 5 Year Summary

Positivity Rate and Number of Specimens Tested for **RSV** by PCR at Wisconsin Laboratories from 2020-25



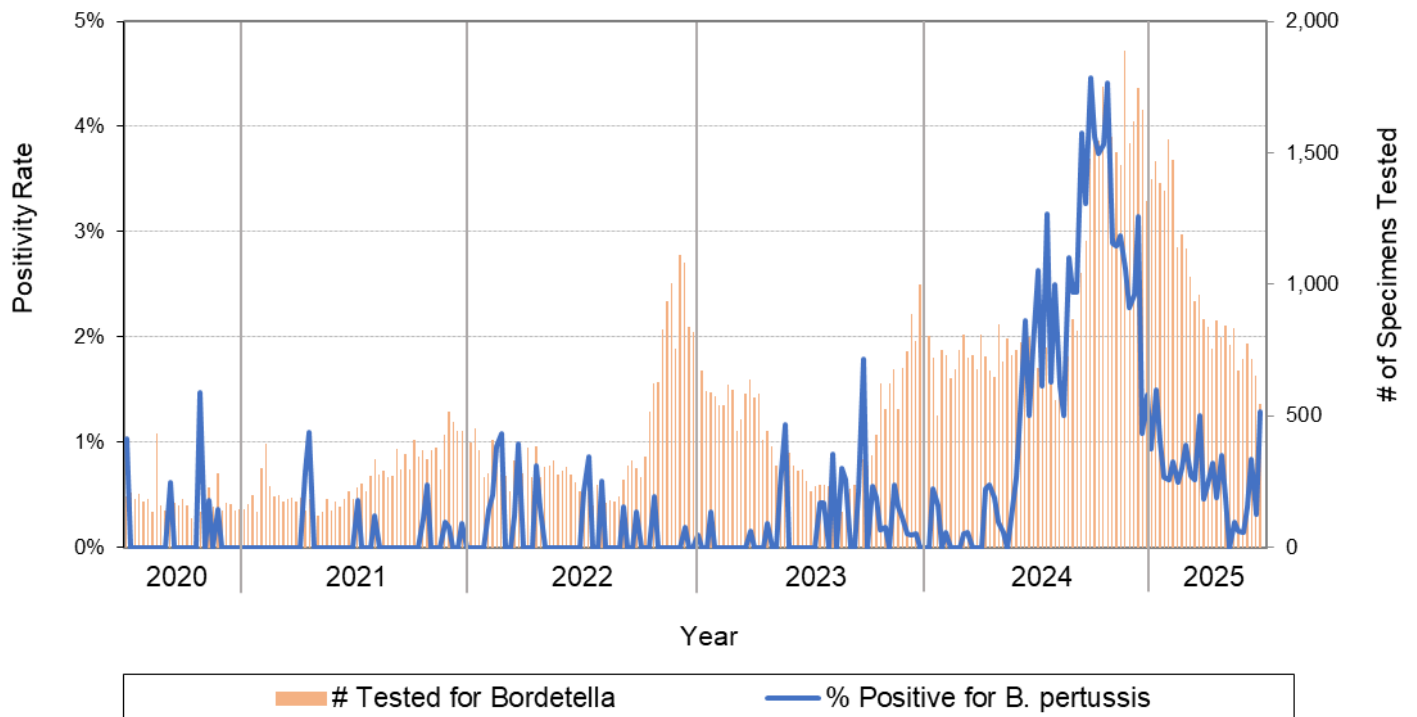
Positivity Rate and Number of Specimens Tested for **Group A Strep** by PCR at Wisconsin Laboratories from 2020-25



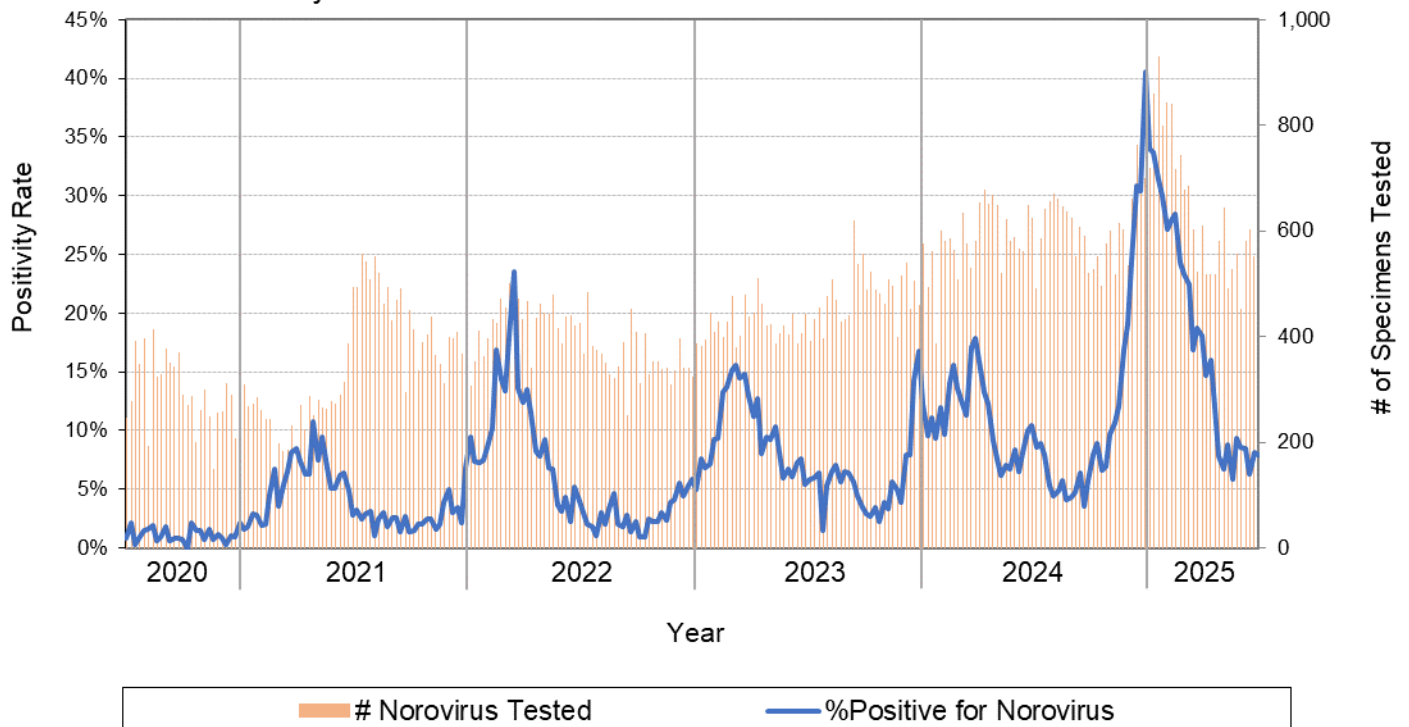


## Surveillance Graphs - 5 Year Summary

Positivity Rate and Number of Specimens Tested for  
***B. pertussis*** by PCR at Wisconsin Laboratories from 2020-25



Positivity Rate and Number of Specimens Tested for **Norovirus**  
by PCR at Wisconsin Laboratories from 2020-25



**\*\*BioMerieux has announced an increased risk of false positive norovirus results with the BioFire FilmArray GI panel, which may increase the statewide norovirus percent positivity\*\***

# AR Lab Network State-wide Surveillance

## **Carbapenem Resistant *Acinetobacter baumannii***

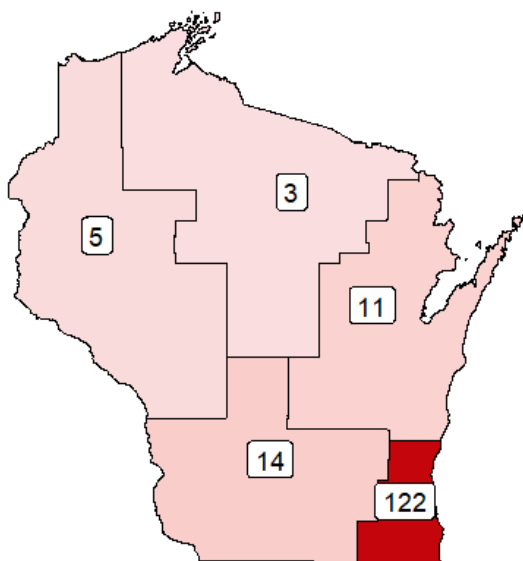
Nearly all carbapenem-resistant *Acinetobacter baumannii* (CRAB) infections occur in patients who recently received care in a healthcare facility. In 2017 CRAB infections attributed to \$281 million in healthcare costs (2019 AR Threats Report; <https://www.cdc.gov/antimicrobial-resistance/media/pdfs/2019-ar-threats-report-508.pdf>). CRAB can survive in the environment for a long time and some CRAB isolates are resistant to nearly all antibiotics available, with few new drugs in development. CRAB can carry carbapenemases, which are encoded on mobile genetic elements and are easily shared between bacteria. 98% of all CRAB isolates submitted from Wisconsin carried a carbapenamase.

*WSLH requests isolates of Acinetobacter baumannii that are resistant to any carbapenems to be submitted for surveillance testing. Testing performed at the WSLH includes identification, antimicrobial susceptibility testing, AR-targeted PCR and molecular subtyping. AR-targeted RT-PCR includes testing for 8 unique carbapenamase genes.*

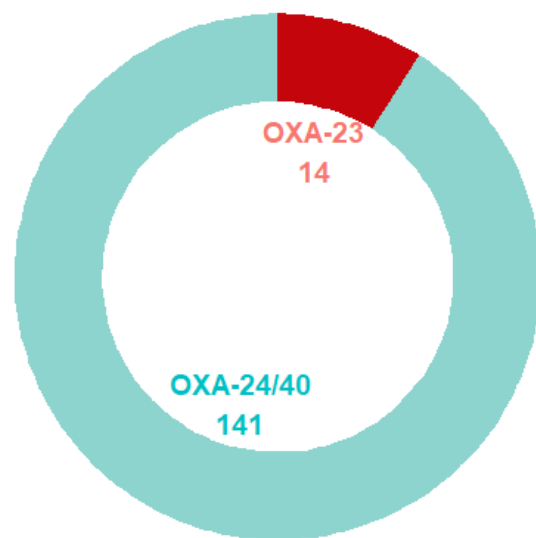
### **2024:**

- WSLH received 158 CRAB isolates from Wisconsin laboratories in 2024.
  - Carbapenemase genes were detected by AR-targeted RT-PCR in 155 isolates (98.1%).
  - 45 CRAB isolates were non-susceptible to all antibiotics tested (28.5%), an 8 percent increase from 2023.

## **Carbapenemases detected at WSLH in clinical CRAB isolates from Wisconsin laboratories in 2024**



Carbapenemase detections in CRAB isolates, by region of the state.



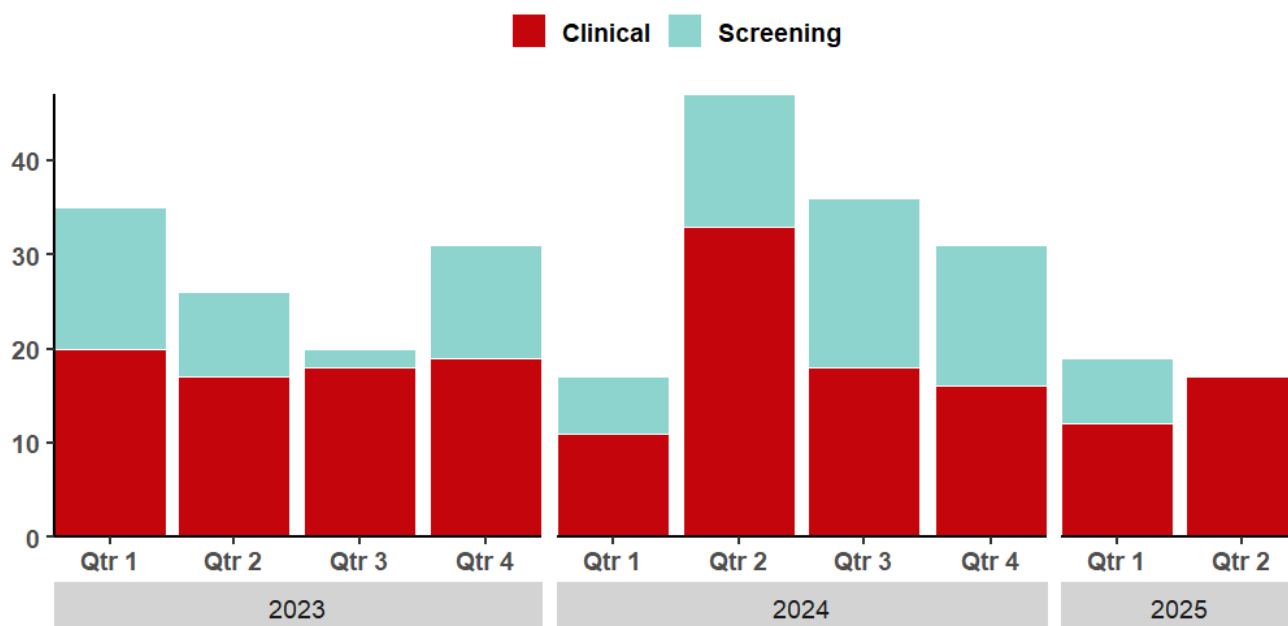
Carbapenemase genes detected in CRAB isolates by AR-targeted RT-PCR.

## AR Lab Network State-wide Surveillance (Cont'd)

### Colonization Testing for CRAB in Wisconsin

- WSLH performed screening on 1717 specimens in 2024 from 15 Wisconsin facilities.
  - CRAB was detected in 74 screening specimens (4.3%).

### CRAB Cases Detected in Wisconsin from Clinical Cases (Infections) and Colonization Screenings



### Carbapenem-Resistant *Pseudomonas aeruginosa* (CRPA)

*Pseudomonas aeruginosa* causes many types of healthcare-associated infections. Some *P. aeruginosa* are resistant to nearly all antibiotics, including carbapenems, and are known as multidrug-resistant (MDR) *P. aeruginosa*. In 2017, MDR *P. aeruginosa* caused an estimated 32,600 infections among hospitalized patients and 2,700 estimated deaths in the United States (2019 AR Threats Report).

WSLH requests *Pseudomonas aeruginosa* isolates that are resistant to carbapenems other than ertapenem AND non-susceptible to cefepime and/or ceftazidime. Testing performed at the WSLH includes identification, antimicrobial susceptibility testing, carbapenemase screening, AR-targeted PCR and molecular subtyping.

#### 2024:

- WSLH received a total of 427 *P. aeruginosa* isolates from Wisconsin Laboratories in 2024.
  - No carbapenemase genes were detected by AR-targeted RT-PCR.
  - 3 CRPA isolates were non-susceptible to all antibiotics tested (0.7%), a decrease of 2 percent.



## AR Lab Network State-wide Surveillance (Cont'd)

### Carbapenem-Resistant *Enterobacterales*

Carbapenem-resistant *Enterobacterales* (CRE) are a major concern for patients in healthcare facilities. In 2017, CRE caused an estimated 13,100 infections in hospitalized patients and about 1,100 deaths in the United States (2019 AR Threats Report). CRE can carry carbapenemases, an antibiotic resistance mechanism which makes carbapenem antibiotics ineffective. Carbapenemases are encoded on mobile genetic elements that are easily shared between bacteria. This increases the risk that antibiotic resistance will spread to other bacteria and/or other patients.

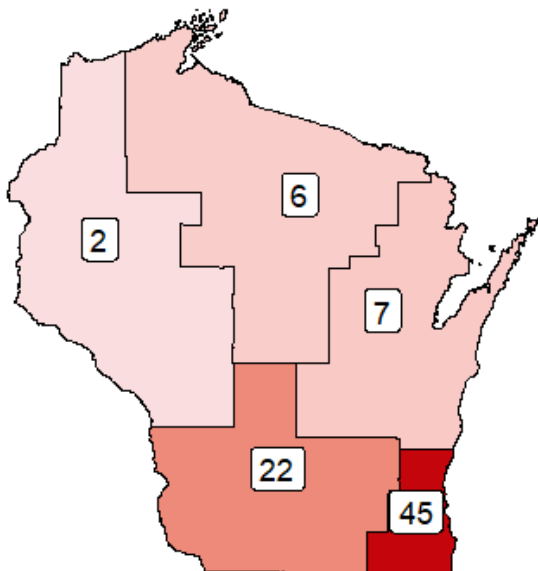


WSLH requests isolates of *Enterobacterales* that are resistant to any carbapenems to be submitted for surveillance testing. Testing performed at the WSLH includes identification, antimicrobial susceptibility testing, carbapenemase screening, AR-targeted PCR and molecular subtyping. AR-targeted RT-PCR includes testing for 5 unique carbapenemase genes.

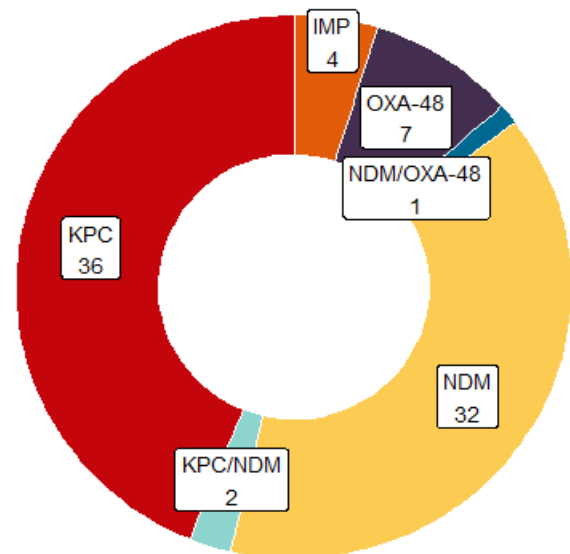
### 2024:

- WSLH received 423 CRE isolates from Wisconsin laboratories in 2024.
- Carbapenemase genes were detected by AR-targeted RT-PCR in 82 isolates (19.4%).
- 4 CRE isolates were non-susceptible to all antibiotics tested (0.9%)

### Carbapenemases detected at WSLH in CRE Isolates from Wisconsin laboratories in 2024



Carbapenemase detections in CRE isolates, by region of the state.



Carbapenemase genes detected in CRE isolates by AR-targeted RT-PCR.

## AR Lab Network State-wide Surveillance (Cont'd)

### **Candida auris**

*Candida auris* is an emerging multi-drug resistant yeast that can cause severe, life-threatening infections. *C. auris* affects ill or immunocompromised patients and is highly transmissible in healthcare settings. Patients can also be colonized with *C. auris* without having symptoms. Colonized patients can easily spread *C. auris* to other patients, and once a patient is colonized with *C. auris*, they are considered colonized for life! Early detection of clinical and colonized cases, followed by measures like colonization screening and infection control, can limit the spread of *C. auris*. The first Wisconsin resident with a *C. auris* infection was identified in 2021.

WSLH requests all *C. auris* isolates for identification, antimicrobial susceptibility testing and molecular subtyping. WSLH also performs *C. auris* colonization screening in response to positive cases or outbreaks at the discretion of the DPH.

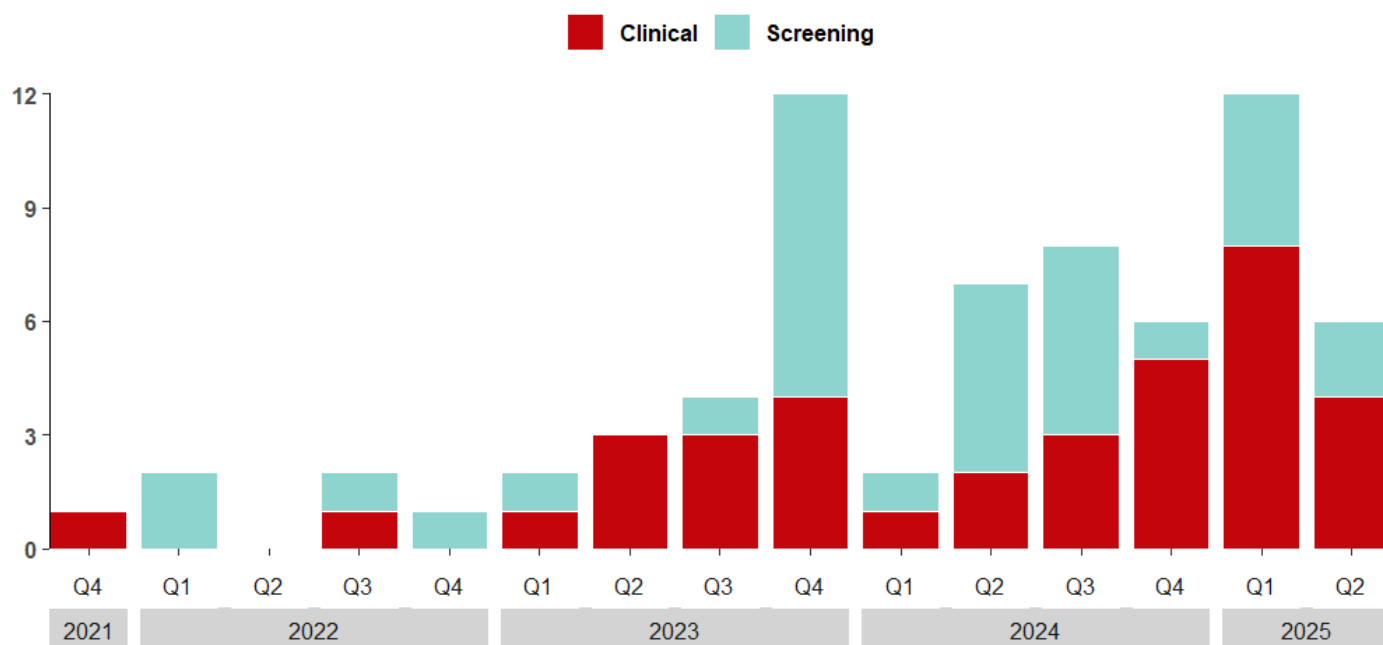
### **2024:**

- WSLH received a total of 18 *C. auris* clinical isolates from Wisconsin Laboratories in 2024.
  - All isolates were susceptible to echinocandins and Amphotericin B
  - 66.7% of *C. auris* isolates from Wisconsin were resistant to azole antifungals.

### **Colonization Testing for *C. auris* in Wisconsin**

- WSLH performed screening on 1547 specimens in 2024 from 15 Wisconsin facilities.
  - *C. auris* was detected in 17 screening specimens (1.1%).

### ***Candida auris* Cases Detected in Wisconsin from Clinical Cases (Infections) and Colonization Screenings**



# Wisconsin Mycobacteriology Data —2024

There were 71 new Report-Verified Cases of Tuberculosis in Wisconsin in 2024. 62 Wisconsin patients had culture-confirmed tuberculosis with susceptibility testing performed‡.

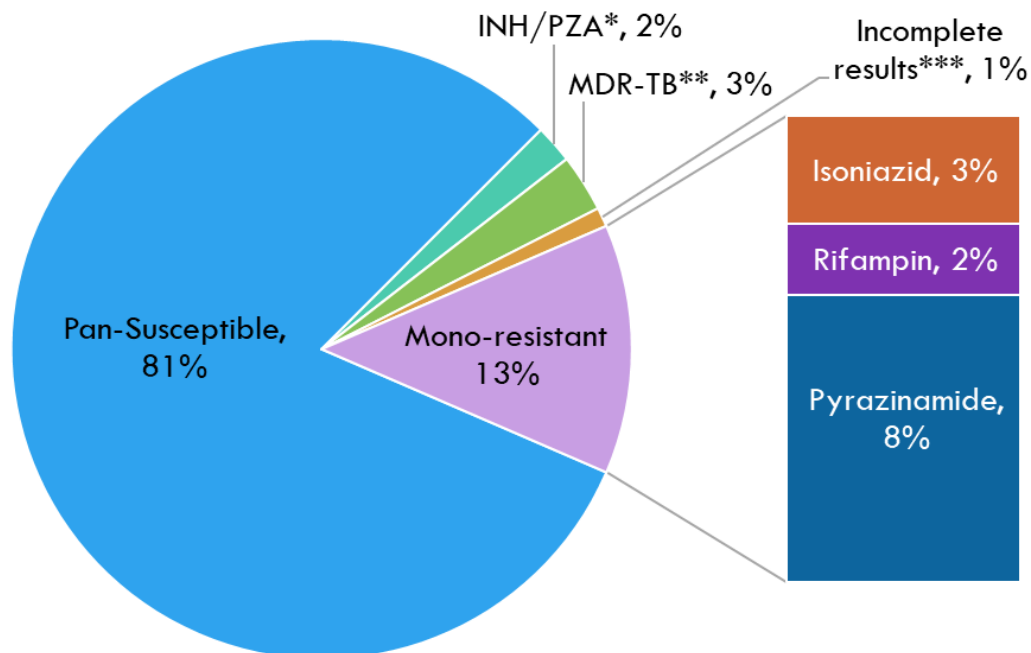
## Wisconsin Patients with New Isolations of *Mycobacterium tuberculosis* complex

County of Residence	Barron	Brown	Dane	Dodge	Iowa	Kenosha	Marathon	Milwaukee	Ozaukee	Pierce	Portage	Price	Rock	Sheboygan	Trempealeau	Waukesha	Winnebago	TOTALS
Pulmonary	1	4	10		1	2	3	14	1	1	1	1	1	2	3	2	1	48
Extra-pulmonary*	1	2	1	1				6								2	1	14
Totals	2	6	11	1	1	2	3	20	1	1	1	1	1	2	3	4	2	62

(‡) 9 cases were clinically diagnosed with no culture growth

(\*)Extra-pulmonary sources of isolation: 3 lymph node, 3 neck, 3 pleural fluid, 2 peritoneal fluid, 1 abscess, 1 gastric aspirate, 1 colon

## 2024 Wisconsin TB Isolate Drug Susceptibility



\*One isolate was resistant to both Isoniazid and Pyrazinamide

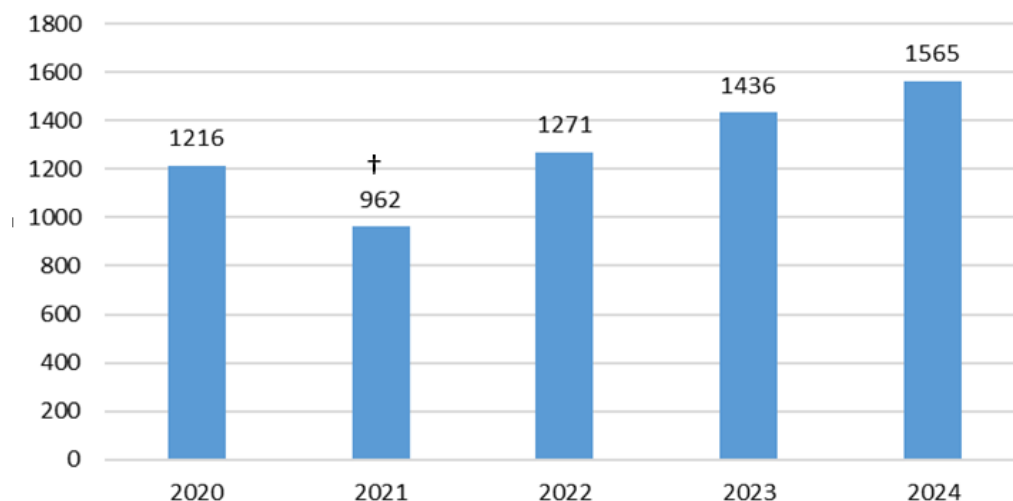
\*\*MDR-TB refers to specimens that are resistant to both Isoniazid and rifampin

\*\*\*One specimen was mixed in culture with a rapidly growing non-tuberculous mycobacteria species. Pure MTBC growth was unable to be isolated for phenotypic susceptibility testing.

## Wisconsin Mycobacteriology Data — 2024

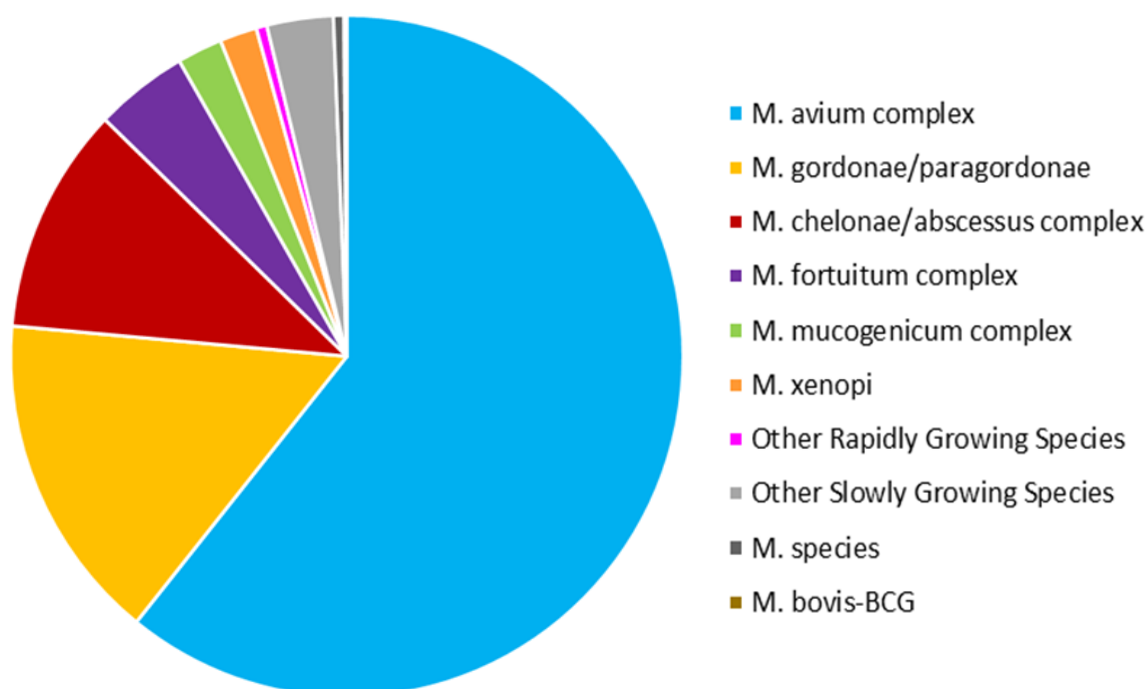
Non-tuberculous mycobacteria (NTM) isolations are self-reported to the Wisconsin State Laboratory of Hygiene (WSLH) by mycobacteriology labs in the state of Wisconsin. These data represent one unique isolation per organism per patient. Subsequent cultures and isolations are not included in this summary. The figures on this page depict NTM isolation trends over the past five years as well as a breakdown of the most common NTM identifications for 2024.

Number of unique NTM isolations in WI by year



†Isolate data for 2021 is incomplete, as one WMLN lab was unable to submit NTM isolation data that year.

WI 2024 NTM Isolation Summary (N=1565)





## 2024 *Legionella* Investigation

### 2024 Legionnaires' Disease Community Cluster Investigation

In September 2024, the Wisconsin Department of Health Services (DHS) collaborated with WSLH and the Department of Agriculture, Trade, and Consumer Protection (DATCP) to investigate a cluster of six Legionnaires' disease cases.

- All six case-patients worked or traveled to the same downtown community within the month of September 2024.
- All case-patients were diagnosed by Legionella urine antigen tests, which detects illnesses caused by Legionella pneumophila serogroup 1.

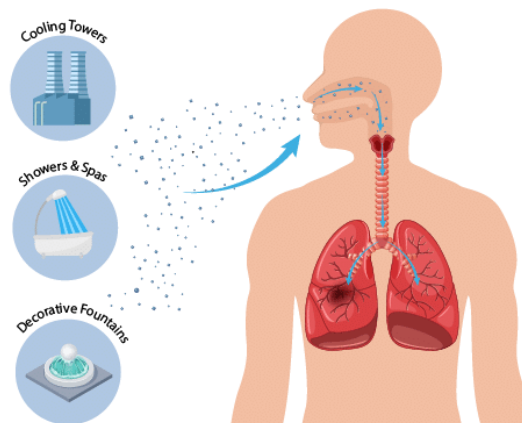


A bronchoalveolar lavage (BAL) specimen was collected from two of the six case-patients. Legionella pneumophila serogroup 1 was isolated by culture from both specimens.

Further investigation revealed that all six case-patients visited locations within 0.5 miles of one another in 14 days prior to illness onset.



- While two patients visited the same lodging facility, there were otherwise no common facility visits between the cases.
- DHS and DATCP collected water samples from the lodging facility's potable water system, as well as cooling towers from six buildings within two city blocks of the lodging facility.
- Samples were analyzed for environmental Legionella PCR and culture.
- Cooling towers from five of the six buildings tested were positive for Legionella; two of the buildings tested positive for Legionella pneumophila serogroup 1.



Source: <https://www.kurita.eu/legionella/>

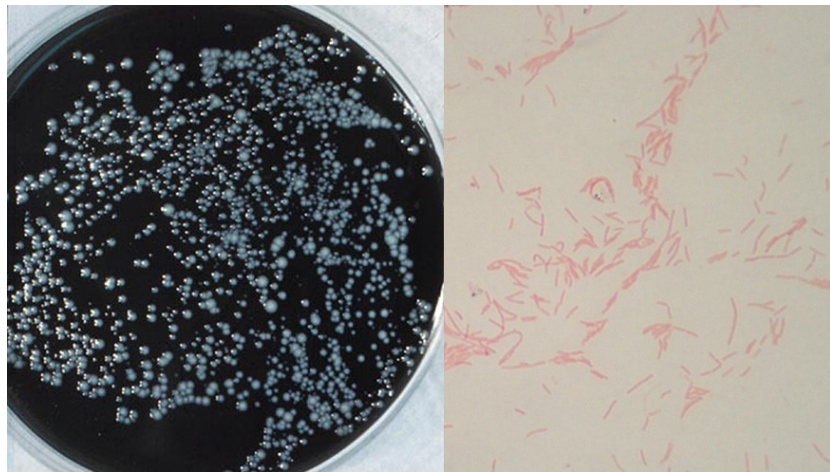
## 2024 Legionella Investigation (Cont'd)

Because both a clinical isolate and two environmental isolates for *Legionella pneumophila* serogroup 1 were stored at WSLH, further characterization was possible. To identify the link between the clinical specimen and a specific cooling tower, sequence-based typing was performed on all three isolates.

- The clinical isolate and one of the cooling towers both resulted with the same, uncommon sequence type.
- The other cooling tower resulted with a different, more common sequence type.
- All cooling towers that tested positive for *Legionella* through public health investigation were either remediated by online disinfection or were shut down for the season.



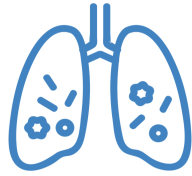
Laboratorians play a pivotal role in community cluster investigations of Legionnaires' disease. Since 2023, 92% of all Legionnaires' disease cases in Wisconsin have been diagnosed by a urine antigen test. However, a urine antigen test result cannot be directly compared to environmental samples. *Legionella* isolates from a sputum or BAL specimen can be directly compared, with sequence-based typing or whole genome sequencing, to clinical isolates from epidemiologically-linked patients and environmental isolates, aiding in the investigation of suspect clusters of community-acquired Legionnaires' disease.



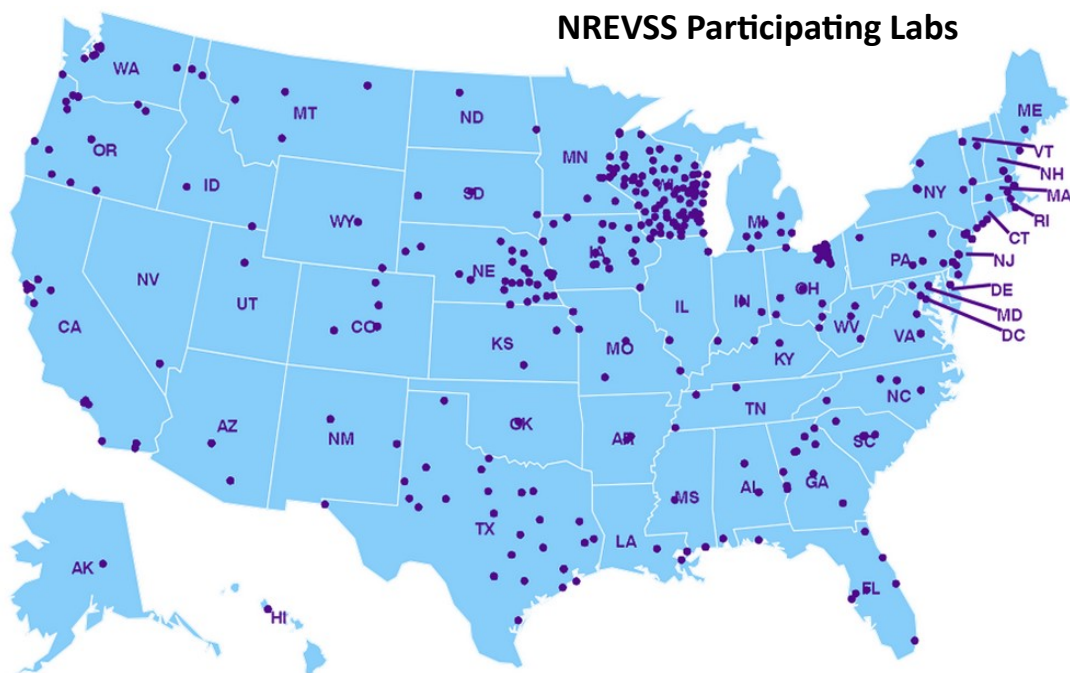
<https://asm.org/articles/2021/march/cooling-tower-of-terror-legionella-s-public-health>

In an effort to obtain more clinical isolates, fee-exempt *Legionella* culture is available at WSLH for patients that have tested positive using a *Legionella* urine antigen or PCR test. If *Legionella* is isolated from a clinical specimen, the isolate should be sent to WSLH for storage. Suspect community clusters of Legionnaires' disease can occur over an extended period of time.





**THANK YOU FOR YOUR  
PARTICIPATION IN  
LABORATORY-BASED SURVEILLANCE!!**





# **Wisconsin State Laboratory of Hygiene**

UNIVERSITY OF WISCONSIN-MADISON

**Communicable Disease Division  
2601 Agriculture Drive  
Madison, WI 53718  
Customer Service: 800-862-1013**



***WWW.SLH.WISC.EDU***